Gigantism

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ABSTRACT

Gigantism is a non-specific term that denotes excessive growth in a pediatric patient. This may rarely result from an over production of growth hormone (GH), which is often termed *pituitary gigantism*, or it may arise from an overgrowth syndrome. Pituitary gigantism can present as early as during infancy or not until adolescence and may be congenital or acquired. Likewise, it may occur as a sporadic condition or in the context of a well described syndrome in which hypersecretion of GH is a potential feature. Conditions in which GH excess occurs include Neurofibromatosis Type 1, McCune-Albright syndrome, Multiple Endocrine Neoplasia Type 1, Carney Complex, Isolated Familial Somatotropinomas and X-Linked Acrogigantism. Therapeutic modalities for the treatment of pituitary gigantism are the same as those for acromegaly and include surgery, medication and radiation. The second major category of gigantism is that which is due to the presence of an overgrowth syndrome. Some of the most notorious overgrowth syndromes are Sotos syndrome, Beckwith-Wiedemann syndrome, Simpson-Golabi-Behmel syndrome and Weaver syndrome. Great strides have been made in identification of the molecular genetic basis for both pituitary gigantism and overgrowth syndromes, affording novel insights into the mechanisms underlying normal and abnormal growth. Etiologies, phenotypic features, and diagnostic and treatment considerations pertaining to the most common forms of gigantism are reviewed. For complete coverage of all related areas of Endocrinology, please visit our on-line FREE web-text, <u>WWW.ENDOTEXT.ORG</u>.

INTRODUCTION

Gigantism refers to a condition characterized by extreme physical size and stature. By definition, this originates during infancy, childhood or adolescence, while epiphyseal growth plates remain open. Although often used to specifically denote growth hormone excess, the term gigantism is also applied to a number of non-hormonally mediated overgrowth conditions in children (1). All forms of gigantism are extremely rare and have in common a complex pathophysiologic origin and extraordinary clinical manifestations. Although many aspects of overgrowth disorders remain to be elucidated, insights into the molecular genetic basis of several forms of gigantism have contributed greatly to our understanding of specific mediators of growth and cellular function.

GROWTH HORMONE EXCESS

The association between gigantism and growth hormone (GH) excess was recognized as early as the 1800's, when it was noted that pituitary giants invariably developed features of acromegaly, which refers to progressive enlargement of the head, face, hands and feet (2). The major difference between these two conditions is that gigantism results from excessive GH production during the period of active growth and acromegaly results from GH excess ensuing after epiphyseal fusion has occurred. A further distinction relates to the overall incidence of these disorders. While acromegaly is

uncommon, occurring at an estimated worldwide annual rate of 2.8-4 cases per million (3), GH excess during childhood and adolescence is extremely rare, with an estimated incidence of 8 per million person-years and the total number of reported cases thus far numbering only in the hundreds. Despite these disparities, some degree of clinical overlap has been suggested by the observation that 10% of acromegalics have tall stature(4), indicating that the onset of GH excess pre-dates epiphyseal fusion in many patients. GH hypersecretion may occur sporadically or may exist within a constellation of abnormalities in the setting of several well-recognized syndromes. Conversely, a genetic predilection to only the development of GH-secreting pituitary adenomas may be present, as is the case in kindreds with isolated familial somatotropinomas. In recent years there has been increased recognition of the underlying molecular genetic abnormalities that lead to pituitary gigantism, one of which can be identified in approximately 50% of cases (5).Regardless of the underlying etiology, the clinical manifestations of GH hypersecretion in childhood are indistinguishable and the initial diagnostic evaluation is standardized. The various categories and sources of GH excess along with their associated genetic abnormalities, if known, are summarized in Table I and will be discussed individually.

Sporadic	1	Syndromic/Familial		
Disorder	Source of GH	Disorder	Source or Genetic Mutation	
Hypothalamic/Pituitary GH excess	Congenital GHRH excess (postulated)	Neurofibromatosi s-1	Tumor infiltration into somatostatinergic	
	Pituitary somatotroph or mammosomatotroph adenoma	McCune- Albright syndrome	Activating mutation of $Gs\alpha$	
	Pituitary hyperplasia	Multiple endocrine neoplasia Type-1 and Multiple endocrine	Defect in tumor suppression from mutations in menin and CDKN1 genes	
	Hypothalamic gangliocytoma/neurocytoma	Carney complex	 Abnormality at 2p16 Mutations in PRKAR1A at 17q22-24 	
		3PA association	Succinate dehydrogenase defects	

Table 1: Etiologies of Growth Hormone Excess

Ectopic GH excess	GHRH or GH production by bronchial, carcinoid or pancreatic neoplasm Ectopic pituitary adenoma	 Familial somatotrophino mas X-LAG syndrome 	 Mutation in acyl hydrocarbon receptor geneat 11q13.3 Abnormality at 2p12-6 Xq26.3 duplications
GH-growth hormone, G regulatory subunit 1	HRH-growth hormone-releasing ho	rmone, PRKARIA-p	rotein kinase A

SPORADIC FORMS OF GROWTH HORMONE EXCESS

Sporadic GH excess may arise from CNS pathology or, rarely, from ectopic GH production. Traditionally, the term "primary growth hormone excess" has been used to differentiate an intrinsic pituitary source of GH from other causes, including hypothalamic abnormalities. In actuality, it may be difficult to clearly distinguish the role of the pituitary from the hypothalamus, particularly in cases of early childhood GH excess, as discussed below.

Hypothalamic-Pituitary Growth Hormone Excess

Unlike in acromegalic adults, in whom discreet pituitary adenomas are present in the overwhelming majority of patients (6), a number of different histopathologic mechanisms underlying childhood GH hypersecretion have been suggested or observed. These relate to the concept that childhood GH excess represents a distinct entity, with different characteristics in terms of pituitary morphology and function. Supporting this view have been reports of diffuse pituitary hyperplasia in the setting of early-onset gigantism, in which congenital growth hormone releasing-hormone (GHRH) excess has been proposed as the inciting cause (7;8). Additionally, the nearly ubiquitous finding of combined GH and prolactin over-secretion in nearly all cases of early childhood gigantism, a feature not universally present in acromegaly, suggests that a separate pathologic process may be involved. This dual hormonal secretion has been attributed to the presence of mammosomatotrophs (9;10), which are rare in adulthood but predominate in fetal life. Even in cases of apparent pituitary microadenomas or macroadenomas arising during early childhood, this unique biochemical feature has been present (11;12). In contrast, prolactin levels are usually normal in cases of pituitary GH-secreting adenomas originating during adolescence, which may be thought of as existing within the spectrum of adult GH hypersecretion. Interestingly, a reversible transformation of pituitary somatotrophs into bihormonal mammosomatotrophs under the influence of ectopic overproduction of GHRH has been observed, lending additional support to the concept that hypothalamic GHRH excess may play a pivotal role in the genesis of early-onset gigantism (13). Although the etiology of sporadic gigantism is often unknown, a number of germline and somatic mutations in genes associated with syndromic and familial GH hypersecretion have been reported in children and adolescents with pituitary gigantism (13;14).

An additional cause of sporadic GH excess linked to CNS pathology is that which occurs in the setting of a hypothalamic gangliocytoma or neurocytoma. These rare tumors, comprised of large hypothalamic-like ganglion cells, have been demonstrated to produce GHRH (15;16) and to be found in close proximity to pituitary growth hormone-secreting adenomas (17). Normalization of serum growth hormone levels following resection of the hypothalamic tumor in some patients has

further supported a central role for abnormal GHRH secretion in the development of gigantism or acromegaly in these cases (18).

Ectopic Growth Hormone Excess

Ectopic GH hypersecretion is a rare but important cause of acromegaly in adults, thought to represent less than 1% of all cases (19;20). In this condition, a paraneoplastic elaboration of GHRH or uncommonly GH (21) occurs, with neuroendocrine tumors being the most common source (22). Specific lesions notorious for this capability include bronchial carcinoid and pancreatic neoplasms (23;24). Extra pituitary GH excess has also been reported in the setting of an ectopic pituitary adenoma located within the sphenoid sinus or clivus (25;26), and in association with an empty sella (27). To our knowledge, an ectopic source of GHRH or GH leading to gigantism in a child has never been described.

SYNDROMIC AND FAMILIAL FORMS OF GROWTH HORMONE EXCESS

A second major category of childhood GH hypersecretion is that which occurs in the setting of a well-recognized syndrome. In these cases, gigantism may be the sole presenting feature of the syndrome or it may be detected during on-going clinical follow-up for other endocrine or nonendocrine problems. Alternatively, biochemical evidence of sub-clinical GH excess may be revealed through routine surveillance in a child known to be at risk for the development of gigantism. As is the case in sporadic GH hypersecretion, a variety of different morphologic abnormalities involving the pituitary gland may be found. Paracrine pituitary GHRH secretion has also been implicated by the discovery of GHRH expression from clusters of cells in the hyperplastic pituitaries of two boys from a family with hereditary early-onset gigantism (28). Syndromes that are associated with the development of childhood GH excess are reviewed below. Table 2 outlines the characteristics of the GH excess and other clinical features in these disorders.

Disorder	Mode of Inheritance	Clinical Features	Frequenc of Gigantism	Age of Onset of gigantism	Pituitary Morphology	Screening recommendations
Neurofibromatosi s -1	Autosomal Dominant or Sporadic	 Optic gliomas Café au lait skin pigmentation 	Extremely rare	6 months on	Optic pathway tumor with normal to small pituitary	Not routine
McCune- Albright Syndrome	Sporadic	 Precocious Puberty Café au lait skin pigmentation Fibrous bone dysplasia Multiple endocrinop athies 	15-20%	Early childhood on	Pituitary adenomas or diffuse pituitary hyperplasi a or no visible abnormalit y	Annually

Table 2: Clinical Characteristics in Syndromes Associated with Growth Hormone Excess

Multiple Endocrine Neoplasia Type 1	Autosomal Dominant or Sporadic	Pituitary, pancreatic and parathyroid adenomas	10-60%	10% by age 40 but has occurred as early as age 5	Pituitary adenoma	Annually beginning at age 5
Multiple Endocrine Neoplasia Type 4	Autosomal Dominant or Sporadic	Pituitary, pancreatic and parathyroid adenomas	Unknown	Unknown	Pituitary adenoma	Not established
Carney Complex	Autosomal Dominant or Sporadic	 Multiple endocrine tumors Skin lentigines Cardiac myxomas Neural 	10%	Usually 3 [∞] & 4 [™] decade	Pituitary adenoma or pituitary hyperplasi a	Annually beginning post pubertally
3PA Association	Autosomal Dominant or Sporadic	Pheochromocy tom paraganglioma , pituitary adenoma	Unknown	Usually 3 [∞] & 4 [⊪] decade	Pituitary adenoma with intracytoplas mic vacuoles	As clinically indicated in unaffected family members
Isolated Familial Somatotropinom as	Autosomal Dominant or Sporadic	Isolated GH- secreting pituitary adenomas	100%	Before 3 rd decade and as early as age 5	Pituitary adenoma	As clinically indicated in unaffected family members
X-linked Acrogigantism	Sporadic or X- linked	Isolated GH excess	100%	Early childhood with onset in late infancy or onset during adolescen ce	Pituitary adenoma or pituitary hyperplasi a or both	As clinically indicated in unaffected family members

Neurofibromatosis-1 (NF-1)

Beginning in the 1970's, several reports of gigantism occurring in young children with NF-1 have appeared in the medical literature (29). In these cases, excessive somatic growth has been noted as early as 6 months of life (30). Neuroimaging in these patients typically reveals an optic glioma (31), usually with infiltration into the medial temporal lobe. However, growth hormone excess has frequently been reported to be a transient phenomenon in children with NF-1, raising questions as to the necessity of treatment (32;33). A number of investigations aimed at identifying the precise etiology of the gigantism in these children have been conducted. In all cases in which tumor tissue has been available, immunostaining for GH, GHRH and somatostatin has been uniformly negative (34;35).

