PITUITARY HISTOPATHOLOGY IN MAN: NORMAL AND ABNORMAL

Updated: June 10, 2007

Authors: Sylvia L. Asa, M.D., Ph.D.

THE NORMAL PITUITARY GLAND

The pituitary is a bean-shaped gland located at the base of the brain in the midline. It measures 0.6 cm SI x 0.9 cm AP x 1.3 cm and an average gland weighs 0.6 grams. Females tend to have larger glands, especially during or after pregnancy, with weights up to 1 g (1;2). The gland lies within the bony sella turcica that surrounds it inferiorly and laterally. Superiorly it is covered by the diaphragma sella, a reflection of the dura mater (3;4). Lateral to the sella are the cavernous sinuses; anteroinferior is the sphenoid sinus; anterosuperior is the optic chiasm; superior to it is the hypothalamus. The pituitary is composed of two anatomically and functionally distinct parts: the neurohypophysis and the adenohypophysis. (Figure 1)



Figure 1. (a) The normal pituitary gland is a bean-shaped structure that hangs from the pituitary stalk (arrow), an extension of the hypothalamus that traverses the diaphragm of the sella turcica. (b) A horizontal section of the gland identifies the anterior lobe (A) or

"adenohypophysis", the posterior lobe (P) or "neurohypophysis" and the vestigial intermediate lobe (I) that is poorly developed in humans. This gland demonstrates "basophil invasion" of the posterior lobe, proliferation of corticotrophs into the neurohypophysis (arrows) that is a normal feature in older patients.

The neurohypophysis is composed of the infundibulum, the pituitary stalk, and the pars nervosa of the pituitary. The cell types of the neurohypophysis include pituicytes, which are modified glial cells, and the axonal processes of neurons whose cell bodies are located in the hypothalamus. The neurohypophysis stores and releases the hypothalamic hormones oxytocin and vasopressin (Figure 2).



Figure 2. (a) The neurohypophysis is composed of "pituicytes", modified glial cells, that surround axonal processes of neurons whose cell bodies are located in the hypothalamus. Occasional dilated terminals of those axons are seen as "Herring bodies" (arrows). (b) The neurohypophysis stores and releases the hypothalamic hormones oxytocin and vasopressin in Herring bodies that are best identified with immunostains for the hormones, as seen here stained for vasopressin (arrows).

The adenohypophysis is of endodermal origin, embryologically derived from Rathke's pouch (5). It has three regions, the pars distalis or anterior lobe, the pars intermedia or intermediate lobe and the pars tuberalis, an extension of epithelium that wraps around the infundibulum of the pituitary stalk. Adenohypophyseal development and cytodifferentiation are regulated by highly specific transcription factors (6-8). (Figure 3) Many of these are implicated in early pituitary organogenesis, including the Rathke's pouch homeobox (Rpx) protein which is identified in the pituitary primordium prior to the onset of known pituitary hormone production, and Pax-6. Mutations of the former have been associated with septo-optic dysplasia and deficiency of the latter is associated with the small-eye mouse phenotype. The bicoid-related pituitary homeobox

factor Ptx1 and structurally related pituitary homeobox factor 2 (Ptx2) are also required for pituitary development. Two members of the Lhx gene family, a group of LIM homeobox genes, Lhx3 and Lhx4 and P-LIM, another LIM homeobox protein transcription factor, are expressed in the pituitary with highest levels at the early stages of Rathke's pouch development. Loss of both Lhx-3 and Lhx-4 results in pituitary hypoplasia but deficiency of Lhx-4 only results in loss of pituitary gonadotrophs alone. Another early determinant of pituitary differentiation is the Prophet of Pit-1 (PROP-1) is a paired-like homeodomain protein that is expressed early in pituitary development. It induces expression of the next phase of development directed by the pituitary transcription factor Pit-1, and plays a role in downregulation of Rpx. Inactivating mutations of PROP-1 have been identified as the cause of Pit-1 deficiency in Ames dwarf mice and of combined pituitary hormone deficiency in humans. Id, a member of the helix-loop-helix (HLH) family of transcription factors, IsI-1, a Lim factor, and several other transcription factors are also found early in development and have been implicated in developmental anomalies.



Figure 3. Pathways of pituitary development and cytodifferentiation. Transcription factors implicated in each step are identified. (see text for details)

The adenohypophysis is composed of acini that contain the specialized cell types, all of which have their own unique hormonal function and characteristics. The molecular factors that determine hormone production are transcription factors that target specific hormone genes. These factors have clarified three main pathways of cell differentiation (6;7) (Figure 3).

Somatotrophs, lactotrophs, mammosomatotrophs, and thyrotrophs all derive from growth hormone (GH)-producing precursors that express the transcription factor Pit-1 (9). (10-12)The

expression of estrogen receptor (ER)a enhances prolactin (PRL) secretion (13), allowing mammosomatotroph differentiation, and a silencing mechanism is thought to repress GH production to allow mature lactotrophs to develop. Thyrotroph embryonic factor (TEF) is the putative factor required for thyrotrophin (TSH)-beta production (14), and GATA-2 appears to be an important contributor to thyrotroph development (15). Mature thyrotrophs also suppress GH production(8). This family of cells is thought to maintain fluidity so that in various situations, there is transdifferentiation: somatotrophs convert to mammosomatotrophs and lactotrophs during pregnancy, and to thyrotrophs in hypothyroidism, and these are thought to be reversible transdifferentiation processes.

GH-producing somatotrophs are located in the lateral wings of the anterior pituitary and account for approximately 50% of the cell population. By light microscopy they are strongly acidophilic cells with centrally located nuclei and diffuse cytoplasmic positivity for GH (Figure 4). Electron microscopy reveals abundant rough endoplasmic reticulum, well-formed Golgi complexes and numerous large dense secretory granules. PRL-secreting lactotrophs are scattered randomly throughout the adenohypophysis; however, they can most often be found in the posterolateral aspects of the gland. In males and nulliparous females, they constitute approximately 9% of the cell population; in multiparous females, they can represent up to 31% of the adenohypophysial cells. They are usually sparsely granulated and chromophobic polygonal cells that wrap cell processes around adjacent cells (Figure 5), usually gonadotrophs; some are densely granulated and acidophilic. The ultrastructural hallmarks of lactotrophs are the elaborate rough endoplasmic reticulum arranged in parallel arrays and occasionally forming concentric structures known as "Nebenkern" formations, the prominent Golgi complexes, and the extrusion of secretory granules at the lateral cell border, a phenomenon known as "misplaced exocytosis". Mammosomatotrophs, which produce both GH and PRL, resemble somatotrophs but contain both GH and PRL by immunohistochemistry. The often have irregular, elongated and pleomorphic large granules and they exhibit the hallmark of PRL secretion, misplaced exocytosis. Thyrotrophs, which produce TSH, represent approximately 5% of adenohypophysial cells and are most numerous in the anteromedial aspect of the gland. They are angular chromophobic cells with multiple elongated cytoplasmic processes (Figure 6). They exhibit cytoplasmic immunoreactivity for alpha-subunit and beta-TSH. By electron microscopy they are characterized by short profiles of dilated rough endoplasmic reticulum and small secretory granules that align along the plasma membrane.



Figure 4. Somatotrophs exhibit diffuse cytoplasmic positivity for GH. They are most numerous in the lateral wings of the anterior pituitary (a) but are scattered throughout the gland including the median wedge (b).



Figure 5. Lactotrophs are polygonal cells that wrap cell processes around adjacent cells, usually gonadotrophs; there is some variation in the intensity of staining for PRL.



Figure 6. Thyrotrophs are angular chromophobic cells with multiple elongated cytoplasmic processes that are well seen on immunostaining for β -TSH.

Corticotrophs, which produce proopiomelanocortin (POMC) and its derivatives including adrenocorticotropin (ACTH), melanotropin (MSH) and lipotropin (LPH), represent approximately 15 to 20 percent of adenohypophysial cells. These basophilic cells (Figure 7a) have strong positivity using the periodic acid-Schiff (PAS) stain. They are concentrated mainly in the central region of the gland known as the "mucoid wedge" because of the PAS-reactivity, and are the main components of the vestigial intermediate lobe that in animals is the source of MSH (Figure 7b). In older patients they spill into the posterior lobe, a phenomenon known as "basophil invasion" (Figure 1b). They have characteristic "enigmatic bodies", large cytoplasmic vacuoles that represent complex lysosomes (Figure 7c). The secretory granules are marked by pleomorphism of size, shape and electron density, and it is not unusual to identify indentations and evaginations of granule membranes, resulting in "heart" and "teardrop" shapes. Small bundles of intermediate filaments identified throughout the cytoplasm by electron microscopy represent keratin filaments. When corticotrophs are exposed to excess glucocorticoids, they undergo a reversible morphologic modification known as Crooke's hyaline change (Figure 8) attributable to concentric accumulations of intermediate filaments that displace the PAS-

positive, ACTH-immunoreactive secretory granules to the juxtanuclear and peripheral cytoplasm. By light microscopy, the accumulated filaments have a pale, homogenous glassy appearance and they stain for low molecular weight cytokeratins. Corticotrophs of the pars intermedia are thought to cleave POMC differently from the ACTH-producing cells of the pars distalis, and in situations of glucocorticoid excess, they do not undergo Crooke's hyaline change. The expression of proopiomelanocortin that defines corticotrophs is dependent on the T-box transcription factor Tpit, (16). Tpit interacts with Ptx1 and Corticotropin upstream transcription-binding element (CUTE) proteins, including neuroD1/beta 2 (17;18).



Figure 7. (a) Corticotrophs are round basophilic cells that are readily identified on H&E-stained sections. (b) They are concentrated mainly in the central region of the gland known as the "mucoid wedge" and are the main components of the vestigial intermediate lobe that is composed of small cysts (C). (c) They have characteristic "enigmatic bodies", large cytoplasmic vacuoles that represent complex lysosomes (arrows).



Figure 8. In situations of glucocorticoid excess, human corticotrophs (arrows) undergo accumulation of keratin filaments in the cytoplasm, resulting in a glassy hyaline appearance; the PAS-positive secretory material is trapped in the juxtanuclear region or at the plasma membrane. This is known as 'Crooke's hyaline change'

Expression of steroidogenic factor -1 (SF-1) (19) and GATA-2 (15) are required for gonadotroph differentiation. Gonadotrophs, which produce the two gonadotropins, follicle-stimulating hormone (FSH) and luteinizing hormone (LH), account for 10% of adenohypophysial cells. They are scattered throughout the pars distalis (Figure 9) and pars tuberalis. With increasing age, these cells tend to undergo oncocytic and squamous metaplasia. These cells are reliably identified by immunoreactivity for beta-subunits of FSH and/or LH, but their ultrastructural appearance is not as characteristic as that of other adenohypophysial cells. They are oval with eccentric spherical nuclei, and their cytoplasm contains short profiles of dilated rough endoplasmic reticulum containing flocculent electron dense material. Secretory granules are generally sparse, small and scattered throughout the cytoplasm.



Figure 9. Gonadotrophs are scattered round to oval cells that contain strong cytoplasmic reactivity for α -subunit, β -FSH and β -LH (β -FSH shown).

The adenohypophysis includes other cells that are not hormonally active. Follicular cells form around small follicles; they are thought to derive from hormone-secreting cells in response to trauma, compression or degeneration. Folliculostellate cells are stromal sustentacular cells that surround acini of the normal gland; they are immunoreactive for S100 protein (Figure 10) or glial fibrillary acidic protein (GFAP). Scattered cells in the gland that have features of adenohypophysial hormone-secreting cells but cannot be classified are called null cells, and the normal gland also contains occasional oncocytes. These latter are characterized by accumulations of dilated, spherulated mitochondria; oncocytic change is most common in gonadotrophs and occurs rarely in other adenohypophysial cells.



Figure 10. Folliculostellate cells are stromal sustentacular cells that surround acini of the normal gland; they are immunoreactive for S100 protein.

The pars intermedia is poorly developed in the human and is composed of small cystic spaces (Figure 1b, Figure 7b) lined by hormone-containing cells types, predominantly corticotrophs. The pars tuberalis is the superior portion of the adenohypophysis that wraps itself around the neural stalk. It is composed primarily of gonadotrophs that with age, undergo squamous metaplasia (20).

The pituitary receives its vascular supply from the superior, middle, and inferior hypophyseal arteries, all of which originate from the internal carotid arteries. The superior hypophysial arteries flow through the infundibulum of the neurohypophysis and form the portal vessels that transport regulatory hormones from the hypothalamus to the pituitary gland. The middle hypophyseal arteries supply blood directly to the adenohypophysis, while the inferior hypophyseal arteries supply the pars nervosa (21-23). Venous blood from the pituitary gland drains mainly into the internal jugular veins, however, there is evidence that reverse flow in the short portal vessels allows adenohypophysial secretion to affect neurohypophysial and hypothalamic function (24;25).

CONGENITAL AND DEVELOPMENTAL DISORDERS

ECTOPIC ADENOHYPOPHYSIS

Residual adenohypophysial tissue can be trapped anywhere along the path followed by Rathke's pouch during fetal development (26-28). The sphenoid sinus is the most common site of ectopic pituitary, followed by the immediate suprasellar region (29;30); these ectopic foci may be incidental findings, or they can undergo hyperplastic or neoplastic change.

PITUITARY APLASIA/HYPOPLASIA

These disorders are usually associated with severe congenital malformations such as those seen in the Cornelia de Lange syndrome(31)or Arnold-Chiari malformation(32). They result in hypopituitarism with subsequent thyroid and adrenal hypoplasia or aplasia(33-37). At least one form of this disorder associated with septo-optic dysplasia has been attributed to a mutation of the Rpx-1 gene (also known as Hesx-1)(38).

Inactivating mutations of PROP-1 result in combined pituitary hormone deficiency (39;40). Pituitary dwarfism with hypothyroidism occurs in patients with mutations of the pit-1 gene (41-43), likely due to hypoplasia of somatotrophs, lactotrophs and thyrotrophs (44).

PITUITARY DUPLICATION OR DYSTOPIA

These lesions are also usually associated with other congenital malformations (45;46).

EMPTY SELLA SYNDROME

A defective or absent diaphragma sella results in transmission of the pulsatile pressure waves of CSF pressure that causes sellar enlargement and flattens the pituitary against the floor of the sella turcica (47). Pituitary function is usually unaffected (48). However, some patients have hyperprolactinemia, attributed to distortion of the pituitary stalk and interference with the tonic dopaminergic inhibition of prolactin (49). This can be difficult to distinguish from prolactinoma that can also develop in the setting of empty sella.

CYSTIC LESIONS

RATHKE'S CLEFT CYSTS

Remnants of Rathke's pouch commonly form small cysts (<5mm) in the vestigial pars intermedia. These small cysts occasionally enlarge into true Rathke's cleft cysts, lined by ciliated cuboidal or columnar epithelium with occasional goblet cells (Figure 11) and areas of squamous metaplasia; as they enlarge they become symptomatic. Although this condition may occasionally be seen in children, it is most common in adults.



Figure 11. Rathke's cleft cysts are lined by ciliated cuboidal or columnar epithelium (a) with occasional goblet cells that stain with the PAS technique (b).

These cysts are non-functional, but may cause hypopituitarism or diabetes insipidus by compression of surrounding structures (50). Suprasellar extension may give rise to mass effects such as visual field defects and headaches. Severe cases can lead to hydrocephalus, aseptic meningitis, and rarely, abscess formation (51). CT scans usually reveal low-density cystic areas with peripheral enhancement; MRI findings tend to be more variable (52).

Treatment by drainage with or without surgical excision usually results in resolution of mass effects; however, hypopituitarism and diabetes insipidus may persist, requiring permanent hormone replacement. Recurrence rates are low.

ARACHNOID CYSTS

Arachnoid cysts originate in the arachnoid of the sellar and parasellar areas; they may be congenital or acquired. They contain clear fluid and the cyst wall is arachnoid laminar connective tissue with incomplete simple flattened epithelium.

Clinically, they can present with mass effects from suprasellar extension, or hypopituitarism and/or diabetes insipidus due to pituitary compression (50). Treatment involves drainage of cyst contents with partial surgical excision.

DERMOID AND EPIDERMOID CYSTS

Dermoid and epidermoid cysts (also known as "cholesteatomas") originate from ectopic or traumatically implanted epithelial cells. Epidermoid cysts have a lining composed of keratinizing squamous epithelium; the lining of dermoid cysts contains skin appendages such as hair follicles and sweat glands. In addition to the sellar and suprasellar regions, these cysts are also

found intracranially, most often at the cerebellopontine angle (53).

Clinical manifestations include hypopituitarism, hyperprolactinemia due to stalk compression, visual field defects, and a variety of nonspecific neurologic symptoms. Radiologic evaluation reveals a cystic lesion.

Surgical resection is usually curative. Complications include rupture of the cyst with subsequent meningitis, or the development of squamous cell carcinoma (54;55).

INFLAMMATORY DISORDERS

Inflammatory conditions can cause mass effects and/or hypothalamic-hypophysial dysfunction. Primary or idiopathic inflammatory conditions include lymphocytic hypophysitis, granulomatous hypophysitis, and xanthomatous hypophysitis(56). Secondary inflammatory conditions affecting the pituitary gland include infections and various systemic diseases.

LYMPHOCYTIC HYPOPHYSITIS

This chronic inflammatory condition of the pituitary gland is seen most commonly in young postpartum or pregnant females (57). The disorder is much less common in males; the female to male ratio is 8.5: 1. The mean ages of presentation are 34.5 years in females and 44.7 years in males. An autoimmune etiology has been proposed as the basis for lymphocytic hypophysitis due to its association with a number of other autoimmune endocrine disorders such as thyroiditis, adrenalitis, atrophic gastritis, and lymphocytic parathyroiditis and there is evidence for pituitary antibodies in patients with this disease (58). Recent data suggest that the precipitating antigen is alpha-enolase, a protein that is expressed by the placenta as well as pituitary, possibly explaining pregnancy as an initiating event.

The symptoms and signs of lymphocytic hypophysitis tend to be nonspecific, mimicking adenoma. The most common manifestation is mild to moderate hyperprolactinemia. Occasionally patients have isolated ACTH deficiency; other hormone deficiencies are rare. In addition, it can present with mass effects such as headache and visual field deficits. Rarely, patients present with isolated diabetes insipidus and the inflammatory process is restricted to the posterior lobe and stalk, which can exhibit localized enlargement; this disorder has been named infundibular neurohypophysitis (59-61).

Radiologic findings can mimic features of an adenoma; the gland is enlarged and may even exhibit suprasellar extension. However, careful MRI examination with contrast enhancement documents diffuse involvement of the gland without discrete delineation and lacking enhancement in normal gland.

At surgery the gland is inflamed, enlarged and soft or may appear atrophic and fibrotic. Microscopic examination reveals a diffuse inflammatory infiltrate composed mainly of lymphocytes and plasma cells forming occasional lymphoid follicles (Figure 12). There is destruction of the adenohypophysial tissue; the remaining parenchymal cells exhibit variable oncocytic change. Rarely, there seems to be preferential destruction of one hormone-containing cell type. The extent of fibrosis varies with the duration of the disease.



Figure 12. Lymphocytic Hypophysitis is characterized by a diffuse inflammatory infiltrate composed mainly of lymphocytes and plasma cells forming occasional lymphoid follicles. The inflammatory cells surround and destroy parenchymal cells that occasionally show oncocytic change.