This, in conjunction with the known temporal lobe location of somatostatin-producing neurons, led to the hypothesis that GH excess in these patients was the result of a hypothalamic regulatory defect. Specifically, tumor infiltration of somatostatinergic pathways would presumably result in reduced somatostatin tone leading to overproduction of GHRH-mediated pituitary GH. Despite this plausible explanation, arginine-induced GH stimulation in a patient with gigantism in the setting of NF-1 was normal, contrary to the expected lack of response to arginine, which is believed to act through somatostatin inhibition (36). Thus, the precise pathogenesis of gigantism in NF-1 remains unclear. Little information is available regarding the overall incidence of GH hypersecretion in patients with NF-1 and optic gliomas, although studies have suggested that it may occur in over 10% of affected patients, some of whom have concurrent central precocious puberty (37). Interestingly, growth hormone excess has also been reported in children with sporadic optic pathway tumors without associated NF-1 (38) Figure 1 demonstrates the café-au-lait pigmentation and linear growth acceleration observed in a young boy with NF-1 and gigantism.

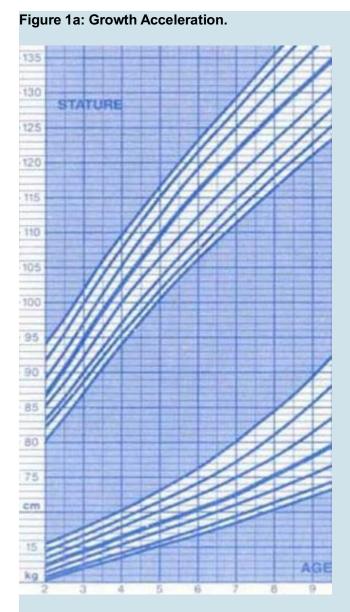


Figure 1b: Characteristic "coast of California" café au lait macules in a child with neurofibromatosis and gigantism.



McCune-Albright syndrome (MAS)

MAS is a complex and heterogenous disorder in which GH excess may arise in conjunction with additional endocrinopathies and other abnormalities. In the classic form, MAS results in the triad of precocious puberty, café-au-lait skin pigmentation, and fibrous dysplasia of bone. It has long been recognized, however, that individuals with MAS have a propensity to develop a number of endocrine problems, including gigantism or acromegaly from excessive growth hormone secretion (39).

Elucidation of the molecular genetic defect in MAS in the early 1990's (40) illuminated the underlying mechanism through which abnormal hormone secretion occurs in this condition. Activating mutations of Gsa, the stimulatory subunit of the heterotrimeric G-protein complex involved in intracellular signaling, have now been shown to form the basis for nearly all of the clinical manifestations of MAS(41). These mutations, which typically involve substitution of arginine at the 201 position with cysteine or histidine, result in unregulated signal transduction leading to increased intracellular cAMP accumulation and downstream gene transcription. The precise timing of the mutation during embryologic life, which occurs in a post-zygotic cell line, will ultimately determine the extent of abnormal cells and severity of the resultant clinical phenotype. The incidence of GH excess in classic MAS has generally been reported to be 15-21% (42;43) and may be more common in males (Yao Y 2017). However, enhanced recognition of the frequency of atypical or forme fruste variants of MAS have the potential to result in an increase of this estimated frequency. Indeed, a number of historical reports of extreme gigantism where fibrous bone dysplasia was also present strongly suggest a diagnosis of MAS in these individuals, a

hypothesis confirmed by molecular genetic analysis in at least one case (44;45). Subclinical growth hormone excess has also been reported in MAS, in which the only clinical manifestation may be the presence of normal stature (rather than short stature) in the context of a history of untreated precocious puberty. Additional phenotypic features in this subgroup of patients with MAS include a higher incidence of vision and hearing deficits, TRH responsiveness, and hyperprolactinemia (46). Growth hormone excess in MAS is typically accompanied by skull base fibrous dysplasia and is notorious for being associated with increased craniofacial morbidity and macrocephaly (47). However, early diagnosis and treatment has been found to decrease the risk of optic neuropathy in these patients (48). A variety of pituitary morphologic abnormalities have been noted in MAS patients with GH hypersecretion (49), ranging from discrete pituitary adenomas (50;51) to diffuse pituitary hyperplasia (7), to no discernible radiographic abnormality (52). Of note is the fact that the identical Gsa mutation found in MAS has also been implicated in the pathogenesis of sporadic GH-secreting pituitary adenomas, where it results in formation of the gsp oncogene. Up to 40% of somatotroph adenomas in adults have been demonstrated to contain either an Arg201 activating mutation, or a related point substitution of glutamine at position 227(53). Interestingly, these sporadic tumors as well as those from patients with MAS and acromegaly display the Gsa mutation exclusively from the maternal allele, providing evidence that the GNAS1 gene is subject to imprinting(54). Figure 2 demonstrates an area of classic café au lait skin pigmentation in a patient with MAS.

Figure 2: Café au lait pigmentation in the typical "coast of Maine" configuration in an individual with McCune-Albright syndrome.



Multiple Endocrine Neoplasia-Type I (MEN-1)

MEN-1 is one of a number of familial cancer syndromes characterized by autosomal dominant inheritance and multi-endocrine gland involvement. Although significant clinical heterogeneity

exists in terms of specific tumor combinations, the most frequent manifestations of MEN-1 are parathyroid, pancreatic, and pituitary adenomas (55). The gene for MEN-1, which had previously been mapped to chromosomal locus 11g13, has been cloned and demonstrated to encode for a 610 amino acid nuclear protein designated menin (56). Many different molecular genetic abnormalities within the menin gene have been identified in kindreds with MEN-1, including nonsense, missense, deletion, insertion and donor-splice mutations(57). Unfortunately, genotype/phenotype correlations have not been observed. In all cases of MEN-1, the development of neoplasia is thought to arise from a defect in normal tumor suppression via a 2-hit hypothesis. The first hit represents inheritance of a germline MEN-1 mutation, leading to a heterozygous loss of the menin gene in every cell (58). As menin is believed to function as a tumor suppressor protein, the second hit involves a somatic MEN-1 mutation in one cell, with subsequent abnormal cellular transformation and clonal expansion. Indeed, somatic biallelic MEN-1 mutations have been demonstrated to be present in at least 15% of sporadic pituitary adenomas, including somatotroph tumors (59). Anterior pituitary adenomas in individuals with known MEN-1 have a reported prevalence of 10-60% and are thought to represent the first clinical manifestation of the disease in up to 25% of sporadic cases (60). Of these, the majority are prolactinomas, with GHsecreting adenomas developing in approximately 10% of individuals with MEN-1 by age 40. The youngest reported case of gigantism in MEN-1 occurred in a 5-year-old boy, who presented with growth acceleration and a GH-secreting mammosomatotroph pituitary adenoma in the context of a known family history of MEN-1 (61). Molecular genetic analysis confirmed the germline and tumor tissue MEN-1 mutations but failed to reveal an etiology for the accelerated presentation in this case. Nonetheless, current recommendations include screening for anterior pituitary hormone excess beginning at age 5 in all individuals with MEN-1, as well as ascertaining MEN-1 carrier status by germline mutation testing in a number of clinical situations (62). Interestingly, GH excess due to ectopic elaboration of GHRH from a pancreatic neuroendocrine tumor has also been reported in several individuals with MEN-1 (63).

Multiple Endocrine Neoplasia-Type 4 (MEN-4)

MEN-4 is a recently recognized entity that is caused by germline mutations in the CDKN1B gene which encodes the putative tumor suppressor p27Kip1 (64). Affected patients are typically heterozygous for mutations in CDKN1B and exhibit a phenotype similar to that seen in MEN-1. Because the number of individuals who have been diagnosed with MEN-4 is low, screening protocols for patients and their family members have not yet been established (65).

Carney Complex (CNC)

Initially described in 1985 (66), CNC is a rare autosomal dominant disorder in which the cardinal features include multiple endocrine tumors, skin lentigines (spotty pigmentation), cardiac myxomas and neural sheath tumors. The condition shares characteristics with several other syndromes, including MEN-1 (multiple endocrine tumors), MAS (endocrine hyperfunction and skin pigmentation) and Peutz-Jeghers (mucosal lentiginoses and gonadal tumors). It has now been demonstrated, however, to have a unique clinical and molecular genetic identity. Two distinct genetic abnormalities have been implicated in the pathogenesis of CNC. The first consists of a locus on 2p16(67), although a specific candidate gene within this region has not been identified. Additionally, mutations in the gene encoding for the protein kinase A regulatory subunit (1 α) (PRKAR1A) at 17q22-24 have been demonstrated in 35-44% of both familial and sporadic cases of CNC(68). This protein, which is intricately involved in endocrine cell signaling pathways, is

thought to function as a tumor suppressor gene. Supporting this theory has been the observation that tumors from patients with CNC (in which diminished levels of PRKAR1A are present) exhibit a 2-fold increase in cAMP responsiveness compared with control tumors (69). The identical mutation has also been found in some sporadic endocrine tumors. As with MEN-1, a germline mutation is thought to be the inciting event for eventual development of the disease. The clinical presentation of CNC is extremely heterogeneous, as is the age at diagnosis. The development of GH excess is rare, occurring usually during the 3rd and 4th decades of life and typically found in only 10% of patients at the time of presentation (70). Thus, annual screening for GH hypersecretion is recommended only in post pubertal patients. As in cases of gigantism/acromegaly in the setting of MAS, diffuse pituitary hyperplasia (71) and concomitant hyperprolactinemia (72) are frequently seen in individuals with CNC and GH excess.

3PA Association

The constellation of paraganglioma, pheochromocytoma and pituitary adenoma is termed 3PA Association and has been shown to be due to germline mutations in a variety of subunits of succinate dehydrogenase (65;73). Growth hormone excess typically occurs in the 3rd and 4th decades of life (74). To date, no pediatric patients with gigantism in the setting of the 3PA phenotype have been reported.

Familial Somatotropinomas

It has long been recognized that isolated pituitary gigantism or acromegaly may occur in a familial pattern. This phenomenon, termed "Familial Isolated Pituitary Adenomas" (FIPA), is defined as the development of GH hypersecretion in two or more members of a family that does not exhibit features of MEN-1 or CNC. At least 46 different affected kindreds have been reported (75). Unlike in MEN-1 and CC, GH excess tends to arise fairly early in life, with 70% of those with the disorder diagnosed before the 3rd decade. Early childhood gigantism in this setting has also occurred. involving sisters with abnormal linear growth since age 5 (76) and a more virulent course than is seen in sporadic somatotropinomas has been suggested by a case series (77). Once assumed to represent a variant of MEN-1, mutations within the menin gene as the etiology for IFS were conclusively excluded (78;79). However, the precise molecular genetic basis for the development of pituitary GH-secreting adenomas in the majority of affected families has eluded detection. Initial investigation revealed loss of heterozygosity and linkage to a 9.7 Mb region of 11g13, suggesting the presence of an additional putative tumor suppressor gene in this region, distinct from that involved in MEN-1. Subsequent studies identified inactivating mutations in the gene encoding aryl hydrocarbon receptor interacting protein (AIP) at 11q13.3 in 15%-25% of families with FIPA (80-82) making it the most common genetic defect found in these kindreds. Although the mechanism by which these mutations cause pituitary adenomas is unknown, the resulting phenotype is characterized by early-onset and aggressive disease. In an amazing case of medical sleuthing, a aermline AIP mutation identified in DNA from the preserved teeth of an 18th century Irish giant was found to be an exact match for the mutation harbored by four contemporary Irish families with FIPA, indicating a common ancestor dating back more than 50 generations ago (83)! Interestingly, a second potential locus for FIPA has mapped to 2p12-16, very close to the region implicated in a number of kindreds with CNC (84). Additional molecular genetic analysis performed in these patients has included a search for germline mutations within the GHRH receptor gene, Gsa and Gi2a genes, all of which were normal. Similar to observations in MEN-1, patients with FIPA have discreet pituitary adenomas, the majority of which are comprised solely of

somatotrophs (75). However, prolactinomas, gonadotropinomas and silent pituitary adenomas may all be seen in different members of the same kindred (85;86). Macroadenomas with invasion into the cavernous sinus are common in the setting of FIPA, and treatment is notoriously difficult (86).

X-Linked Acrogigantism

An additional cause of familial gigantism and acromegaly has been linked to microduplication of Xq26.3 and termed X-linked acrogigantism (X-LAG). This genomic duplication was initially identified in 14 patients with gigantism and is associated with both sporadic and familial cases (87;88). Of the four genes contained in the duplicated region, the growth hormone excess appears to result from an abnormality of GPR101, a gene that encodes for an orphan G-protein coupled receptor. This gene is markedly over-expressed in pituitary tissue from affected patients. Functional studies suggest a proliferative role for mutant GPR101, although the precise mechanism for how this aberration contributes to GH hypersecretion is not yet clear. The condition can result from either germline or somatic duplications in GPR101 and has a female predominance (89;90). Mosaicism for GPR101 duplication resulting in X-LAG has also been reported in sporadic cases involving boys (91). Patients harboring the Xq26.3 microduplication exhibit a distinct phenotype characterized by strikingly early gigantism with a median age of onset of 12 months. In addition to hypersecretion of GH, elevated circulating GHRH and prolactin have also been noted (92). Both pituitary adenomas and pituitary hyperplasia have been seen among cases testing positive for X-LAG. This discovery highlights new biological processes that will undoubtedly lead to novel insights regarding the central regulation of human growth.

CLINICAL AND BIOCHEMICAL FEATURES OF GIGANTISM

As would be predicted, linear growth acceleration is the cardinal feature of excessive GH production in a child or adolescent. Based on numerous case reports, however, it is clear that the excessive linear growth observed in young children with gigantism may be accompanied or even preceded by macrocephaly and or obesity (9;11). In a large international study of patients with pituitary gigantism, the median onset of rapid growth was 13 years and occurred earlier in girls than in boys (93). Additional clinical features frequently encountered include frontal bossing, broad nasal bridge, prognathism, excessive sweating, voracious appetite, coarse facial features and enlargement of the hands and feet. Bone age radiographs in these patients have variably been reported to be normal or advanced, even in the complete absence of sex steroid production. Figure 3 demonstrates the prognathism, coarse facial features and typical tall stature seen in a 12-year-old boy with gigantism, and Figure 4 illustrates enlargement of the hands in this same patient.

Figure 3: Twelve-year-old boy with pituitary gigantism measuring 6'5" with his mother. Note the coarse facial features and prominent jaw.



Figure 4: Enlarged hand of the same patient in comparison with the hand of an adult male with a height of 6'1". The patient's middle digit has a circumference of 9 centimeters.



The most consistent biochemical abnormality observed in patients with gigantism is an elevated IGF-1, which is known to exhibit an excellent correlation with 24-hour GH secretion (94). As previously mentioned, hyperprolactinemia is extremely common in early-onset GH hypersecretion. Depending on the individual situation, the additional pituitary screening evaluation may be normal, indicative of hypopituitarism or central precocious puberty. Concurrent endocrinopathies may also be present, particularly in patients with syndromes such as MAS or MEN-1. Rarely, alterations in glucose tolerance brought about by GH excess may result in the development of overt diabetes, leading to transient diabetic ketoacidosis (95-97) which may even be the presenting feature in rare instances (98). An additional physiologic effect of GH excess that may have clinical significance is that of increased erythropoiesis, as demonstrated by a case of acromegaly-induced polycythemia vera that resolved following surgical resection of the GH-secreting adenoma(99).. The importance of GH in the regulation of red blood cell production has further been supported by the observation that pre-treatment hemoglobin concentrations in children with idiopathic growth hormone deficiency are lower than controls (100)

DIAGNOSTIC EVALUATION OF GH EXCESS

The gold standard for making the diagnosis of GH excess relies on the inability to suppress serum GH to an appropriate level following an oral glucose load. While the OGTT has been the diagnostic test of choice for many years, numeric guidelines for the expected degree of suppression in a normal individual have steadily decreased. This trend is the direct result of newer assays with an improved threshold of sensitivity for detection (101). A normal response to a standardized glucose bolus (1.75 gm/kg up to 75 grams) utilizing the newer IRMA/ICMA assays is considered to be a GH level below 1 ng/ml (102). However, given the observation that biochemical recurrence of GH excess may be detected in patients with a GH nadir less than 1 ng/dl, and that healthy subjects nearly always suppress to below 0.14 ng/ml, some investigators have suggested that this cut-off is

too liberal (103). Typically, the nadir in serum GH is expected to occur within the first 2 hours of the test. Along with a lack of suppression following oral glucose, individuals with GH hypersecretion characteristically exhibit a paradoxical response to other forms of hypothalamic-pituitary stimulation testing.

Occasionally, 24 hour integrated GH assessment may be helpful in cases in which an equivocal response to OGTT is seen (104). Despite the development of highly sensitive GH assays, generalizability of results across institutions or regions is hampered by significant heterogeneity in the availability of reference preparations and methods used by specific laboratories (105). Depending on the individual circumstance, measurement of peripheral GHRH may also be indicated to investigate the possibility of ectopic GHRH secretion. Once biochemical evidence of GH excess has been demonstrated, MRI scanning of the H-P region is obviously the next step. Figure 5 illustrates the typical appearance of a GH-secreting pituitary macroadenoma in an adolescent with gigantism.



Figure 5: Pituitary somatotroph macroadenoma in an adolescent with gigantism.

A potential pitfall in the evaluation of gigantism in adolescents is the fact that significant elevations of IGF-1 may be present during normal puberty (106). Moreover, growth hormone response to an oral glucose load in normal children has been found to be gender and pubertal-stage specific, with the

highest nadir GH occurring in Tanner stage 2-3 girls (107). The effect of sex steroids on IGF-1 and GH suppression must also be taken into account when a diagnosis of gigantism is being considered in a child with concurrent precocious puberty, as may be the case in NF-1 or MAS. Adding to the possible diagnostic ambiguity is the fact that a significant percentage of normal tall adolescents fail to suppress serum GH in response to oral glucose testing (108). Therefore, both screening and definitive testing for GH excess should be performed in the context of clinical suspicion, and IGF-1 levels interpreted according to age and pubertal stage-adjusted normal ranges.

TREATMENT

No large-scale studies evaluating various therapeutic approaches to the treatment of GH excess in pediatric patients are available. Therefore, the optimal treatment of gigantism has traditionally been extrapolated from the adult literature as well as case reports or small series involving children. As is the case in adults, the three separate modalities available for the treatment of children and adolescents with GH hypersecretion are surgery, radiation and medical therapy. Of these, the greatest recent advances by far have occurred in the realm of pharmacologic agents, resulting in an exciting armamentarium of drugs promising truly enhanced efficacy and excellent safety. Regardless of the individual treatment strategy, the goals of therapy remain the same, namely the restoration of GH and IGF-1 levels to normal ranges (109). Of all parameters investigated, GH levels themselves appear to correlate most with overall morbidity and mortality in acromegaly (110). Table 3 summarizes the current therapeutic options as they pertain to pediatric patients, each of which is discussed below.

Modality	Specific Options	Current Indications	Pediatric Experience
Surgery	Transphenoidal resection	Pituitary microadenoma or	Performed safely in children as young as 2 years old
Radiation	Conventional radiation	Adjuvant to surgical or medical therapy	Typically avoided if at all possible, but has been used as adjuvant therapy
Naulation	Stereotactic radiosurgery, ex: gamma knife	Adjuvant therapy in patients with residual GH hypersecretion	No experience with use in children
Medical Therapy	Somatostatin analogues Octreotide (Sandostatin) Lanreotide 	 Primary therapy in cases of diffuse pituitary hyperplasia or severe bone disease Adjuvant to surgery or radiation 	Used safely in children with both sporadic and syndromic gigantism for extended periods of time alone and in combination with dopamine analogues

Table 3: Therapeutic Modalities in GH	H Excess and Experience with	Use in Pediatric Patients.
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Depot somatostatin analogues • Sandostatin LAR SR-lanreotide	Same as above	Safety and efficacy appear comparable to non-depot preparations
Dopamine agonists Bromocriptine Cabergoline	 Adjuvant to somatostatin analogues and other therapies Particularly useful when concurrent hyperprolactinemia is present 	Used safely in children in combination with somatostatin analogues
GH receptor antagonists Pegvisomant	Particularly useful for treatment of refractory disease	Has been used alone and in combination with somatostatin analogues Preliminary experience in children appears promising

Surgery

Transphenoidal resection is the treatment of choice for discreet pituitary microadenomas or macroadenomas (111), with the objective being the preservation of pituitary function in association with cure of the GH excess. Not surprisingly, individual surgeon expertise has a significant impact on the likelihood of success (112), which is exemplified by a rapid normalization of serum GH levels (often within one hour) and response to OGTT. However, while surgery cures the majority of patients with microadenomas, less than 50% of patients with macroadenomas experience this optimal outcome (113;114). Moreover, extended post-operative follow-up has revealed a gradual return of GH excess over time in a substantial number of patients in whom the disease was previously deemed to be well controlled (115;116). Experience with surgical treatment of gigantism in children and adolescents has been comparable to that observed in adults (117;118), and it has been employed safely in patients as young as 24 months (12). Although further investigation is needed, a potential role for pre-operative medical therapy has been suggested by studies indicating higher surgical remission rates and lower anesthesia risk following a several month course of a somatostatin analogue (119).

Radiation

Although traditionally included as a therapeutic option, significant problems exist with the use of conventional radiotherapy in gigantism or acromegaly. These include a low level of efficacy, delayed normalization of GH levels, and a high incidence of hypopituitarism. In the setting of MAS, radiation therapy for GH hypersecretion may contribute to malignant transformation of dysplastic bone tissue (120). Additional concerns particularly relevant to children include potential neurocognitive effects and the possible development of hypothalamic obesity, both of which have been linked to cranial irradiation in pediatric patients (120;121). Therefore, radiation therapy would be considered a last resort for the treatment of childhood GH hypersecretion. Improved precision and safety is observed with use of stereotactic radiosurgery in the form of the gamma knife technique, which has been successfully employed as adjuvant therapy in adults with acromegaly (120;122-124).