The natural history of untreated lymphocytic hypophysitis is variable; it may result in permanent hypopituitarism due to extensive destruction of adenohypophysial cells, or it may run a self-limited course followed by a full recovery. Treatment for this condition is supportive with appropriate hormone replacement. Corticosteroids have been proposed to decrease inflammation, but the efficacy of this treatment has yet to be determined. Transsphenoidal surgery should be considered if the patient suffers progressive mass effects, or deterioration as evidenced by radiologic or neurologic changes. Surgical biopsy may be required to establish the diagnosis. It should be noted however, that surgery has resulted in deleterious effects in occasional case reports.

GRANULOMATOUS HYPOPHYSITIS

Idiopathic granulomatous hypophysitis is a rare chronic inflammatory disorder of unknown pathogenesis first described in 1917 (62-65). It represents one percent of all pituitary disorders with an annual incidence of 1 in 10 million. As of 1991, only 31 cases were described in publications, 21 from autopsy material. Unlike lymphocytic hypophysitis, there is no gender predilection. The mean age of presentation in females is 21.5 years; in males it is 50 years.

Patients may present with visual field deficits, cranial nerve palsies, or headaches, which may be accompanied by nausea and vomiting; this is in contrast to headaches caused by adenomas that are not associated with nausea and vomiting (66;67). Other clinical manifestations include variable degrees of adenohypophysial failure (68), hyperprolactinemia (69), diabetes insipidus, and meningitis with CSF leucocytosis (70).

Radiologic evaluation usually reveals an intrasellar mass with or without suprasellar extension (67). Sometimes, a tongue-like extension along the basal hypothalamus can be seen. Microscopically, this condition is characterized by collections of histiocytes with scattered lymphocytes and plasma cells; multinucleated giant cells may be present (Figure 13). By definition, a diagnosis of granulomatous hypophysitis cannot be made until systemic granulomatous disease has been excluded.



Figure 13. Granulomatous hypophysitis is characterized by collections of histiocytes with scattered lymphocytes and plasma cells; occasional giant cells and areas of necrosis mey be seen but are not identified in this area.

Treatment is somewhat controversial; transphenoidal biopsy/resection with subsequent administration of corticosteroids has been proposed (56;71).

XANTHOMATOUS HYPOPHYSITIS

This relatively new clinicopathologic entity is characterized by a chronic inflammatory infiltrate composed mainly of foamy histiocytes with scattered lymphocytes and plasma cells. The patients reported have been young females. Clinical presentation included headache, nausea, menstrual irregularity, and diabetes insipidus. One patient has had elevated prolactin levels. In most patients, a preoperative diagnosis of pituitary adenoma was suspected based on the presence of a localized lesion in the pituitary (56;72). Histologically, the condition is characterized by infiltration of the adenohypophysis by foamy lipid-containing histiocytes with areas of granulation tissue. This disorder may represent a reactive process (56), however, it remains idiopathic. A diagnosis of xanthomatous hypophysitis can only be made after causes of secondary hypophysitis are ruled out.

SECONDARY HYPOPHYSITIS

These are inflammatory lesions of the pituitary gland, which have a definite etiology (73) or occur as part of a systemic process. A number of infectious agents can involve the pituitary, including fungi, mycobacteria, brucellosis, and syphilis (74). They can result in acute or chronic hypophysitis with occasional abscess formation. Other causes of secondary hypophysitis include sarcoidosis (75), vasculitides such as Takayasu's Disease (76) and Wegener's granulomatosis (77), Crohn's disease (78), Whipple's disease, ruptured Rathke's cleft cyst (79;80), necrotizing adenoma (73), and meningitis (70).

The acquired immunodeficiency syndrome (AIDS) may involve the pituitary gland (81). The involvement is usually infectious in nature and results in acute or chronic inflammation with necrosis. The pathogens are generally opportunistic organisms including PCP, toxoplasma gondii, and CMV.

VASCULAR DISORDERS

PITUITARY INFARCTION

Ischemic necrosis of the pituitary can result from a number of insults including head injury, hemorrhagic shock, disseminated intravascular coagulation, thrombocytopenia, and stroke

(82-85). Two unique conditions that result in pituitary infarction are Sheehan's syndrome and pituitary apoplexy.

SHEEHAN'S SYNDROME

Sheehan's syndrome is postpartum necrosis of the pituitary gland. This disorder is usually related to hypotension caused by postpartum hemorrhage (86). There is usually extensive central necrosis of the gland with a rim of viable cells at the periphery, but various degrees of necrosis can occur. Clinical manifestations appear only with significant tissue destruction. The neurohypophysis, with its independent blood supply, is usually spared.

PITUITARY APOPLEXY

This condition constitutes a true endocrine emergency in which acute hemorrhagic infarction of a sellar tumor (usually an adenoma), results in rapid expansion with symptoms and signs of elevated intracranial pressure (87). Factors predisposing to pituitary apoplexy include carotid angiography, radiation therapy, trauma, coagulopathy, temporal arteritis, diabetes mellitus, and atherosclerosis but none of these may be present.

Histologic examination reveals extensive infarction and hemorrhage. These features are common as focal changes in many pituitary tumors, however, true pituitary apoplexy refers to those extreme cases where hemorrhagic infarction of the pituitary is accompanied by the appropriate clinical features.

METABOLIC DISORDERS

AMYLOIDISIS

Amyloid deposition involving the pituitary usually occurs as part of a systemic disorder but is occasionally seen in pituitary adenomas, most commonly prolactinomas (88-92). The amyloid is deposited either in vessel walls or the interstitium, where it is an extracellular, amorphous, eosinophilic substance that stains with Congo Red and exhibits characteristic apple-green birefringence under polarized light.

HEMOSIDEROSIS

Deposition of iron occurs in the pituitary glands of patients with hemochromatosis; there is preferential deposition in gonadotrophs (93;94).

HYPERPLASIA

Hyperplasia is an increase in the number of cells of an organ or tissue in response to a stimulus. Any cell population within the pituitary gland can undergo hyperplasia. This process can be physiological or pathological (3;95;96) and when prolonged may progress to adenoma formation

(97-102).

Somatotroph hyperplasia is seen in patients with ectopic production of GH-releasing hormone (GRH) by pheochromocytomas, endocrine tumors of lung, pancreas or other elements of the dispersed endocrine system (97;103;104). Rarely, it may be associated with a gangliocytoma of the hypothalamus (105). Mammosomatotroph hyperplasia is the characteristic pituitary lesion in McCune-Albright syndrome (106;107); rarely, it may be due to GRH excess or it may be idiopathic (108). Lactotroph hyperplasia is physiologic during pregnancy or other conditions of estrogen excess, but pathological idiopathic lactotroph hyperplasia is a rare cause of hyperprolactinemia (109;110). Corticotroph hyperplasia is a cause of Cushing's disease that may be associated with a corticotroph adenoma (98-100). In rare patients, it is attributed to ectopic or eutopic excess of corticotropin-releasing hormone (CRH). It is also physiological in patients with untreated Addison's disease. Thyrotroph hyperplasia is seen in patients with prolonged primary hypothyroidism(111-113). Gonadotroph hyperplasia is seen in patients with prolonged primary hypogonadism (101;102).

Radiologic evaluation of patents with hyperplasia usually reveals diffuse sellar enlargement without enhancing normal tissue on contrast administration. Hyperplasia is usually reversible if the underlying condition is appropriately treated. However, in patients with idiopathic hyperplasia, the underlying stimulus is not known. Patients with lactotroph hyperplasia can be treated with dopaminergic agonists. Those with Cushing's syndrome may require total hypophysectomy to achieve clinical control.

The key to distinguishing adenohypophysial hyperplasia from adenoma lies in the reticulin stain (Figure 14). Hyperplasia is characterized by expanded acini with an intact reticulin framework whereas adenomas have breakdown of the reticulin fiber network. Immunohistochemistry shows predominance of the hyperplastic cell type with other hormone-containing cells interspersed. Ultrastructural examination is not a reliable method to distinguish hyperplasia from adenoma, however, it was the method used to describe the cell enlargement that defines "thyroidectomy" cells (Figure 15) in patients with primary hypothyroidism, and "gonadectomy" cells in patients with primary hypothyroidism, the target cells develop abundant vacuolated cytoplasm that is occupied almost entirely by dilated rough endoplasmic reticulum with secretory material.



Figure 14. The reticulin stain is the most valuable tool to distinguish normal acinar architecture (a) from the expanded acini of hyperplasia (b) and to confirm total breakdown of the reticulin fiber network in adenomas (c).



Figure 15. (a) Thyrotroph hyperplasia in a patient with primary hypothyroidism retains acinar architecture but the expanded acini are dominated by large cells with pale cytoplasm; a few smaller acidophils and basophils are identified (arrow). (b) The large "thyroidectomy" cells have abundant cytoplasm with scattered PAS-positive globules (large arrows); the PAS stain also identifies corticotrophs (short arrows).

PRIMARY TUMORS OF ADENOHYPOPHYSEAL CELLS

Primary tumors of the adenohypophysis include pituitary adenomas and the rare malignant

pituitary carcinoma. Pituitary adenomas are benign neoplasms that arise from adenohypophysial cells and represent up to 25% of all intracranial tumors (3). They are present in approximately 17% of the general population (3, 114). Their prevalence increases with advancing age; both sexes are affected equally.