Medical Therapy

Although most commonly considered adjunctive to surgery or radiation, a primary role for medical therapy has always existed for those patients with diffuse pituitary hyperplasia or severe bony deformities precluding a surgical approach. As tremendous improvements in the pharmacologic agents available for use in GH excess continues to evolve (125), the number of patients offered medical therapy as first-line treatment will surely expand. The three currently existing classes of drugs for suppression of GH and IGF-1 levels are reviewed below.

SOMATOSTATIN ANALOGUES

Ever since their development in the mid-1980's, long acting analogues of somatostatin have held a pivotal place in the medical treatment of GH excess. These agents exert their effect through selective binding to somatostatin receptors within somatotroph adenomas (126). By far the greatest experience in the United States has been with octreotide, which is typically administered subcutaneously in three divided doses. Short-term administration of octreotide results in a decrease in GH levels within one hour in > 90% of patients with acromegaly (127), while sustained use

normalizes GH and IGF-1 levels in up to 65% of patients (128). Experience with the use of octreotide in children has been similarly favorable, where it has been beneficial in the treatment of sporadic as well as syndromic gigantism (129;130). Continuous subcutaneous infusion of octreotide has also resulted in superior efficacy in controlling GH hypersecretion in a pubertal patient (131).Long-acting depot preparations of octreotide in the form of Sandostatin LAR and SR-lanreotide are also available, in which a slow release of drug is achieved through degradation of a polymer in which microspheres are encapsulated (132). This allows for monthly IM administration, resulting in a safety and efficacy profile that is comparable to or improved in contrast to traditional dosing (133). Both slow-release preparations have also been used in the treatment of ectopic forms of GH excess (134) and in MAS associated gigantism (135-137), and have been noted to have equivalent safety and efficacy (138). The development of novel somatostatin analogues has the potential to improve efficacy over existing compounds (139). The major side effect of all the somatostatin analogues is a significantly increased risk of biliary sludge and gallstones after sustained use, necessitating periodic ultrasound examinations in patients treated long-term (140).

DOPAMINE AGONISTS

Although rarely effective alone, dopamine agonists have a valuable role as adjunctive agents in the treatment of GH excess. Due to their suppressive effects on prolactin, these drugs are particularly advantageous when hyperprolactinemia is present, as is often the case in childhood-onset gigantism. Both bromocriptine and the more potent dopamine agonists such as cabergoline have been administered to children in combination with octreotide long-term with no apparent adverse effects (136).

GH RECEPTOR ANTAGONISTS

The latest development in the realm of medical therapy has been the emergence of pegvisomant, a genetically engineered human GH analogue that acts as a highly selective GH antagonist (141). This is achieved through alterations in affinity binding of pegvisomant compared to the native GH molecule (129), resulting in prevention of the normal extracellular dimerization of the growth hormone receptor. Administration of pegvisomant long-term to adults with acromegaly has been shown to result in normalization of serum IGF-1 levels in 97% of patients (142). Despite these extremely promising results, the implications of the nearly ubiquitous elevations in serum GH levels observed in conjunction with pegvisomant treatment initially created some concerns. Although early reports recounted an increase in tumor volume and abnormal liver enzymes in association with peqvisomant use (143:144), long-term follow has demonstrated that these complications are rare and that efficacy is very good (145;146). Combination therapy using pegvisomant along with a dopamine agonist or somatostatin analogue also appears promising (145). Thus far, preliminary experience with the use of pegvisomant alone or in combination with a somatostatin analogue for the treatment of gigantism in children also appears favorable (147). This approach resulted in successful normalization of IGFI levels in a 4 year old with NF-1 (148), a 12 year old with MAS (149), and a couple of children with persistent GH hypersecretion following surgical removal of a pituitary adenoma who had failed a somatostatin analogue (1 5 0 ; 1 5 1). Even more reassuring is a report of long-term (up to 3.5 years) treatment using pegvisomant in 3 children w it h gigantism, all of whom experienced a decline in growth velocity and resolution of acromegalic features (152).

Treatment of Tall Stature

Depending on the absolute height and the degree of growth potential remaining, one of the goals in the treatment of gigantism may be prevention of further linear growth. When this is the case, acceleration of epiphyseal fusion can be achieved with exogenous sex steroids (153). Short-term administration of both high dose testosterone and estrogen have been utilized for this purpose in children with gigantism, resulting in significant improvements in terms of adult height (154;155). However, such an approach would require great caution given reports of subfertility in women who were treated with high dose estrogen during adolescence with the goal of attenuating growth in the setting of constitutional tall stature (156;157).

OVERGROWTH SYNDROMES

The overgrowth syndromes comprise a diverse group of conditions with unique clinical, behavioral and molecular genetic features. While considerable overlap in presentation sometimes exists (158), advances in identification of the precise etiology of specific overgrowth disorders continues to improve the clinician's ability to make an accurate diagnosis. In this chapter, only the most common syndromes characterized by generalized somatic overgrowth will be reviewed, with specific aspects pertaining to each disorder summarized in Table 4. Additional syndromes in which tall stature (such as Marfan syndrome) or obesity (such as Prader-Willi syndrome) are the predominant features will be discussed elsewhere.

Syndrome	Clinical Features	Mode of Inheritance	Etiology
Sotos syndrome	 Prenatal overgrowth with early transient growth acceleration 	SporadicRare familial cases	 Mutations within the NSD1 gene in 90% of cases
	 Macrocephaly, hypertelorism, prominent forehead, pointed chin 		
	 Speech and language delay 		
	clumsiness		

Table 4: Comparison of Overgrowth Syndromes

Beckwith- Wiedemann syndrome	 Prenatal and postnatal overgrowth Macroglossia, abdominal wall defects, ear creases, visceromegaly Neonatal hypoglycemia Increased incidence of embryonal tumors 	 Sporadic Rare familial cases 	Abnormal imprinting in growth regulatory genes at 11p15 including IGF-2
Simpson- Golabi- Behmel syndrome	 Prenatal and postnatal overgrowth Macroglossia, skeletal/hand anomalies, supernumerary nipples, visceromegaly, cardiac abnormalities Increased incidence of 	X-linkedSporadic	• Mutations within the GPC3 gene at Xq26
Weaver syndrome NSD1-nuclear re	 Prenatal or postnatal overgrowth Macrocephaly, hypertelorism, large ears, micrognathia Advanced skeletal maturation 	 Autosomal dominant Sporadic taining protein 1, GPG3-glyp 	Mutations within EZH2

Sotos Syndrome

Sotos syndrome, also known as cerebral gigantism, was first described in 1964 (159). Since then, several hundred cases have been reported. Cardinal features of the disorder include early onset overgrowth, a characteristic facial configuration and stereotypical behavioral profile. The overgrowth in Sotos syndrome is of prenatal onset, with length being the most significantly affected parameter.

After birth, acceleration of all growth parameters ensues, with OFC measuring above the 97th percentile in nearly all affected infants by 12 months of age (160). Although the growth velocity slows by age 3 or 4, height invariably remains above the normal range throughout childhood, typically in association with somewhat lower weight percentiles. In contrast, adult stature in Sotos

syndrome is usually within the normal range for the general population (161), which has been attributed to the combination of an advanced bone age and a relatively early onset of puberty. Classic facial features include macrocephaly with dolichocephaly, hypertelorism, high-arched palate, prominent forehead and a pointed chin (162). Additional oral findings may include premature tooth eruption and supernumerary teeth(163). Developmental delay is ubiquitous, particularly in the area of speech and language acquisition (163-165) .Children with Sotos syndrome are often described as being clumsy, with a tendency toward aggressive behavior (163;166). A minority have seizures, as well as structural abnormalities of the brain such as enlarged ventricles and absence of the corpus callosum.

Sotos syndrome is typically sporadic, although autosomal dominant transmission has been reported(163;167). Isolated cases of identical twin pairs who are concordant as well as discordant for the condition have also been described (163;168). Historically, the diagnosis was based entirely on clinical criteria. However, it is now known that Sotos syndrome is caused by a variety of molecular genetic alterations resulting in haploinsufficiency of the nuclear receptor-binding SET domain-containing protein 1 (NSD1) gene at 5q35 (163;169-171) in ~90% of cases. The NSD1 gene encodes for a nuclear protein believed to function as a basic transcription factor and transcriptional regulator. Heterozygous mutations in the NFIX gene (Nuclear Factor I, X) have also been identified in some children with Sotos syndrome (172). While genotype-phenotype correlations have been suggested (173;174), this needs to be confirmed by additional studies of affected patients.

Beckwith-Wiedemann Syndrome (BWS)

Two physicians independently reported the first recognized cases of BWS in the 1960's (175;176). Since that time, tremendous progress has been made in unraveling several aspects of this complex disorder. BWS is typified by the combination of prenatal and postnatal overgrowth, congenital malformations and a predisposition to embryonal tumors. Characteristic features noted in the neonatal period include macroglossia, abdominal wall defects such as umbilical hernia, ear creases, visceromegaly and hyperinsulinemic hypoglycemia (177). A variety of additional abnormalities are found in a subset of patients, including hemihypertrophy or isolated facial asymmetry(178). While intelligence may be normal, mild to moderate developmental delay may also be present. Although usually sporadic, several families manifesting heterogeneous inheritance patterns have been reported in whom there are several generations of affected individuals (179). The reported incidence of malignancy in children with BWS varies between 4-21% (180), with the majority consisting of Wilms tumor. Therefore, frequent screening via abdominal ultrasonography during infancy and early childhood is essential (181), especially in patients with hemihypertrophy, which is known to be associated with an increased risk of cancer (182). Insights into the pathophysiology of the abnormal growth in this condition emerged with the discovery of abnormalities in imprinting of a number of growth regulatory genes within three regions of chromosome 11p15, including IGF-2, H19 and CDKN1C (181;183). The molecular genetic defects resulting in BWS are extremely heterogeneous, and include maternal hypomethylation of 11p15, paternal uniparental disomy of this region, and unbalanced translocations leading to trisomy of the 11p15 locus (184;185). Enhanced understanding of the relationship between tumor risk and the molecular subtype in BWS will result in improvements in targeted screening (186;187). Interestingly, an association has been noted between assisted reproduction and risk of imprinting disorders such as BWS (188), although the risk appears to be small (189). Figure 6 demonstrates several classic clinical features in a child with BWS.

Figure 6: Young child with Beckwith-Wiedemann syndrome. Note the macroglossia, prominent eyes, eyelid nevus flammeus and barely visible linear ear creases.



Simpson-Golabi-Behmel syndrome (SGBS)

SGBS is a complex X-linked overgrowth disorder sharing many features with BWS. It is characterized by prenatal and postnatal overgrowth, coarse facial features and congenital anomalies. Some of the most commonly reported abnormalities include skeletal/hand defects, supernumerary nipples, macroglossia and visceromegaly However, a wide spectrum in severity has been noted, ranging from mild features in carrier females to a lethal form of the disorder in affected males (190). Similarly, cognitive abilities vary from within the normal range to severe developmental delays. Approximately 36% of patients have a cardiac abnormality, the most common of which is a cardiovascular malformation (191). As is the case in BWS, an increased incidence of embryonal tumors during early life is present. Delineation of the molecular genetic cause of SGBS has provided significant insight as to the reason for the striking similarities between this disorder and BWS. Inactivating mutations of the glypican-3 (GPC3) gene at Xq26 have been demonstrated in 28-70% of individuals with SGBS (192;193). GPC3 is a member of a multigene family known to have critical roles in growth and development through the modulation of cellular responses to growth factors, including IGF-2 (194). Exactly how abnormal levels of GPC3 promote tumorigenesis is poorly understood, but it may be through a disruption of the normal GPC3/IGF-2 complex, which is believed to be involved in IGF-2 modulation (195). An alternative proposal is that the physical manifestations of SGBS are due to abnormal interaction between GPC3 and CD26, a protein with important roles in the regulation of cell growth and immunologic response (196). Application of GPC3 mutational analysis in patients with unspecified overgrowth conditions has resulted in an extension of the SGBS phenotype (197) and the establishment of an international registry will be invaluable in providing information regarding the natural history and pathophysiology of this interesting condition. The oldest case of SGBS on record was discovered in an anatomical museum in the form of a macrosomic newborn who had

died neonatally from unknown causes and was traced through following the family tree of a newly identified GPC3 mutation in an affected patient (198).