The cause of sporadic pituitary adenomas is not known, however, there has been significant investigation that has shed light on some of the molecular pathogenetic events associated with pituitary neoplasia (115,116). A minority of these lesions is associated with Multiple Endocrine Neoplasia (MEN) syndromes, familial disorders in which several endocrine glands develop neoplasms or hyperplasias (117). Pituitary adenomas are most commonly associated with MEN-1 (Wermer's syndrome), an autosomal dominant disorder with incomplete penetrance (117). This syndrome is characterized by the development of parathyroid hyperplasia or adenoma, pancreatic endocrine cell hyperplasia, dysplasia and tumor, and pituitary adenoma. The various tumors develop at different times rather than synchronously in individual patients and in no specific order. Approximately two-thirds of affected patients develop a pituitary adenoma, most often producing prolactin and/or GH (118). This disease results from a germline mutation of the MEN 1 gene on chromosome 11g13 that encodes the tumor suppressor protein "menin" (115,116). Loss of heterozygosity of the intact allele is responsible for subsequent tumor formation. However, alterations of menin are not found in the more common sporadic adenomas. A number of etiologic factors have been implicated, including genetic events, hormonal stimulation and growth factors (6,115,116) and it is likely that all of these interact to initiate transformation and promote tumor cell proliferation.

Ectopic pituitary adenomas can arise in embryonic remnants of Rathke's cleft that are found in extrasellar locations including the sphenoid sinus, parapharyngeal area, suprasellar regions, middle nasal meatus, petrous temporal bone, clivus, hypothalamus, and third ventricle (119-127). Epidemiologically, these tumors resemble their intrasellar counterparts. Compared to intrasellar pituitary adenomas, higher proportions of ectopic adenomas are functional with the most common clinical presentation being Cushing's disease. Silent ectopic adenomas usually present with mass effects and their diagnosis depends on careful examination of resected tissue specimens.

CLASSIFICATION OF PITUITARY ADENOMAS

There are several classification schemes for pituitary adenomas: functional, anatomic/radiologic, histologic, immunohistochemical, ultrastructural, and clinicopathologic (3).

The functional classification is used clinically. It groups adenomas according to the hormonal syndromes with which they are associated. This includes the various functioning adenomas and the clinically "silent" or nonfunctioning adenomas.

The anatomic/radiologic classification categorizes pituitary adenomas based on size and degree of invasion (128). Sellar enlargement can be demonstrated in most patients with CT scan but MRI is the imaging technique of choice to identify a pituitary adenoma and delineate residual normal tissue that enhances with contrast more readily than tumor tissue.

The histologic classification, based on histochemical stains, divides adenomas into those that are acidophilic, basophilic, and chromophobic. This classification is of limited value and has largely been abandoned.

The immunohistochemical classification categorizes pituitary adenomas based primarily on hormone content with additional information provided by immunoreactivity for transcription factors and keratins (3).

Although electron microscopy can be extremely valuable in the diagnosis of certain tumors, the ultrastructural classification of adenomas is less commonly utilized.

The clinicopathologic classification represents the most effective classification scheme (3,129). It categorizes adenomas using both morphologic and clinical features as shown in Table 1.

Table 1. Clinicopathological Classification of Pituitary Adenomas

Functioning Adenomas	Nonfunctioning Adenomas
GH-PRL-TSH Family	
Adenomas causing GH-excess	Silent lactotroph adenomas
1. Densely granulated somatotroph	
adenomas	
2. Sparsely granulated somatotroph	
adenomas	
3. Mammosomatotroph adenomas	
Adenomas causing hyperprolactinemia	Silent lactotroph adenomas
1. Lactotroph adenomas	
2. Acidophil stam cell adenomas	
Adenomas causing ISH excess	Slient thyrotroph adenomas
1 Thurotroph adenomas	
Adenomas causing ACTH excess	Silent corticotroph adenomas
Additional causing No TT excess	Cheft context opri adenomas
1 Densely granulated corticotroph	
adenomas	
2. Sparsely granulated corticotroph	
adenomas	
Gonadotroph Family	
Adenomas causing gonadotropin excess	Silent gonadotroph adenomas (null cell
	adenomas, oncocytomas)
1. Gonadotroph adenomas	
Unclassified Adenomas	
1. Unusual plurihormonal adenomas	Immunonegative adenomas

FUNCTIONING SOMATOTROPH ADENOMAS

Somatotroph adenomas arise from GH-producing cells and represent 10 to 15% of pituitary adenomas. GH excess in adults manifests as acromegaly, and gigantism results from excessive GH prior to epiphyseal plate closure (130). The details of clinical and biochemical parameters of acromegaly and gigantism are covered elsewhere in this text. Hyperprolactinemia can be prominent in some patients; this may be the result of stalk compression in macroadenomas, or the elaboration of PRL in addition to GH by the tumor.

Grossly, these tumors are usually well demarcated and are located in the lateral wing of the adenohypophysis. Microscopically, there are several subtypes of somatotroph adenomas.

Densely Granulated Somatotroph Adenomas are composed of acidophilic cells arranged in a trabecular, sinusoidal, or diffuse architecture (Figure 16). The tumor cells show strong, diffuse cytoplasmic immunoreactivity for GH and strong nuclear immunoreactivity for Pit-1 (3). Tumor cells exhibit moderate perinuclear positivity with antibodies to low molecular weight cytokeratins. There is variable cytoplasmic immunoreactivity for alpha-subunit of glycoprotein hormones. By electron microscopy (129), the tumor cells resemble nontumorous somatotrophs; they have spherical nuclei with prominent nucleoli. Parallel arrays of rough endoplasmic reticulum are found and Golgi complexes are well formed. The numerous secretory granules are homogeneous, dense and spherical with diameters ranging from 150 to 600 nm.



Figure 16. Densely Granulated Somatotroph Adenomas are composed of acidophilic cells arranged in a trabecular, sinusoidal, or diffuse architecture (a). The tumor cells show diffuse cytoplasmic immunoreactivity for GH (b) and strong nuclear immunoreactivity for Pit-1 (c). In contrast to the sparsely granulated variant (see keratin positivity is perinuclear and faint (d).

Sparsely Granulated Somatotroph Adenomas are composed of solid sheets of chromophobic cells with striking nuclear pleomorphism and prominent nucleoli that can mimic metastatic carcinomas (Figure 17). Immunohistochemical stains for low molecular weight cytokeratins reveal the characteristic feature of this tumor type, the fibrous body, which manifests as juxtanuclear globular reactivity (3). There is focal, weak cytoplasmic immunoreactivity for GH and occasionally for alpha-subunit. Nuclear immunoreactivity for Pit-1 is usually present. Ultrastructurally (129), the tumor cells are irregularly shaped with eccentric, pleomorphic, and often multilobulated nuclei. The rough endoplasmic reticulum can be either poorly or well developed. The characteristic fibrous body is a juxtanuclear, spherical mass composed of intermediate filaments. Secretory granules are sparse and range in size from 100 to 250 nm.



Figure 17. Sparsely Granulated Somatotroph Adenomas are composed of chromophobic cells with striking nuclear pleomorphism and prominent nucleoli that can mimic metastatic carcinoma (a). Immunohistochemical stains for low molecular weight cytokeratins reveal the characteristic feature of this tumor type, the fibrous body, which manifests as juxtanuclear globular reactivity (b).

Mammosomatotroph Adenomas produce both GH and PRL. They are associated with acromegaly and are the most frequent findings in patients with gigantism and in young patients with acromegaly. Microscopically, the tumor is composed mainly of acidophilic cells arranged in a diffuse or solid pattern. Chromophobic cells may be scattered throughout. Immunohistochemically, the tumor cells are strongly immunoreactive for GH and variably immunoreactive for alpha-subunit and PRL(3). Staining for low molecular weight cytokeratins yields a perinuclear pattern of staining similar to that of normal somatotrophs and the cells of densely granulated somatotroph adenomas. There is strong nuclear immunoreactivity for Pit-1 and occasionally for ER. Very rarely, beta-TSH can be demonstrated in the cytoplasm. Ultrastructurally, the tumor cells resemble densely granulated somatotrophs(129), however, secretory granules have mottled cores, are variably pleomorphic, and can measure up to 1000nm. In addition, the cells exhibit misplaced exocytosis, which is the classic feature of PRL secretion.

Drug Effects

These tumors rarely show a response to dopaminergic agents. Treatment with somatostatin analogues often results in inhibition of GH excess (131), most successfully in patients with densely granulated as compared with sparsely granulated tumors (132). There is, however, no consistent tumor size reduction or morphologic alteration attributable to this therapy (133).

FUNCTIONING LACTOTROPH ADENOMAS

Tumors arising from PRL-secreting adenohypophysial cells are the most common type of pituitary adenoma (134;135). Although almost half of adenomas found incidentally at autopsy are of this type (136), the incidence is much lower in surgical series, probably because these tumors are often treated medically. This tumor is more common in females, who tend to present at a younger age with hormonal disturbances. In contrast, men tend to present later, with larger tumors that more often result in mass effects and hypopituitarism secondary to adenohypophysial destruction (137). While this has traditionally been attributed to sexual dimorphism of the perception of the relevant symptoms, recent evidence suggests that the tumors grow faster in men (138). There are three variants of PRL-secreting pituitary adenomas: sparsely granulated and densely granulated lactotroph adenomas and the rare but aggressive acidophil stem cell tumor.

An elevated serum PRL level greater than 250ng/mL (5000mU/I) is virtually diagnostic of a prolactin secreting adenoma. Values less than this must be interpreted with more caution, since a number of physiologic, pharmacologic, and pathologic conditions may cause elevated serum PRL levels. Lactotroph adenomas tend to have a good correlation between tumor size and PRL level. However, this is not often true for patients with acidophil stem cell adenomas.

Microadenomas are most commonly located in the posterolateral portions of the gland. Macroadenomas may invade into dura mater, nasal sinuses, and bone. They can be soft and red, or gray and firm. Occasionally, the presence of psammoma bodies results in a gritty consistency.