Weaver Syndrome

Weaver syndrome is a rare condition that was first reported in 1974 (199). Major features include prenatal or postnatal overgrowth, characteristic facies and advanced skeletal maturation. The typical appearance includes tall stature, macrocephaly, hypertelorism, large ears and micrognathia. A subset of patients have been reported to have cervical spine abnormalities (200), and the occasional development of neoplasia has also been noted in this population. The majority of individuals with Weaver syndrome have developmental delay, which is typically mild. Initially believed to be sporadic, multiple instances of familial occurrence have pointed strongly toward an autosomal dominant form of transmission (201). In 2011, mutations in the histone methytransferase, EZH2, were shown to cause Weaver syndrome (174). Heterozygous mutations in embryonic ectoderm development (EED) have also been identified in patients with Weaver syndrome (202;203). Significant phenotypic overlap between Weaver syndrome and Sotos syndrome often makes it difficult to differentiate between these overgrowth conditions (158;202). Thus, the availability of molecular genetic testing will aid in the diagnostic process.

CONCLUSION

In summary, the differential diagnosis of gigantism includes a significant number of heterogeneous disorders exhibiting a vast array of clinical and genetic features (204). In most cases, the history, physical examination and adjunctive biochemical and/or molecular genetic testing will ultimately reveal the likely diagnosis. Albeit rare, diseases resulting in gigantism afford the unique opportunity for a glimpse into the complex mechanisms of growth regulation. Thus, continued clinical and scientific investigation will enhance not only individual patient care, but also collective insight into the intricacies of the fundamental processes of human growth.

REFERENCES

1. Chentli F, Azzoug S, Amani MA et al. Etiologies and clinical presentation of gigantism in Algeria. Horm Res Paediatr 2012; 77(3):152-155.

2. Daughaday WH. Pituitary gigantism. Endocrinol Metab Clin North Am 1992; 21(3):633-647.

3. Etxabe J, Gaztambide S, Latorre P, Vazquez JA. Acromegaly: an epidemiological study. J Endocrinol Invest 1993; 16(3):181-187.

- 4. Sotos JF. Overgrowth. Hormonal Causes. Clin Pediatr (Phila) 1996; 35(11):579-590.
- 5. Lodish MB, Trivellin G, Stratakis CA. Pituitary gigantism: update on molecular biology and management. Curr Opin Endocrinol Diabetes Obes 2016; 23(1):72-80.
- 6. Melmed S. Acromegaly. N Engl J Med 1990; 322(14):966-977.
- 7. Moran A, Asa SL, Kovacs K et al. Gigantism due to pituitary mammosomatotroph hyperplasia. N Engl J Med 1990; 323(5):322-327.
- 8. Zimmerman D, Young WF, Jr., Ebersold MJ et al. Congenital gigantism due to growth hormone-releasing hormone excess and pituitary hyperplasia with adenomatous transformation. J Clin Endocrinol Metab 1993; 76(1):216-222.

9. Dubuis JM, Deal CL, Drews RT et al. Mammosomatotroph adenoma causing gigantism in an 8-year old boy: a possible pathogenetic mechanism. Clin Endocrinol (Oxf) 1995; 42(5):539-

549.

 Felix IA, Horvath E, Kovacs K, Smyth HS, Killinger DW, Vale J. Mammosomatotroph adenoma of the pituitary associated with gigantism and hyperprolactinemia. A morphological study including immunoelectron microscopy. Acta Neuropathol (Berl) 1986; 71(1-2):76-82.
 Blumberg DL, Sklar CA, David R, Rothenberg S, Bell J. Acromegaly in an infant.

Pediatrics 1989; 83(6):998-1002.

12. Gelber SJ, Heffez DS, Donohoue PA. Pituitary gigantism caused by growth hormone excess from infancy. J Pediatr 1992; 120(6):931-934.

13. Vidal S, Horvath E, Kovacs K, Lloyd RV, Smyth HS. Reversible transdifferentiation: interconversion of somatotrophs and lactotrophs in pituitary hyperplasia. Mod Pathol 2001; 14(1):20-28.

14. Hannah-Shmouni F, Trivellin G, Stratakis CA. Genetics of gigantism and acromegaly. Growth Horm IGF Res 2016; 30-31:37-41.

15. Araki Y, Sakai N, Andoh T, Yoshimura S, Yamada H. Central neurocytoma presenting with gigantism: case report. Surg Neurol 1992; 38(2):141-145.

16. Isidro ML, Iglesias DP, Matias-Guiu X, Cordido F. Acromegaly due to a growth hormone-releasing hormone-secreting intracranial gangliocytoma. J Endocrinol Invest 2005; 28(2):162-165.

17. Asada H, Otani M, Furuhata S, Inoue H, Toya S, Ogawa Y. Mixed pituitary adenoma and gangliocytoma associated with acromegaly--case report. Neurol Med Chir (Tokyo) 1990; 30(8):628-632.

18. Asa SL, Scheithauer BW, Bilbao JM et al. A case for hypothalamic acromegaly: a clinicopathological study of six patients with hypothalamic gangliocytomas producing growth hormone-releasing factor. J Clin Endocrinol Metab 1984; 58(5):796-803.

19. Ghazi AA, Amirbaigloo A, Dezfooli AA et al. Ectopic acromegaly due to growth hormone releasing hormone. Endocrine 2013; 43(2):293-302.

20. Faglia G, Arosio M, Bazzoni N. Ectopic acromegaly. Endocrinol Metab Clin North Am 1992; 21(3):575-595.

21. Beuschlein F, Strasburger CJ, Siegerstetter V et al. Acromegaly caused by secretion of growth hormone by a non-Hodgkin's lymphoma. N Engl J Med 2000; 342(25):1871-1876.

22. Garby L, Caron P, Claustrat F et al. Clinical characteristics and outcome of acromegaly induced by ectopic secretion of growth hormone-releasing hormone (GHRH): a French nationwide series of 21 cases. J Clin Endocrinol Metab 2012; 97(6):2093-2104.

23. Biswal S, Srinivasan B, Dutta P et al. Acromegaly caused by ectopic growth hormone: a rare manifestation of a bronchial carcinoid. Ann Thorac Surg 2008; 85(1):330-332.

24. Doga M, Bonadonna S, Burattin A, Giustina A. Ectopic secretion of growth hormonereleasing hormone (GHRH) in neuroendocrine tumors: relevant clinical aspects. Ann Oncol 2001; 12 Suppl 2:S89-S94.

25. Madonna D, Kendler A, Soliman AM. Ectopic growth hormone-secreting pituitary adenoma in the sphenoid sinus. Ann Otol Rhinol Laryngol 2001; 110(1):99-101.

26. Appel JG, Bergsneider M, Vinters H, Salamon N, Wang MB, Heaney AP. Acromegaly due to an ectopic pituitary adenoma in the clivus: case report and review of literature. Pituitary 2012; 15 Suppl 1:S53-S56.

 Osella G, Orlandi F, Caraci P et al. Acromegaly due to ectopic secretion of GHRH by bronchial carcinoid in a patient with empty sella. J Endocrinol Invest 2003; 26(2):163-169.
 Glasker S, Vortmeyer AO, Lafferty AR et al. Hereditary pituitary hyperplasia with infantile gigantism. J Clin Endocrinol Metab 2011; 96(12):E2078-E2087.

29. Costin G, Fefferman RA, Kogut MD. Hypothalamic gigantism. J Pediatr 1973; 83(3):419-

425.

30. Drimmie FM, MacLennan AC, Nicoll JA, Simpson E, McNeill E, Donaldson MD. Gigantism due to growth hormone excess in a boy with optic glioma. Clin Endocrinol (Oxf) 2000; 53(4):535-538.

31. Duchowny MS, Katz R, Bejar RL. Hypothalamic mass and gigantism in neurofibromatosis: treatment with bromocriptine. Ann Neurol 1984; 15(3):302-304.

32. Josefson JL, Listernick R, Charrow J, Habiby RL. Growth Hormone Excess in Children with Optic Pathway Tumors Is a Transient Phenomenon. Horm Res Paediatr 2016; 86(1):35-38.

33. Sani I, Albanese A. Endocrine Long-Term Follow-Up of Children with Neurofibromatosis Type 1 and Optic Pathway Glioma. Horm Res Paediatr 2017; 87(3):179-188.

34. Fuqua JS, Berkovitz GD. Growth hormone excess in a child with neurofibromatosis type 1 and optic pathway tumor: a patient report. Clin Pediatr (Phila) 1998; 37(12):749-752.

35. Manski TJ, Haworth CS, Duval-Arnould BJ, Rushing EJ. Optic pathway glioma infiltrating into somatostatinergic pathways in a young boy with gigantism. Case report. J Neurosurg 1994; 81(4):595-600.

36. Waguespack SG, Eugster EA, Pescovitz OH. Growth hormone (GH) excess in a child with neurofibromatosis type 1 (NF1) an optic pathway glioma. Pediatric Research 49[6 Suppl 2 of 2], 82A. 2001. Ref Type: Abstract

37. Cambiaso P, Galassi S, Palmiero M et al. Growth hormone excess in children with neurofibromatosis type-1 and optic glioma. Am J Med Genet A 2017; 173(9):2353-2358.
38. Josefson JL, Listernick R, Charrow J, Habiby RL. Growth Hormone Excess in Children with Optic Pathway Tumors Is a Transient Phenomenon. Horm Res Paediatr 2016; 86(1):35-38.

39. Salenave S, Boyce AM, Collins MT, Chanson P. Acromegaly and McCune-Albright syndrome. J Clin Endocrinol Metab 2014; 99(6):1955-1969.

40. Weinstein LS, Shenker A, Gejman PV, Merino MJ, Friedman E, Spiegel AM. Activating mutations of the stimulatory G protein in the McCune-Albright syndrome. N Engl J Med 1991; 325(24):1688-1695.

41. Lumbroso S, Paris F, Sultan C. McCune-Albright syndrome: molecular genetics. J Pediatr Endocrinol Metab 2002; 15 Suppl 3:875-882.

42. Christoforidis A, Maniadaki I, Stanhope R. McCune-Albright syndrome: growth hormone and prolactin hypersecretion. J Pediatr Endocrinol Metab 2006; 19 Suppl 2:623-625.

43. Yao Y, Liu Y, Wang L et al. Clinical characteristics and management of growth hormone excess in patients with McCune-Albright syndrome. Eur J Endocrinol 2017; 176(3):295-303.
44. Tinschert S, Gerl H, Gewies A, Jung HP, Nurnberg P. McCune-Albright syndrome: clinical and molecular evidence of mosaicism in an unusual giant patient. Am J Med Genet 1999; 83(2):100-108.