Sparsely Granulated Lactotroph Adenomas are the more common variant. The chromophobic tumor cells are arranged in papillae, trabeculae, or solid sheets; tumor cells may form pseudorosettes around vascular spaces (3). Calcification with the formation of frank psammoma bodies is occasionally present. Amyloid deposition is a rare feature. The tumor cells show strong immunoreactivity for PRL in a juxtanuclear globular pattern that corresponds to the Golgi region (Figure 18). Nuclear staining for Pit-1 is usually present and estrogen receptor positivity may be found by immunohistochemistry. Ultrastructurally (129), the cells have large nuclei with prominent nucleoli. The rough endoplasmic reticulum is prominent and arranged in distinctive parallel arrays; when arranged in concentric whorls, they are known as "Nebenkern formations". The large, well-developed Golgi complexes harbor immature pleomorphic granules. Secretory granules are spherical, sparse, and range in size from 150 to 300 nm. Misplaced exocytosis, or the expulsion of secretory granules from the lateral cell membrane into the extracellular space, is a diagnostic feature of PRL-producing tumors.



Figure 18. Sparsely Granulated Lactotroph Adenomas are characterized by strong immunoreactivity for PRL in a juxtanuclear globular pattern that corresponds to the Golgi region.

Densely Granulated Lactotroph Adenomas are much less common than the sparsely granulated variant. These tumors are composed of acidophilic cells that exhibit diffuse cytoplasmic positivity for PRL, unlike the juxtanuclear Golgi pattern seen in the sparsely granulated adenoma. Ultrastructurally, densely granulated cells have abundant rough endoplasmic reticulum; secretory granules are numerous and can measure up to 700 nm. Misplaced exocytosis is again a diagnostic feature.

The Acidophil Stem Cell Adenoma is usually composed of solid sheets of large cells that are slightly acidophilic due to the accumulation of mitochondria. Large cytoplasmic vacuoles corresponding to giant mitochondria can be easily appreciated by light microscopy (3). The classic immunohistochemical profile shows strong diffuse immunoreactivity for PRL and scant immunoreactivity for GH. Some tumors may lack detectable immunoreactivity for GH. Staining with low molecular weight cytokeratins usually allows the detection of scattered fibrous bodies (3). Electron microscopy may be necessary to render a definitive diagnosis of this tumor (129). Ultrastructurally, the cells are elongated with oval or irregular nuclei. The cytoplasm is occupied

by numerous enlarged mitochondria; distinctive giant mitochondria containing electron dense tubules are frequently seen. Fibrous bodies, or juxtanuclear bundles of cytokeratin intermediate filaments, identical to those seen in sparsely granulated somatotroph adenomas, are scattered throughout the tumor. Secretory granules are scant and range in size form 150 to 200 nm. Misplaced exocytosis can be seen.

Drug Effects. Administration of dopamine agonists such as bromocriptine results in a dramatic clinical response in patients with prolactinomas. The rapid fall in serum PRL is accompanied by an almost equally rapid reduction in tumor size that is due to tumor cell shrinkage (139). In the absence of relevant history, the changes may be a source of diagnostic confusion. The cytoplasm of the tumor cells shrinks, resulting in a marked increase in cellularity. The resulting picture can histologically resemble a lymphoma. These changes are reversible upon discontinuation of therapy; however, the alterations may be permanent in a small population of tumor cells. After chronic therapy, there is occasionally interstitial and perivascular fibrosis along with hemorrhage and hemosiderin deposition.

The acidophil stem cell adenoma is generally resistant to treatment with bromocriptine (140). Surgical resection is necessary for these aggressive tumors; careful postoperative monitoring is required, as recurrence is common. Radiation therapy may play a role in the management of recurrent acidophil stem cell tumors.

FUNCTIONING THYROTROPH ADENOMAS

Thyrotroph adenomas account for less than one percent of all pituitary neoplasms. Patients with pituitary-dependent TSH excess may exhibit features of hyperthyroidism or hypothyroidism, or may be euthyroid (141). Because most thyrotroph tumors are invasive macroadenomas, mass effects with visual field disturbances are common.Diffuse thyrotroph hyperplasia can mimic this disorder therefore thyroid function tests should be performed in all patients with elevated levels of serum TSH to exclude primary thyroid failure, and radiological evaluation with an MRI scan is essential to identify the presence of a discrete sellar tumor rather than diffuse hyperplasia.

Grossly, thyrotroph adenomas tumors tend to be invasive and fibrotic macroadenomas at the time of diagnosis.

By light microscopy, these tumors are composed of chromophobic cells with indistinct cell borders and varying degrees of nuclear pleomorphism (Figure 19). Architecturally, the tumors most commonly exhibit a solid or sinusoidal pattern. Stromal fibrosis is relatively common and occasional psammoma bodies may be present. The tumor cells show variable immunoreactivity for alpha-subunit and beta-TSH. Immunohistochemistry highlights the polygonal structure of the tumor cells that usually have elongated processes (3). Ultrastructurally, thyrotroph adenoma cells resemble normal thyrotrophs (129). The polygonal cells have euchromatic nuclei and long interdigitating cytoplasmic processes that contain abundant rough endoplasmic reticulum with prominent and spherical Golgi bodies. Secretory granules, which are spherical and range in size from 150 to 250 nm, tend to accumulate along the cell membrane. Some densely granulated tumors occasionally have larger granules measuring up to 350 nm.



Figure 19. Thyrotroph adenomas are composed of chromophobic cells with indistinct cell borders and varying degrees of nuclear pleomorphism with hyperchromasia and marked atypia (a). Immunohistochemistry identified α -subunit and β -TSH (b) in tumor cells.

FUNCTIONING CORTICOTROPH ADENOMAS

Tumors composed of ACTH-secreting corticotrophs represent 10 to 15% of all pituitary adenomas. There are three recognized variants: densely granulated corticotroph adenomas, sparsely granulated corticotroph adenomas, and Crooke's cell adenomas. Cushing's disease, defined as pituitary-dependent hypercortisolism, has a number of clinical manifestations that are reviewed elsewhere in this text.

The most common cause of Cushing's disease is a basophilic microadenoma (3). These small tumors may be centrally located, as corticotrophs are most abundant in the median wedge of the anterior pituitary, but they usually exhibit lateralization of blood supply, justifying inferior petrosal sinus sampling. Macroadenomas are associated with Nelson's syndrome, or represent chromophobic or sparsely granulated adenomas in patients with less florid hormone excess. The possibility of corticotroph hyperplasia must be considered in the differential diagnosis of this disorder.

Densely granulated corticotroph adenomas are the most common type of corticotroph adenoma. By light microscopy, these tumors are composed of basophilic cells arranged in a sinusoidal architecture. The tumor cells exhibit cytoplasmic PAS positivity. They are immunoreactive for ACTH, beta-endorphin and other POMC-derived peptides. Positivity for low molecular weight cytokeratins is seen in patients with Cushing's disease; in patients with Nelson's syndrome, the tumor cells do not accumulate filaments of keratin. Ultrastructurally (129), corticotroph cells are large and polygonal with ovoid or irregular nuclei that harbor nucleoli in contact with the inner nuclear membrane. The cytoplasm contains prominent rough endoplasmic reticulum with abundant free ribosomes, spherical Golgi complexes and perinuclear intermediate filaments composed of cytokeratins that are prominent in patients with Cushing's syndrome but are not conspicuous in Nelson's syndrome. The secretory granules range in size from 150 to 450nm in diameter and are distinctive because of their marked variability in shape and electron density.

Sparsely granulated corticotroph adenomas are less common than the densely granulated variant. By light microscopy, the tumor cells are chromophobic (Figure 20) and are negative or only focally positive with the PAS stain. They exhibit strong immunoreactivity for cytokeratins and weak immunoreactivity for ACTH and other POMC-derived peptides (3). Ultrastructurally (129), the cells contain less well-developed organelles and scant secretory granules, but the characteristic variability of size, shape and density of the granules characterizes them as corticotrophs.



Figure 20. Corticotroph adenomas may be basophilic or chromophobic (a). They exhibit strong immunoreactivity for cytokeratins (b) and may have diffuse or focal, strong or weak immunoreactivity for ACTH and other POMC-derived peptides.

Crooke's cell adenomas are rare tumors that exhibit Crooke's hyaline change (142;143). Usually, Crooke's hyalinization is restricted to nontumorous corticotrophs, but rarely this marker of feedback suppression by glucocorticoids is seen in adenomas. These tumors can be associated with Cushing's disease, but it is generally an unusual form of the disease, such as cyclical Cushing's. The tumor cells can exhibit marked cytologic and nuclear atypia (Figure 21). The perinuclear ring of pale hyaline material represents the accumulation of low molecular weight cytokeratin filaments that are intermediate filaments on electron microscopy (144). The cells exhibit a rim of peripheral positivity when stained with PAS, and this is due to immunohistochemically detectable ACTH in secretory granules located either at the periphery of the cell or in the perinuclear region.



Figure 21. Crooke's cell adenomas exhibit marked cytologic and nuclear atypia. The perinuclear ring of pale hyaline material represents the accumulation of low molecular weight cytokeratin filaments in these tumor cells.

FUNCTIONING GONADOTROPH ADENOMAS

These tumors are mainly diagnosed in middle-aged men with no prior history of gonadal dysfunction (145). Although they occur in women, the clinical diagnosis is more often missed because elevation of gonadotropins is considered to be physiological in postmenopausal women and the tumors are considered to be nonfunctional (146).

Grossly, the tumors are large, soft, well vascularized, and occasionally have foci of hemorrhage or necrosis. Microscopically, they are characterized by chromophobic cells arranged in a trabecular, papillary, or sinusoidal pattern (147). There is usually prominent pseudorosette formation around vascular spaces (Figure 22). Focal oncocytic change is quite common. Scant PAS positivity may be demonstrated in some tumor cells. The tumor cells exhibit, with variable intensity of immunoreactivity for alpha-subunit, beta-FSH, and beta-LH. As well, there is strong nuclear staining with steroidogenic factor-1 (3). It is common for gonadotroph adenomas to

exhibit ultrastructural diversity (129;148). Well-differentiated tumor cells are elongated with the nucleus occupying one pole and secretory granules accumulate at the opposite pole. Poorly differentiated cells are generally ovoid or polygonal and lack polarity. Rough endoplasmic reticulum is usually composed of short dilated profiles that contain flocculent material. Golgi bodies are perinuclear, large, and globular. Secretory granules are generally small (250nm), variable in number, and located close to the cell membrane. Cells exhibiting oncocytic change have abundant mitochondria.



Figure 22. Gonadotroph adenomas are characterized by cuboidal to columnar chromophobic cells arranged in a trabecular, papillary, or sinusoidal pattern with prominent pseudorosette formation around vascular spaces (a). They often exhibit focal oncocytic change with nests of round to polygonal cells that have abundant granular cytoplasm (b).

CLINICALLY NON-FUNCTIONING PITUITARY ADENOMAS

These tumors account for approximately one third of all pituitary adenomas. Due to their lack of clinically detectable hormonal activity, they tend to present with mass effects such as headache, visual field deficits, cranial nerve defects, or rarely, cavernous sinus syndrome (3). If there is extensive tissue destruction, hypopituitarism results in clinical symptomatology. Less commonly, pituitary apoplexy with hemorrhage into the tumor causes a medical emergency. Patients may have varying degrees of hypopituitarism depending on the amount of adenohypophysial tissue destruction. There is no evidence of hormone excess, however, stalk compression without significant adenohypophysial destruction can result in mild hyperprolactinemia.

The diagnosis of silent pituitary adenomas is based solely on morphologic features of the tumor (3). Silent somatotroph adenomas have morphologic features similar to those of sparsely granulated somatotroph adenomas. Silent lactotroph adenomas and silent thyrotroph adenomas exhibit morphologic features corresponding to those of their functioning counterparts. Silent corticotroph adenomas are usually associated with hyperprolactinemia even in cases without obvious stalk involvement. There are two morphologic variants. Type I silent corticotroph

adenomas correspond morphologically to the functioning densely granulated corticotroph adenoma. Type II silent corticotroph adenomas are similar to the sparsely granulated functioning corticotroph adenomas. The clinical inactivity of some corticotroph adenomas may be due to aberrant cleavage of the POMC molecule. Silent gonadotroph adenomas are morphologically identical to the functioning gonadotroph adenomas and represent the largest group of clinically nonfunctioning adenomas. Most tumors classified as null cell adenomas are silent gonadotroph adenomas composed of poorly differentiated cells with scattered foci exhibiting histologic features consistent with gonadotroph differentiation; these tumors generally exhibit SF-1 staining despite lack of detectable gonadotropin content. Oncocytomas represent silent gonadotroph adenomas with extensive oncocytic change (Figure 23). The tumor cells are usually arranged sheets or nests, and contain abundant granular eosinophilic cytoplasm, which corresponds ultrastructurally to mitochondrial accumulation in the cytoplasm. These tumors also generally exhibit SF-1 nuclear reactivity (3).



Figure 23. Oncocytomas represent silent gonadotroph adenomas with extensive oncocytic change. The tumor cells are arranged solid nests, and contain abundant granular eosinophilic cytoplasm (a). Immunohistochemistry identifies SF-1 nuclear reactivity and cytoplasmic staining for α -subunit as well as β -subunits of the gonadotropins, e.g. β -FSH in this tumor (b).

Despite advances in morphologic classification of adenohypophysial cells, due to improved tissue fixation, more specific and sensitive antibodies, and transcription factors that identify cell differentiation, there remain a minority of pituitary adenomas that defy definitive classification on based on histological, immunohistochemical, and ultrastructural examination. Poorly differentiated adenomas are negative for all hormones and transcription factors and exhibit no ultrastructural markers of the known adenohypophysial cell types (3).

The tumor identified as "female type gonadotroph adenoma" (149) is a tumor that is usually clinically silent, immunohistochemically plurihormonal, and characterized by a distinctive ultrastructural feature of dilated, saccular Golgi bodies known as "honeycomb Golgi". The cytogenesis of this lesion is not known, but recent data suggest that the honeycomb Golgi is not

a specific finding and some of these tumors may actually represent corticotroph adenomas (149a).

PLURIHORMONAL PITUITARY ADENOMAS

Occasionally, pituitary adenomas elaborate multiple hormones. Most often, this is due to known regulatory factors; rarely, products of different hormone families are produced. These adenomas may be fully functioning, partially functioning (in which only one component is clinically apparent), or silent. The most common combination is excessive production of GH and PRL, or GH, PRL and TSH, resulting in acromegaly/gigantism accompanied by hyperprolactinemia and even hyperthyroidism. This pattern of plurihormonality is accounted for by the expression of Pit-1 that regulates the expression of these various hormones by related cells. Other combinations of unrelated hormones have been reported (3). The interpretation of the individual combinations must be evaluated with caution, and may in some cases be artefacts of antibody cross-reactivity.

Monomorphous plurihormonal adenomas are composed of one cell type that can produce multiple hormones; this is supported by cytoplasmic immunoreactivity for two or more hormones within the same cell. Plurimorphous plurihormonal adenomas are composed of at least two cell types, each of which exhibits a characteristic immunohistochemical and ultrastructural profile. These may represent "collision" tumors (150;151). Silent subtype III adenomas are rare and aggressive plurihormonal tumors that are identified by unique ultrastructural features (152). The large tumor cells have nuclei that are located at one pole and may contain spheridia. There is abundant well-developed rough endoplasmic reticulum with prominent, tortuous Golgi complexes, and abundant groups of mitochondria. Small secretory granules are localized to attenuated, interdigitating cell processes. This lesion is considered more aggressive and local recurrence is not uncommon; there may be a role for radiotherapy for this relatively radiosensitive lesion.

PITUITARY CARCINOMA

Malignant tumors of adenohypophysial origin are defined by the ability to metastasize (3;153). Although many pituitary adenomas are widely invasive, destructive of adjacent tissues, and lethal, they are not classified as malignancies. The pathogenesis of the very rare pituitary carcinoma is not known. H-ras point mutations have been reported in some metastatic foci, but not in the corresponding primary tumor (154;155). Immunoreactivity for p53 has been reported, but p53 mutations are not found in these lesions and there is insufficient evidence at this point to draw any conclusions regarding the mechanism of the apparent accumulation of p53 protein (156).

Pituitary carcinomas generally present initially as pituitary adenoma. They can be associated with any form of hormone excess (157-162), or they may be clinically nonfunctioning lesions (163;164). Only the subsequent development of metastases identified the lesion as malignant. The most common sites of metastasis include the subarachnoid space, brain parenchyma (not including areas of direct invasion), cervical lymph nodes, bone, liver, and lungs. Examination of

the primary tumor usually reveals non-specific morphologic features such as hypercellularity, hemorrhage, necrosis, mitoses, nuclear pleomorphism, and invasion; none of these features either individually, or in combination, are reliable indicators of malignancy.

Immunohistochemistry and electron microscopy are used mainly to characterize the tumor based on the classification scheme applied to the more common adenomas (165).

OTHER PRIMARY TUMORS OF THE SELLAR REGION

CRANIOPHARYNGIOMA

This benign but locally invasive tumor, which originates from the remnants of Rathke's pouch, represents 2 to 4 percent of all intracranial neoplasms (166). It is the most common sellar tumor in children and accounts for 10% of all childhood CNS tumors. Craniopharyngiomas can occur at all ages but the peak incidence occurs from 5 to 20 years old with a second smaller peak occurring in the sixth decade. Some series show a male predominance.

Three quarters of patients have mass effects with headaches and visual field disturbances (50). Patients may have psychiatric disturbances, nausea, vomiting, and somnolence. Hypopituitarism is identified in the majority but is not often the presenting complaint. In contrast to patients with large pituitary adenomas, hyperprolactinemia is found in less than half of patients (167;168) and about 25% of patients have diabetes insipidus. Children may present with dwarfism.

Radiologic evaluation reveals a variably cystic lesion; only 10% are entirely solid with no cystic component. An enlarged or eroded sella turcica is encountered in 50% of cases; suprasellar calcification is present in more than 50% of cases. MRI is the preferred technique to determine the extent of the lesion but unlike CT does not show the calcification; there is often a strong T1 signal on MRI in the absence of contrast due to high lipid content.

The natural history of these lesions is extensive infiltration with significant tissue damage. Infiltration may involve the hypothalamus or extend to as high as the third ventricle. Complete surgical resection is curative (169). However, the highly infiltrative nature of this lesion often results in incomplete resection with a subsequent high recurrence rate of 10 to 62%; this is especially true in younger patients. Post-operative radiation has been advocated to reduce recurrence. Hormone replacement may be necessary for persistent hypopituitarism.

Complications of untreated disease include hydrocephalus if there is extension and obstruction of the third ventricle, and rupture with abscess formation. A single case of malignant transformation in a craniopharyngioma has been reported (170).

Craniopharyngiomas are entirely suprasellar in 85% of cases; an intrasellar component is present in only 15%. Most of these tumors are larger than 1 cm at the time of diagnosis. They are well circumscribed but not necessarily encapsulated and usually contain a thick oil-like fluid resembling "black sludge". The cyst fluid has been shown to contain HCGI (170a). Other features recognized grossly include the presence of cholesterol and calcification.