45. Vogl TJ, Nerlich A, Dresel SH, Bergman C. CT of the "Tegernsee Giant": juvenile gigantism and polyostotic fibrous dysplasia. J Comput Assist Tomogr 1994; 18(2):319-322.
46. Akintoye SO, Chebli C, Booher S et al. Characterization of gsp-mediated growth hormone excess in the context of McCune-Albright syndrome. J Clin Endocrinol Metab 2002; 87(11):5104-5112.

47. Collins MT, Singer FR, Eugster E. McCune-Albright syndrome and the extraskeletal manifestations of fibrous dysplasia. Orphanet J Rare Dis 2012; 7 Suppl 1:S4.

48. Boyce AM, Glover M, Kelly MH et al. Optic neuropathy in McCune-Albright syndrome: effects of early diagnosis and treatment of growth hormone excess. J Clin Endocrinol Metab 2013; 98(1):E126-E134.

49. Vortmeyer AO, Glasker S, Mehta GU et al. Somatic GNAS mutation causes widespread and diffuse pituitary disease in acromegalic patients with McCune-Albright syndrome. J Clin Endocrinol Metab 2012; 97(7):2404-2413.

50. Dotsch J, Kiess W, Hanze J et al. Gs alpha mutation at codon 201 in pituitary adenoma causing gigantism in a 6-year-old boy with McCune-Albright syndrome. J Clin Endocrinol Metab 1996; 81(11):3839-3842.

51. Zumkeller W, Jassoy A, Lebek S, Nagel M. Clinical, endocrinological and radiography features in a child with McCune-Albright syndrome and pituitary adenoma. J Pediatr Endocrinol Metab 2001; 14(5):553-559.

52. Cuttler L, Jackson JA, Saeed uz-Zafar M, Levitsky LL, Mellinger RC, Frohman LA. Hypersecretion of growth hormone and prolactin in McCune-Albright syndrome. J Clin Endocrinol Metab 1989; 68(6):1148-1154.

53. Shimon I, Melmed S. Genetic basis of endocrine disease: pituitary tumor pathogenesis. J Clin Endocrinol Metab 1997; 82(6):1675-1681.

54. Mantovani G, Bondioni S, Lania AG et al. Parental origin of Gsalpha mutations in the McCune-Albright syndrome and in isolated endocrine tumors. J Clin Endocrinol Metab 2004; 89(6):3007-3009.

55. Skogseid B, Rastad J, Oberg K. Multiple endocrine neoplasia type 1. Clinical features and screening. Endocrinol Metab Clin North Am 1994; 23(1):1-18.

56. Guru SC, Goldsmith PK, Burns AL et al. Menin, the product of the MEN1 gene, is a nuclear protein. Proc Natl Acad Sci U S A 1998; 95(4):1630-1634.

57. Bassett JH, Forbes SA, Pannett AA et al. Characterization of mutations in patients with multiple endocrine neoplasia type 1. Am J Hum Genet 1998; 62(2):232-244.

58. Mutch MG, Dilley WG, Sanjurjo F et al. Germline mutations in the multiple endocrine neoplasia type 1 gene: evidence for frequent splicing defects. Hum Mutat 1999; 13(3):175-185.

59. Boggild MD, Jenkinson S, Pistorello M et al. Molecular genetic studies of sporadic pituitary tumors. J Clin Endocrinol Metab 1994; 78(2):387-392.

60. Carty SE, Helm AK, Amico JA et al. The variable penetrance and spectrum of manifestations of multiple endocrine neoplasia type 1. Surgery 1998; 124(6):1106-1113.

61. Stratakis CA, Schussheim DH, Freedman SM et al. Pituitary macroadenoma in a 5-yearold: an early expression of multiple endocrine neoplasia type 1. J Clin Endocrinol Metab 2000; 85(12):4776-4780.

62. Brandi ML, Gagel RF, Angeli A et al. Guidelines for diagnosis and therapy of MEN type 1 and type 2. J Clin Endocrinol Metab 2001; 86(12):5658-5671.

63. Sala E, Ferrante E, Verrua E et al. Growth hormone-releasing hormone-producing pancreatic neuroendocrine tumor in a multiple endocrine neoplasia type 1 family with an uncommon phenotype. Eur J Gastroenterol Hepatol 2013; 25(7):858-862.

64. Lee M, Pellegata NS. Multiple endocrine neoplasia type 4. Front Horm Res 2013; 41:63-78.

65. Lodish MB, Trivellin G, Stratakis CA. Pituitary gigantism: update on molecular biology and management. Curr Opin Endocrinol Diabetes Obes 2016; 23(1):72-80.

66. Carney JA, Gordon H, Carpenter PC, Shenoy BV, Go VL. The complex of myxomas, spotty pigmentation, and endocrine overactivity. Medicine (Baltimore) 1985; 64(4):270-283.
67. Stratakis CA, Carney JA, Lin JP et al. Carney complex, a familial multiple neoplasia and lentiginosis syndrome. Analysis of 11 kindreds and linkage to the short arm of chromosome 2. J Clin Invest 1996; 97(3):699-705.

68. Kirschner LS, Carney JA, Pack SD et al. Mutations of the gene encoding the protein

kinase A type I-alpha regulatory subunit in patients with the Carney complex. Nat Genet 2000; 26(1):89-92.

69. Sandrini F, Stratakis C. Clinical and molecular genetics of Carney complex. Mol Genet Metab 2003; 78(2):83-92.

70. Stratakis CA, Kirschner LS, Carney JA. Clinical and molecular features of the Carney complex: diagnostic criteria and recommendations for patient evaluation. J Clin Endocrinol Metab 2001; 86(9):4041-4046.

71. Pack SD, Kirschner LS, Pak E, Zhuang Z, Carney JA, Stratakis CA. Genetic and histologic studies of somatomammotropic pituitary tumors in patients with the "complex of spotty skin pigmentation, myxomas, endocrine overactivity and schwannomas" (Carney complex). J Clin Endocrinol Metab 2000; 85(10):3860-3865.

72. Raff SB, Carney JA, Krugman D, Doppman JL, Stratakis CA. Prolactin secretion abnormalities in patients with the "syndrome of spotty skin pigmentation, myxomas, endocrine overactivity and schwannomas" (Carney complex). J Pediatr Endocrinol Metab 2000; 13(4):373-379.

73. O'Toole SM, Denes J, Robledo M, Stratakis CA, Korbonits M. 15 YEARS OF PARAGANGLIOMA: The association of pituitary adenomas and phaeochromocytomas or paragangliomas. Endocr Relat Cancer 2015; 22(4):T105-T122.

74. Denes J, Swords F, Rattenberry E et al. Heterogeneous genetic background of the association of pheochromocytoma/paraganglioma and pituitary adenoma: results from a large patient cohort. J Clin Endocrinol Metab 2015; 100(3):E531-E541.

75. Soares BS, Frohman LA. Isolated familial somatotropinoma. Pituitary 2004; 7(2):95-101.

76. Matsuno A, Teramoto A, Yamada S et al. Gigantism in sibling unrelated to multiple endocrine neoplasia: case report. Neurosurgery 1994; 35(5):952-955.

77. Nozieres C, Berlier P, Dupuis C et al. Sporadic and genetic forms of paediatric somatotropinoma: a retrospective analysis of seven cases and a review of the literature. Orphanet J Rare Dis 2011; 6:67.

78. Gadelha MR, Prezant TR, Une KN et al. Loss of heterozygosity on chromosome 11q13 in two families with acromegaly/gigantism is independent of mutations of the multiple endocrine neoplasia type I gene. J Clin Endocrinol Metab 1999; 84(1):249-256.

79. Jorge BH, Agarwal SK, Lando VS et al. Study of the multiple endocrine neoplasia type 1, growth hormone-releasing hormone receptor, Gs alpha, and Gi2 alpha genes in isolated familial acromegaly. J Clin Endocrinol Metab 2001; 86(2):542-544.

80. Beckers A, Aaltonen LA, Daly AF, Karhu A. Familial isolated pituitary adenomas (FIPA) and the pituitary adenoma predisposition due to mutations in the aryl hydrocarbon receptor interacting protein (AIP) gene. Endocr Rev 2013; 34(2):239-277.

81. Daly AF, Vanbellinghen JF, Khoo SK et al. Aryl hydrocarbon receptor-interacting protein gene mutations in familial isolated pituitary adenomas: analysis in 73 families. J Clin Endocrinol Metab 2007; 92(5):1891-1896.

82. Martucci F, Trivellin G, Korbonits M. Familial isolated pituitary adenomas: an emerging clinical entity. J Endocrinol Invest 2012; 35(11):1003-1014.

83. Chahal HS, Stals K, Unterlander M et al. AIP mutation in pituitary adenomas in the 18th century and today. N Engl J Med 2011; 364(1):43-50.

84. Gadelha MR, Une KN, Rohde K, Vaisman M, Kineman RD, Frohman LA. Isolated familial somatotropinomas: establishment of linkage to chromosome 11q13.1-11q13.3 and evidence for a potential second locus at chromosome 2p16-12. J Clin Endocrinol Metab 2000; 85(2):707-714.

85. Raverot G, Arnous W, Calender A et al. Familial pituitary adenomas with a heterogeneous

functional pattern: clinical and genetic features. J Endocrinol Invest 2007; 30(9):787-790. 86. Cansu GB, Taskiran B, Trivellin G, Faucz FR, Stratakis CA. A novel truncating AIP mutation, p.W279*, in a familial isolated pituitary adenoma (FIPA) kindred. Hormones (Athens) 2016; 15(3):441-444.

87. Trivellin G, Daly AF, Faucz FR et al. Gigantism and acromegaly due to Xq26 microduplications and GPR101 mutation. N Engl J Med 2014; 371(25):2363-2374.
 88. Iacovazzo D, Korbonits M. Gigantism: X-linked acrogigantism and GPR101 mutations. Growth Horm IGF Res 2016; 30-31:64-69.

89. Iacovazzo D, Caswell R, Bunce B et al. Germline or somatic GPR101 duplication leads to X-linked acrogigantism: a clinico-pathological and genetic study. Acta Neuropathol Commun 2016; 4(1):56.

90. Rodd C, Millette M, Iacovazzo D et al. Somatic GPR101 Duplication Causing X-Linked Acrogigantism (XLAG)-Diagnosis and Management. J Clin Endocrinol Metab 2016; 101(5):1927-1930.

91. Daly AF, Yuan B, Fina F et al. Somatic mosaicism underlies X-linked acrogigantism syndrome in sporadic male subjects. Endocr Relat Cancer 2016; 23(4):221-233.

92. Daly AF, Lysy PA, Desfilles C et al. GHRH excess and blockade in X-LAG syndrome. Endocr Relat Cancer 2016; 23(3):161-170.

93. Rostomyan L, Daly AF, Petrossians P et al. Clinical and genetic characterization of pituitary gigantism: an international collaborative study in 208 patients. Endocr Relat Cancer 2015; 22(5):745-757.

94. Barkan AL, Beitins IZ, Kelch RP. Plasma insulin-like growth factor-l/somatomedin-C in acromegaly: correlation with the degree of growth hormone hypersecretion. J Clin Endocrinol Metab 1988; 67(1):69-73.

95. Ali O, Banerjee S, Kelly DF, Lee PD. Management of type 2 diabetes mellitus associated with pituitary gigantism. Pituitary 2007; 10(4):359-364.

96. Alvi NS, Kirk JM. Pituitary gigantism causing diabetic ketoacidosis. J Pediatr Endocrinol Metab 1999; 12(6):907-909.

97. Kuzuya T, Matsuda A, Sakamoto Y, Yamamoto K, Saito T, Yoshida S. A case of pituitary gigantism who had two episodes of diabetic ketoacidosis followed by complete recovery of diabetes. Endocrinol Jpn 1983; 30(3):329-334.

98. Kuo SF, Chuang WY, Ng S et al. Pituitary gigantism presenting with depressive mood disorder and diabetic ketoacidosis in an Asian adolescent. J Pediatr Endocrinol Metab 2013; 26(9-10):945-948.

99. Grellier P, Chanson P, Casadevall N, Abboud S, Schaison G. Remission of polycythemia vera after surgical cure of acromegaly. Ann Intern Med 1996; 124(5):495-496.

100. Eugster EA, Fisch M, Walvoord EC, DiMeglio LA, Pescovitz OH. Low hemoglobin levels in children with in idiopathic growth hormone deficiency. Endocrine 2002; 18(2):135-136.

101. Chapman IM, Hartman ML, Straume M, Johnson ML, Veldhuis JD, Thorner MO. Enhanced sensitivity growth hormone (GH) chemiluminescence assay reveals lower postglucose nadir GH concentrations in men than women. J Clin Endocrinol Metab 1994; 78(6):1312-1319.

102. Melmed S, Jackson I, Kleinberg D, Klibanski A. Current treatment guidelines for acromegaly. J Clin Endocrinol Metab 1998; 83(8):2646-2652.

103. Freda PU, Nuruzzaman AT, Reyes CM, Sundeen RE, Post KD. Significance of "abnormal" nadir growth hormone levels after oral glucose in postoperative patients with acromegaly in remission with normal insulin-like growth factor-I levels. J Clin Endocrinol Metab 2004; 89(2):495-500.

104. Patel YC, Ezzat S, Chik CL et al. Guidelines for the diagnosis and treatment of acromegaly: a Canadian perspective. Clin Invest Med 2000; 23(3):172-187.

105. Bidlingmaier M, Strasburger CJ. Growth hormone assays: current methodologies and their limitations. Pituitary 2007; 10(2):115-119.

106. Le Roith D. Seminars in medicine of the Beth Israel Deaconess Medical Center. Insulinlike growth factors. N Engl J Med 1997; 336(9):633-640.

107. Misra M, Cord J, Prabhakaran R, Miller KK, Klibanski A. Growth hormone suppression after an oral glucose load in children. J Clin Endocrinol Metab 2007; 92(12):4623-4629.
108. Holl RW, Bucher P, Sorgo W, Heinze E, Homoki J, Debatin KM. Suppression of growth

hormone by oral glucose in the evaluation of tall stature. Horm Res 1999; 51(1):20-24. 109. Giustina A, Barkan A, Casanueva FF et al. Criteria for cure of acromegaly: a consensus statement. J Clin Endocrinol Metab 2000; 85(2):526-529.

110. Rajasoorya C, Holdaway IM, Wrightson P, Scott DJ, Ibbertson HK. Determinants of clinical outcome and survival in acromegaly. Clin Endocrinol (Oxf) 1994; 41(1):95-102.

111. Nomikos P, Buchfelder M, Fahlbusch R. The outcome of surgery in 668 patients with acromegaly using current criteria of biochemical 'cure'. Eur J Endocrinol 2005; 152(3):379-387. 112. Gittoes NJ, Sheppard MC, Johnson AP, Stewart PM. Outcome of surgery for acromegaly-the experience of a dedicated pituitary surgeon. QJM 1999; 92(12):741-745.

113. Jane JA, Jr., Starke RM, Elzoghby MA et al. Endoscopic transsphenoidal surgery for acromegaly: remission using modern criteria, complications, and predictors of outcome. J Clin Endocrinol Metab 2011; 96(9):2732-2740.

114. Swearingen B, Barker FG, Katznelson L et al. Long-term mortality after transsphenoidal surgery and adjunctive therapy for acromegaly. J Clin Endocrinol Metab 1998; 83(10):3419-3426.

115. Laws ER, Jr., Thapar K. Pituitary surgery. Endocrinol Metab Clin North Am 1999; 28(1):119-131.

116. Rostomyan L, Daly AF, Petrossians P et al. Clinical and genetic characterization of pituitary gigantism: an international collaborative study in 208 patients. Endocr Relat Cancer 2015; 22(5):745-757.

117. Abe T, Tara LA, Ludecke DK. Growth hormone-secreting pituitary adenomas in childhood and adolescence: features and results of transnasal surgery. Neurosurgery 1999; 45(1):1-10.
118. Williams F, Hunter S, Bradley L et al. Clinical experience in the screening and management of a large kindred with familial isolated pituitary adenoma due to an aryl hydrocarbon receptor interacting protein (AIP) mutation. J Clin Endocrinol Metab 2014; 99(4):1122-1131.

119. Katznelson L, Atkinson JL, Cook DM, Ezzat SZ, Hamrahian AH, Miller KK. American Association of Clinical Endocrinologists medical guidelines for clinical practice for the diagnosis and treatment of acromegaly--2011 update. Endocr Pract 2011; 17 Suppl 4:1-44. 120. Liu F, Li W, Yao Y et al. A case of McCune-Albright syndrome associated with pituitary GH adenoma: therapeutic process and autopsy. J Pediatr Endocrinol Metab 2011; 24(5-6):283-287.

121. Sklar C, Boulad F, Small T, Kernan N. Endocrine complications of pediatric stem cell transplantation. Front Biosci 2001; 6:G17-G22.

122. Attanasio R, Epaminonda P, Motti E et al. Gamma-knife radiosurgery in acromegaly: a 4year follow-up study. J Clin Endocrinol Metab 2003; 88(7):3105-3112.

123. Castinetti F, Taieb D, Kuhn JM et al. Outcome of gamma knife radiosurgery in 82 patients with acromegaly: correlation with initial hypersecretion. J Clin Endocrinol Metab 2005; 90(8):4483-4488.

124. Swords FM, Allan CA, Plowman PN et al. Stereotactic radiosurgery XVI: a treatment for previously irradiated pituitary adenomas. J Clin Endocrinol Metab 2003; 88(11):5334-5340.
125. Grasso LF, Pivonello R, Colao A. Investigational therapies for acromegaly. Expert Opin Investig Drugs 2013; 22(8):955-963.

126. Patel YC, Greenwood M, Panetta R et al. Molecular biology of somatostatin receptor subtypes. Metabolism 1996; 45(8 Suppl 1):31-38.

127. Ezzat S, Snyder PJ, Young WF et al. Octreotide treatment of acromegaly. A randomized, multicenter study. Ann Intern Med 1992; 117(9):711-718.

128. Newman CB, Melmed S, George A et al. Octreotide as primary therapy for acromegaly. J Clin Endocrinol Metab 1998; 83(9):3034-3040.

129. Feuillan PP, Jones J, Ross JL. Growth hormone hypersecretion in a girl with McCune-Albright syndrome: comparison with controls and response to a dose of long-acting somatostatin analog. J Clin Endocrinol Metab 1995; 80(4):1357-1360.

130. Schoof E, Dorr HG, Kiess W et al. Five-year follow-up of a 13-year-old boy with a pituitary adenoma causing gigantism--effect of octreotide therapy. Horm Res 2004; 61(4):184-189.

131. Nanto-Salonen K, Koskinen P, Sonninen P, Toppari J. Suppression of GH secretion in pituitary gigantism by continuous subcutaneous octreotide infusion in a pubertal boy. Acta Paediatr 1999; 88(1):29-33.

132. Freda PU. Somatostatin analogs in acromegaly. J Clin Endocrinol Metab 2002; 87(7):3013-3018.

133. Flogstad AK, Halse J, Bakke S et al. Sandostatin LAR in acromegalic patients: long-term treatment. J Clin Endocrinol Metab 1997; 82(1):23-28.

134. Drange MR, Melmed S. Long-acting lanreotide induces clinical and biochemical remission of acromegaly caused by disseminated growth hormone-releasing hormone-secreting carcinoid. J Clin Endocrinol Metab 1998; 83(9):3104-3109.

135. Ciresi A, Amato MC, Galluzzo A, Giordano C. Complete biochemical control and pituitary adenoma disappearance in a child with gigantism: efficacy of octreotide therapy. J Endocrinol Invest 2011; 34(2):162-163.

136. Tajima T, Tsubaki J, Ishizu K, Jo W, Ishi N, Fujieda K. Case study of a 15-year-old boy with McCune-Albright syndrome combined with pituitary gigantism: effect of octreotide-long acting release (LAR) and cabergoline therapy. Endocr J 2008; 55(3):595-599.

137. Zacharin M. Paediatric management of endocrine complications in McCune-Albright syndrome. J Pediatr Endocrinol Metab 2005; 18(1):33-41.

138. Murray RD, Melmed S. A critical analysis of clinically available somatostatin analog formulations for therapy of acromegaly. J Clin Endocrinol Metab 2008; 93(8):2957-2968.

139. Hofland LJ, van der HJ, van Koetsveld PM et al. The novel somatostatin analog SOM230 is a potent inhibitor of hormone release by growth hormone- and prolactin-secreting pituitary adenomas in vitro. J Clin Endocrinol Metab 2004; 89(4):1577-1585.

140. Schmidt K, Leuschner M, Harris AG et al. Gallstones in acromegalic patients undergoing different treatment regimens. Clin Investig 1992; 70(7):556-559.

141. Trainer PJ, Drake WM, Katznelson L et al. Treatment of acromegaly with the growth hormone-receptor antagonist pegvisomant. N Engl J Med 2000; 342(16):1171-1177.

142. van der Lely AJ, Hutson RK, Trainer PJ et al. Long-term treatment of acromegaly with pegvisomant, a growth hormone receptor antagonist. Lancet 2001; 358(9295):1754-1759. 143. Bernabeu I, Marazuela M, Lucas T et al. Pegvisomant-induced liver injury is related to the UGT1A1*28 polymorphism of Gilbert's syndrome. J Clin Endocrinol Metab 2010; 95(5):2147-2154.

144. Buhk JH, Jung S, Psychogios MN et al. Tumor volume of growth hormone-secreting

pituitary adenomas during treatment with pegvisomant: a prospective multicenter study. J Clin Endocrinol Metab 2010; 95(2):552-558.

145. Higham CE, Atkinson AB, Aylwin S et al. Effective combination treatment with cabergoline and low-dose pegvisomant in active acromegaly: a prospective clinical trial. J Clin Endocrinol Metab 2012; 97(4):1187-1193.

146. van der Lely AJ, Biller BM, Brue T et al. Long-term safety of pegvisomant in patients with acromegaly: comprehensive review of 1288 subjects in ACROSTUDY. J Clin Endocrinol Metab 2012; 97(5):1589-1597.

147. Mangupli R, Rostomyan L, Castermans E et al. Combined treatment with octreotide LAR and pegvisomant in patients with pituitary gigantism: clinical evaluation and genetic screening. Pituitary 2016; 19(5):507-514.

148. Main KM, Sehested A, Feldt-Rasmussen U. Pegvisomant treatment in a 4-year-old girl with neurofibromatosis type 1. Horm Res 2006; 65(1):1-5.

149. Rix M, Laurberg P, Hoejberg AS, Brock-Jacobsen B. Pegvisomant therapy in pituitary gigantism: successful treatment in a 12-year-old girl. Eur J Endocrinol 2005; 153(2):195-201.
150. Bergamaschi S, Ronchi CL, Giavoli C et al. Eight-year follow-up of a child with a GH/prolactin-secreting adenoma: efficacy of pegvisomant therapy. Horm Res Paediatr 2010; 73(1):74-79.