By light microscopy, the tumor is characterized by islands of epithelial cells and cysts within a loose fibrous stroma (Figure 24). Cholesterol clefts are common. There is often keratin debris, which forms the nidus for calcification. Occasionally, there is a mixed chronic inflammatory infiltrate composed of lymphocytes, plasma cells and macrophages. Although grossly well delineated, microscopically these tumors frequently have infiltrative borders with associated gliosis of the adjacent brain. Two histologic types are identified. The adamantinomatous variant has a prominent stellate component and resembles the dental ameloblastic organ and other adamantinomas. The less common papillary variant is found in adults. This tumor is characterized by pseudopapillary squamous epithelium in a solid or cystic pattern; palisading, fibrosis, and cholesterol accumulation are usually absent. This variant has a somewhat better prognosis than the adamantinomatous variant. By immunohistochemistry, the presence of cytokeratin reactivity confirms the epithelial nature of these tumors. Immunoreactive HCGI can be demonstrated in some of the cells (171a). Ultrastructural examination reveals tonofilaments, intercellular junctions, and the absence of secretory granules.



Figure 24. Craniopharyngioma is characterized by islands of epithelial cells with a loose fibrous stroma.

Beta-catenin gene mutations have been documented in adamantinomatous but not papillary craniopharyngiomas (171b), suggesting a role for \Box -catenin in tumorigenesis. Cytoplasmic and nuclear localization of immunohistochemical reactivity for \Box -catenin correlates with mutation; in contrast papillary craniopharyngiomas with no mutations show exclusively membranous expression of this protein (171b). There is evidence that stromal cells may also harbor \Box -catenin mutations, suggesting that the stroma may be a true neoplastic component rather than reaction to the proliferating epithelium (171b).

NEURONAL TUMORS

These tumors are also known as "gangliocytomas" or "ganglioneuromas" (3). They are composed of mature neurons, most likely derived from the ganglion cells of the hypothalamus. Clinically, they can present with mass effects, hypothalamic dysregulation, hypopituitarism, and hyperprolactinemia (171). Because these tumors have the ability to synthesize hypothalamic peptides, they may sometimes be associated with other hormonal syndromes including acromegaly, precocious puberty, or Cushing's disease (171). Histologically, they are composed of mature ganglion cells with abundant cytoplasm that contains Nissl substance, and large nuclei with prominent nucleoli (Figure 25). Binucleate or even multinucleated cells are not uncommon. The tumor cells are distributed within a variable stroma composed of neuroglia, or fibrous tissue with small vessel proliferation. These tumors are immunoreactive for synaptophysin and neurofilaments and may contain hypothalamic peptides. Ultrastructurally, the tumor cells resemble mature neurons with abundant endoplasmic reticulum, mitochondria, and neurofilaments. Secretory granules are concentrated in neuronal processes. The neuronal lesions associated with acromegaly have often been composite tumors with a pituitary adenoma component, usually a sparsely granulated somatotroph adenoma.



Figure 25. Hypothalamic gangliocytomas are composed of mature neurons that may be binucleate (arrows). This tumor is associated with a sparsely granulated somatotroph adenoma (bottom) and the neurons contained immunoreactivity for GRH.

GLIOMAS

These are neoplasms of neuroglia and include astrocytomas, oligodendrogliomas, and ependymomas (3). The pilocytic astrocytoma, which is most common young patients, is the most common glial tumor of the sellar region (172). These lesions can be sporadic, associated with inherited conditions, or may follow cranial irradiation (173;174). Low-grade gliomas occurring in children have a good prognosis. Post irradiation gliomas and those affecting the optic nerve are aggressive and rapidly lethal.

MENINGIOMAS

Meningiomas are neoplasms of the meninges, most commonly of arachnoid origin. They occur more frequently in females. Meningiomas of the sellar region account for up to 20% of all meningiomas. They can present with neurological deficits, visual field defects, hypopituitarism,

and hyperprolactinemia due to stalk compression (175). Completely intrasellar meningiomas are rare (176). They have been reported following radiotherapy for pituitary adenoma (177-179).

GRANULAR CELL TUMORS

Granular cell tumors are benign neoplasms of uncertain histogenesis found in the neurohypophysis or distal pituitary stalk (180;181). Most of these tumors are small and are incidental autopsy findings. Occasionally, they present with visual field deficits. Diabetes insipidus is rare. These tumors are unencapsulated and composed of cells with abundant, granular, eosinophilic cytoplasm. The granules are PAS positive and diastase resistant. The tumor cells exhibit variable immunoreactivity for the histiocytic markers alpha-1-antitrypsin, alpha-1-antichymotrypsin, and cathepsin B. They are usually nonimmunoreactive for GFAP and S-100 protein (182). Ultrastructurally, the granular cells have phagolysosomes containing debris and electron dense material.

CHORDOMAS

Chordomas are rare midline tumors derived from notochord remnants (183). They usually occur in patients over thirty years old. These tumors are slow growing but locally aggressive. Clinically, patients with parasellar chordomas can present with hypopituitarism. Radiologically, they are lobulated, osteolytic lesions with foci of calcification. Elevation of the periosteum is a characteristic feature of chordomas. The treatment of choice is surgical excision. Tumors that are incompletely resected can be irradiated. Mean survival from time of diagnosis is approximately 5 years. Grossly, chordomas are gelatinous, lobulated, and calcified. Microscopically, the tumors are composed of large polygonal cells called "physaliphorous" cells because of their bubbly cytoplasmic vacuoles containing glycogen and neutral mucins. The cells are arranged in solid sheets or trabeculae within a stroma of acidic mucin. The tumor cells are immunoreactive for low molecular weight cytokeratins, epithelial membrane antigen (EMA), S100 protein, and sometimes carcinoembryonic antigen (CEA). Ultrastructurally, desmosomes and microvilli are present. The rough endoplasmic reticulum forms concentric rings around mitochondria.

SCHWANNOMAS

These tumors are also known as neurolemmoma. Schwannomas of this region are derived from the Schwann cells surrounding cranial nerves. They are rare tumors that can present as a sellar mass with or without hypopituitarism or hyperprolactinemia (184-186). These tumors are usually benign. Surgical resection is the treatment of choice. Histologically, these tumors are usually encapsulated lesions composed of spindle-shaped cells arranged into the classic Antoni A and Antoni B areas. Verocay bodies result from palisading of tumor cells in Antoni A areas. The tumor cells are immunoreactive for S-100 protein. Ultrastructural examination reveals basal lamina and pathognomonic long-spacing collagen.

GERM CELL TUMORS

These are midline tumors that arise from residual germ cells. They include germinomas, embryonal carcinomas, teratomas, endodermal sinus tumors, and choriocarcinomas. These tumors represent less than 1% of intracranial tumors in adults (187). However, in children, germ cell tumors account for approximately 6.5% of all intracranial neoplasms (188). They are relatively rare after age twenty, with males more frequently affected than females. The most common site of intracranial involvement is the pineal gland, followed by the suprasellar region (189). Within the sellar region, pure germinomas and pure teratomas are the predominant subtypes. Mixed germ cell tumors are also common; they usually contain a germinoma in combination with some other component (190;191). Tumors that produce beta-hCG may result in precocious puberty.

HEMATOLOGIC TUMORS

Neoplastic proliferations of myeloid, lymphoid or plasmacytoid cells within the hypophysis and hypothalamus usually occur as part of a systemic disorder. However, very rarely, the hypophysis and/or hypothalamus may be the primary site of involvement (192-194). Histologically, these neoplasms are similar to their extracranial counterparts. They are most commonly non-Hodgkin's lymphomas composed of B-cells (195).

LANGERHANS' CELL HISTIOCYTOSIS

Langerhans' cell histiocytosis is characterized by proliferation of the Langerhans' cell, which is a special type of histiocyte with dendritic processes and antigen presenting capabilities. This disease can be unifocal, multifocal, or disseminated, and is separated into three clinicopathologic entities: Letterer-Siwe disease, eosinophilic granuloma, and Hand-Schuller-Christian disease (3). Characteristically, the granulomas may produce cranial diabetes insipidus with or without growth hormone and gonadotrophin and occasionally other hormone deficiencies due to anterior hypothalamic infiltration although there may also be infiltration of the pituitary stalk itself.

These lesions are characterized by large Langerhans' cells that have abundant pink cytoplasm with characteristic indented or "kidney-bean" shaped nuclei. They are usually accompanied by a mixed inflammatory infiltrate including lymphocytes, plasma cells, and eosinophils. The Langerhans' cells are immunoreactive for CD1a and S-100 protein. Ultrastructurally, the presence of Birbeck granules within the cytoplasm is diagnostic (195;196).

MESENCHYMAL TUMORS

Tumors arising from mesenchyme can be derived from vessels, fat, bone, cartilage, or fibrous tissue. They can be benign or malignant. Involvement of the sellar turcica by such neoplasms is uncommon; they usually manifest with mass effects and variable hypofunction of anterior or posterior pituitary function. Those reported in the sellar region include hemangioma (197;198), glomangioma (199), hemangioblastoma (200), lipoma (3), enchondroma (201), chondroma (202;203), chondrosarcoma (204), chondromyxoid fibroma (205), giant cell tumor (206), alveolar soft part sarcoma (207), osteosarcoma (208), and fibrosarcoma (209). They can be sporadic, or

can occur as part of a clinical syndrome such as von Hippel Lindau disease (200). However, most commonly, sarcomas of the sellar region develop as a result of previous irradiation for lesions such as pituitary adenoma or craniopharyngioma (210).

METASTATIC TUMORS TO THE HYPOPHYSIS AND SELLA TURCICA

The pituitary gland, being a highly vascular organ, can be the target of blood-borne metastases from many malignancies. Metastatic tumors to the pituitary gland are not an uncommon event; the frequency has been reported to be as high as 27 percent. Involvement of the neurohypophysis is more common than the adenohypophysis. The most common sites of origin are lung, breast, and gastrointestinal tract (211-214).