151. Daniel A, d'Emden M, Duncan E. Pituitary gigantism treated successfully with the growth hormone receptor antagonist, pegvisomant. Intern Med J 2013; 43(3):345-347.

152. Goldenberg N, Racine MS, Thomas P, Degnan B, Chandler W, Barkan A. Treatment of pituitary gigantism with the growth hormone receptor antagonist pegvisomant. J Clin Endocrinol Metab 2008; 93(8):2953-2956.

153. Drop SL, De Waal WJ, De Muinck Keizer-Schrama SM. Sex steroid treatment of constitutionally tall stature. Endocr Rev 1998; 19(5):540-558.

154. Lu PW, Silink M, Johnston I, Cowell CT, Jimenez M. Pituitary gigantism. Arch Dis Child 1992; 67(8):1039-1041.

155. Minagawa M, Yasuda T, Someya T, Kohno Y, Saeki N, Hashimoto Y. Effects of octreotide infusion, surgery and estrogen on suppression of height increase and 20K growth hormone ratio in a girl with gigantism due to a growth hormone-secreting macroadenoma. Horm Res 2000; 53(3):157-160.

156. Albuquerque EV, Scalco RC, Jorge AA. MANAGEMENT OF ENDOCRINE DISEASE:
Diagnostic and therapeutic approach of tall stature. Eur J Endocrinol 2017; 176(6):R339-R353.
157. Hendriks AE, Drop SL, Laven JS, Boot AM. Fertility of tall girls treated with high-dose estrogen, a dose-response relationship. J Clin Endocrinol Metab 2012; 97(9):3107-3114.
158. Baujat G, Rio M, Rossignol S et al. Clinical and molecular overlap in overgrowth syndromes. Am J Med Genet C Semin Med Genet 2005; 137(1):4-11.

159. Sotos JF, DODGE PR, MUIRHEAD D, CRAWFORD JD, TALBOT NB. CEREBRAL GIGANTISM IN CHILDHOOD. A SYNDROME OF EXCESSIVELY RAPID GROWTH AND ACROMEGALIC FEATURES AND A NONPROGRESSIVE NEUROLOGIC DISORDER. N Engl J Med 1964; 271:109-116.

160. Cole TR, Hughes HE. Sotos syndrome: a study of the diagnostic criteria and natural history. J Med Genet 1994; 31(1):20-32.

161. Agwu JC, Shaw NJ, Kirk J, Chapman S, Ravine D, Cole TR. Growth in Sotos syndrome. Arch Dis Child 1999; 80(4):339-342.

162. Sotos JF. Overgrowth. Section VI. Genetic syndromes and other disorders associated with overgrowth. Clin Pediatr (Phila) 1997; 36(3):157-170.

163. Raitz R, Laragnoit A. Supernumerary teeth and dental management in Sotos syndrome. J

Dent Child (Chic) 2009; 76(3):246-250.

164. Ball LJ, Sullivan MD, Dulany S, Stading K, Schaefer GB. Speech-language characteristics of children with Sotos syndrome. Am J Med Genet A 2005; 136(4):363-367.
165. Lane C, Milne E, Freeth M. Cognition and Behaviour in Sotos Syndrome: A Systematic Review. PLoS One 2016; 11(2):e0149189.

166. Mauceri L, Sorge G, Baieli S, Rizzo R, Pavone L, Coleman M. Aggressive behavior in patients with Sotos syndrome. Pediatr Neurol 2000; 22(1):64-67.

167. van Haelst MM, Hoogeboom JJ, Baujat G et al. Familial gigantism caused by an NSD1 mutation. Am J Med Genet A 2005; 139(1):40-44.

168. Brown WT, Wisniewski KE, Sudhalter V et al. Identical twins discordant for Sotos syndrome. Am J Med Genet 1998; 79(4):329-333.

169. Malan V, de Blois MC, Prieur M et al. Sotos syndrome caused by a paracentric inversion disrupting the NSD1 gene. Clin Genet 2008; 73(1):89-91.

170. Melchior L, Schwartz M, Duno M. dHPLC screening of the NSD1 gene identifies nine novel mutations--summary of the first 100 Sotos syndrome mutations. Ann Hum Genet 2005; 69(Pt 2):222-226.

171. Saugier-Veber P, Bonnet C, Afenjar A et al. Heterogeneity of NSD1 alterations in 116 patients with Sotos syndrome. Hum Mutat 2007; 28(11):1098-1107.

172. Sotos JF. Sotos syndrome 1 and 2. Pediatr Endocrinol Rev 2014; 12(1):2-16.

173. Cecconi M, Forzano F, Milani D et al. Mutation analysis of the NSD1 gene in a group of 59 patients with congenital overgrowth. Am J Med Genet A 2005; 134(3):247-253.

174. Tatton-Brown K, Douglas J, Coleman K et al. Genotype-phenotype associations in Sotos syndrome: an analysis of 266 individuals with NSD1 aberrations. Am J Hum Genet 2005; 77(2):193-204.

175. Beckwith J. Macroglossia, omphalocete, adrenal cytomegaly:fifantism and hyperplastic visceromegaly. Birth Defects 1969; 5:188-196.

176. WIEDEMANN HR. [FAMILIAL MALFORMATION COMPLEX WITH UMBILICAL HERNIA AND MACROGLOSSIA--A "NEW SYNDROME"?]. J Genet Hum 1964; 13:223-232.

177. Pettenati MJ, Haines JL, Higgins RR, Wappner RS, Palmer CG, Weaver DD.

Wiedemann-Beckwith syndrome: presentation of clinical and cytogenetic data on 22 new cases and review of the literature. Hum Genet 1986; 74(2):143-154.

178. Sathienkijkanchai A, Prucka SK, Grant JH, Robin NH. Isolated facial hemihyperplasia: manifestation of Beckwith-Wiedemann syndrome. J Craniofac Surg 2008; 19(1):279-283.

179. Wangler MF, An P, Feinberg AP, Province M, Debaun MR. Inheritance pattern of Beckwith-Wiedemann syndrome is heterogeneous in 291 families with an affected proband. Am J Med Genet A 2005; 137(1):16-21.

180. Rump P, Zeegers MP, van Essen AJ. Tumor risk in Beckwith-Wiedemann syndrome: A review and meta-analysis. Am J Med Genet A 2005; 136(1):95-104.

181. Weksberg R, Shuman C, Beckwith JB. Beckwith-Wiedemann syndrome. Eur J Hum Genet 2010; 18(1):8-14.

182. Debaun MR, Tucker MA. Risk of cancer during the first four years of life in children from The Beckwith-Wiedemann Syndrome Registry. J Pediatr 1998; 132(3 Pt 1):398-400.

183. Cerrato F, Sparago A, Verde G et al. Different mechanisms cause imprinting defects at the IGF2/H19 locus in Beckwith-Wiedemann syndrome and Wilms' tumour. Hum Mol Genet 2008; 17(10):1427-1435.

184. Cohen MM, Jr. Beckwith-Wiedemann syndrome: historical, clinicopathological, and etiopathogenetic perspectives. Pediatr Dev Pathol 2005; 8(3):287-304.

185. Slavotinek A, Gaunt L, Donnai D. Paternally inherited duplications of 11p15.5 and

Beckwith-Wiedemann syndrome. J Med Genet 1997; 34(10):819-826.

186. Brioude F, Lacoste A, Netchine I et al. Beckwith-Wiedemann syndrome: growth pattern and tumor risk according to molecular mechanism, and guidelines for tumor surveillance. Horm Res Paediatr 2013; 80(6):457-465.

187. Cooper WN, Luharia A, Evans GA et al. Molecular subtypes and phenotypic expression of Beckwith-Wiedemann syndrome. Eur J Hum Genet 2005; 13(9):1025-1032.

188. Uyar A, Seli E. The impact of assisted reproductive technologies on genomic imprinting and imprinting disorders. Curr Opin Obstet Gynecol 2014; 26(3):210-221.

189. Bowdin S, Allen C, Kirby G et al. A survey of assisted reproductive technology births and imprinting disorders. Hum Reprod 2007; 22(12):3237-3240.

190. Zimmermann N, Stanek J. Perinatal Case of Fatal Simpson-Golabi-Behmel Syndrome with Hyperplasia of Seminiferous Tubules. Am J Case Rep 2017; 18:649-655.

191. Lin AE, Neri G, Hughes-Benzie R, Weksberg R. Cardiac anomalies in the Simpson-Golabi-Behmel syndrome. Am J Med Genet 1999; 83(5):378-381.

192. Cottereau E, Mortemousque I, Moizard MP et al. Phenotypic spectrum of Simpson-Golabi-Behmel syndrome in a series of 42 cases with a mutation in GPC3 and review of the literature. Am J Med Genet C Semin Med Genet 2013; 163C(2):92-105.

193. Pilia G, Hughes-Benzie RM, MacKenzie A et al. Mutations in GPC3, a glypican gene, cause the Simpson-Golabi-Behmel overgrowth syndrome. Nat Genet 1996; 12(3):241-247. 194. Debaun MR, Ess J, Saunders S. Simpson Golabi Behmel syndrome: progress toward understanding the molecular basis for overgrowth, malformation, and cancer predisposition. Mol Genet Metab 2001; 72(4):279-286.

195. Mariani S, lughetti L, Bertorelli R et al. Genotype/phenotype correlations of males affected by Simpson-Golabi-Behmel syndrome with GPC3 gene mutations: patient report and review of the literature. J Pediatr Endocrinol Metab 2003; 16(2):225-232.

196. Davoodi J, Kelly J, Gendron NH, MacKenzie AE. The Simpson-Golabi-Behmel syndrome causative glypican-3, binds to and inhibits the dipeptidyl peptidase activity of CD26. Proteomics 2007; 7(13):2300-2310.

197. Li M, Shuman C, Fei YL et al. GPC3 mutation analysis in a spectrum of patients with overgrowth expands the phenotype of Simpson-Golabi-Behmel syndrome. Am J Med Genet 2001; 102(2):161-168.

198. Kehrer C, Hoischen A, Menkhaus R et al. Whole exome sequencing and array-based molecular karyotyping as aids to prenatal diagnosis in fetuses with suspected Simpson-Golabi-Behmel syndrome. Prenat Diagn 2016; 36(10):961-965.

199. Weaver DD, Graham CB, Thomas IT, Smith DW. A new overgrowth syndrome with accelerated skeletal maturation, unusual facies, and camptodactyly. J Pediatr 1974; 84(4):547-552.

200. Kelly TE, Alford BA, Abel M. Cervical spine anomalies and tumors in Weaver syndrome. Am J Med Genet 2000; 95(5):492-495.

201. Proud VK, Braddock SR, Cook L, Weaver DD. Weaver syndrome: autosomal dominant inheritance of the disorder. Am J Med Genet 1998; 79(4):305-310.

202. Cooney E, Bi W, Schlesinger AE, Vinson S, Potocki L. Novel EED mutation in patient with Weaver syndrome. Am J Med Genet A 2017; 173(2):541-545.

203. Imagawa E, Higashimoto K, Sakai Y et al. Mutations in genes encoding polycomb repressive complex 2 subunits cause Weaver syndrome. Hum Mutat 2017; 38(6):637-648. 204. Neylon OM, Werther GA, Sabin MA. Overgrowth syndromes. Curr Opin Pediatr 2012; 24(4):505-511.