Metastatic tumors to the pituitary gland are usually not among the most prominent clinical complaints of patients with disseminated malignancy and are usually discovered at autopsy. They may occasionally present as a sellar tumor in a patient with an occult primary. Clinically they are distinguished from primary pituitary adenomas by the prominence of diabetes insipidus, and mass effects such as headaches, visual field defects, ptosis, and ophthalmoplegia; hypopituitarism is less evident and occurs when adenohypophysial involvement is extensive. In rare cases, metastatic involvement of a pituitary adenoma may result in rapid increase in tumor size and/or sudden worsening symptoms (215-218).

MISCELLANEOUS LESIONS

Other rare and unusual lesions of the pituitary region include inflammatory pseudotumors (219), aneurysm (220), meningoencephlocele (221), hamartomas and choristomas (222;223), and brown tumor of bone (126).

References

1. Asa SL, Penz G, Kovacs K, Ezrin C 1982 Prolactin cells in the human pituitary. A quantitative immunocytochemical analysis. Archives of Pathology and Laboratory Medicine 106:360-363

2. Elster AD, Sanders TG, Vines FS 1991 Size and shape of the pituitary gland during pregnancy and post partum: measurement with MR imaging. Radiology (Easton,PA) 181:531-535

3. Asa SL 1998 Tumors of the Pituitary Gland. Atlas of Tumor Pathology, Third Series, Fascicle 22, Armed Forces Institute of Pathology, Washington, D.C.

4. Asa SL, Kovacs K, Melmed S 1995 The hypothalamic-pituitary axis. In: Melmed S (ed). The Pituitary.Blackwell Scientific Publication Inc., Boston:3-44

5. Asa SL, Kovacs K 1984 Functional morphology of the human fetal pituitary. Pathology Annual

19 Pt 1:275-315

6. Asa SL, Ezzat S 1998 The cytogenesis and pathogenesis of pituitary adenomas. Endocrine Reviews 19:798-827

7. Asa SL, Ezzat S 1999 Molecular determinants of pituitary cytodifferentiation. Pituitary 1:159-168

8. Scully KM, Rosenfeld MG 2002 Pituitary development: regulatory codes in mammalian organogenesis. Science 295:2231-2235

9. Rosenfeld MG 1991 POU-domain transcription factors: pou-er-ful developmental regulators. Genes and Development 5:897-907

10. Asa SL, Puy LA, Lew AM, Sundmark VC, Elsholtz HP 1993 Cell type-specific expression of the pituitary transcription activator Pit-1 in the human pituitary and pituitary adenomas. Journal of Clinical Endocrinology and Metabolism (Baltimore) 77:1275-1280

11. Friend KE, Chiou Y-K, Laws ER, Jr., Lopes MBS, Shupnik MA 1993 Pit-1 messenger ribonucleic acid is differentially expressed in human pituitary adenomas. Journal of Clinical Endocrinology and Metabolism (Baltimore) 77:1281-1286

12. Pellegrini I, Barlier A, Gunz G, Figarella-Branger D, Enjalbert A, Grisoli F, Jaquet P 1994 Pit-1 gene expression in the human pituitary and pituitary adenomas. Journal of Clinical Endocrinology and Metabolism (Baltimore) 79:189-196

13. Day RN, Koike S, Sakai M, Muramatsu M, Maurer RA 1990 Both Pit-1 and the estrogen receptor are required for estrogen responsiveness of the rat prolactin gene. Molecular Endocrinology (Baltimore) 4:1964-1971

14. Drolet DW, Scully KM, Simmons DM, Wegner M, Chu K, Swanson LW, Rosenfeld MG 1991 TEF, a transcription factor expressed specifically in the anterior pituitary during embryogenesis, defines a new class of leucine zipper proteins. Genes and Development 5:1739-1753

15. Dasen JS, O'Connell SM, Flynn SE, Treier M, Gleiberman AS, Szeto DP, Hooshmand F, Aggarwal AK, Rosenfeld MG 1999 Reciprocal interactions of Pit1 and GATA2 mediate signaling gradient- induced determination of pituitary cell types. Cell 97:587-598

16. Lamolet B, Pulichino AM, Lamonerie T, Gauthier Y, Brue T, Enjalbert A, Drouin J 2001 A pituitary cell-restricted T box factor, Tpit, activates POMC transcription in cooperation with Pitx homeoproteins. Cell 104:849-859

17. Therrien M, Drouin J 1993 Cell-specific helix-loop-helix factor required for pituitary expression of the pro-opiomelanocortin gene. Molecular and Cellular Biology 13:2342-2353

18. Poulin G, Turgeon B, Drouin J 1997 NeuroD1/beta2 contributes to cell-specific transcription

of the proopiomelanocortin gene. Molecular and Cellular Biology 17:6673-6682

19. Ingraham HA, Lala DS, Ikeda Y, Luo X, Shen W-H, Nachtigal MW, Abbud R, Nilson JH, Parker KL 1994 The nuclear receptor steroidogenic factor 1 acts at multiple levels of the reproductive axis. Genes and Development 8:2302-2312

20. Asa SL, Kovacs K, Bilbao JM 1983 The pars tuberalis of the human pituitary. A histologic, immunohistochemical, ultrastructural and immunoelectron microscopic analysis. Virchows Archiv A, Pathological Anatomy and Histopathology (Berlin) 399:49-59

21. Stanfield JP 1960 The blood supply of the human pituitary gland. Journal of Anatomy (London) 94:257-273

22. Daniel PM, Prichard MML 1966 Observations on the vascular anatomy of the pituitary gland and its importance in pituitary function. American Heart Journal 72:147-152

23. Gorczyca W, Hardy J 1987 Arterial supply of the human anterior pituitary gland. Neurosurgery (Baltimore) 20:369-368

24. Bergland RM, Page RB 1978 Can the pituitary secrete directly to the brain? (Affirmative anatomical evidence). Endocrinology (Baltimore) 102:1325-1338

25. Bergland RM, Page RB 1979 Pituitary-brain vascular relations: a new paradigm. Science 204:18-24

26. Melchionna RH, Moore RA 1938 The pharyngeal pituitary gland. American Journal of Pathology 14:763-771

27. Boyd JD 1956 Observations of the human pharyngeal hypophysis. Journal of Endocrinology (London) 14:66-77

28. Ciocca DR, Puy LA, Stati AO 1985 Identification of seven hormone-producing cell types in the human pharyngeal hypophysis. Journal of Clinical Endocrinology and Metabolism (Baltimore) 60:212-216

29. Hori A 1985 Suprasellar peri-infundibular ectopic adenohypophysis in fetal and adult brains. Journal of Neurosurgery (Baltimore) 62:113-115

30. Colohan ART, Grady MS, Bonnin JM, Thorner MO, Kovacs K, Jane JA 1987 Ectopic pituitary gland simulating a suprasellar tumor. Neurosurgery (Baltimore) 20:43-48

31. Björklöf K, Brundelet PJ 1965 Typus degenerativus amstelodamensis (Cornelia de Lange first syndrome). Congenital hypopituitarism due to a cyst of Rathke's cleft? Acta Pediatrica Scandinavica 54:275-287

32. Fujita K, Matsuo N, Mori O, Koda N, Mukai E, Okabe Y, Shirakawa N, Tamia S, Itagane Y,

Hibi I 1992 The association of hypopituitarism with small pituitary, invisible stalk, type 1 Arnold-Chiari malformation, and syringomyelia in several patients born in breech position: a further proof of birth injury theory on the pathogenesis of "idiopathic hypopituitarism". European Journal of Pediatrics 151:266-270

33. Ehrlich RM 1957 Ectopic and hypoplastic pituitary with adrenal hypoplasia. Journal of Pediatrics (St Louis) 51:377-384

34. Moncrieff MW, Hill DS, Archer J, Arthur LJH 1972 Congenital absence of pituitary gland and adrenal hypoplasia. Archives of Disease in Childhood 47:136-137

35. Kauschansky A, Genel M, Walker Smith GJ 1979 Congenital hypopituitarism in female infants. Its association with hypoglycemia and hypothyroidism. American Journal of Diseases of Children 133:165-169

36. Kosaki K, Matsuo N, Tamai S, Miyama S, Momoshima S 1991 Isolated aplasia of the anteior pituitary as a cause of congenital panhypopituitarism. Hormone Research (Basel) 35:226-228

37. Pholsena M, Young J, Couzinet B, Schaison G 1994 Primary adrenal and thyroid insufficiencies associated with hypopituitarism: A diagnostic challenge. Clinical Endocrinology (Oxf) 40:693-695

38. Dattani MT, Martinez-Barbera JP, Thomas PQ, Brickman JM, Toresson H, Fox M, Hindmarsh PC, Krauss S, Robinson IC 1998 Mutations in the homeobox gene HESX1/Hesx1 associated with septo-optic dysplasia in human and mouse. Nature Genetics 19:125-133

39. Wu W, Cogan JD, Pfäffle RW, Dasen JS, Frisch H, O'Connell SM, Flynn SE, Brown MR, Mullis PE, Parks JS, Phillips JAI, Rosenfeld MG 1998 Mutations in PROP1 cause familial combined pituitary hormone deficiency. Nature Genetics 18:147-149

40. Fofanova O, Takmura N, Kinoshita E, Parks JS, Brown MR, Peterkova VA, Evgrafov OV, Goncharov NP, Bulatov AA, Dedov II, Yamashita S 1998 Compound heterozygous deletion of the prop-1 gene in children with combined pituitary hormone deficiency. Journal of Clinical Endocrinology and Metabolism (Baltimore) 83:2601-2604

41. Tatsumi K, Miyai K, Notomi T, Kaibe K, Amino N, Mizuno Y, Kohno H 1992 Cretinism with combined hormone deficiency caused by a mutation in the Pit-1 gene. Nature Genetics 1:56-58

42. Pfäffle RW, DiMattia GE, Parks JS, Brown MR, Wit JM, Jansen M, van der Nat H, van den Brande JL, Rosenfeld MG, Ingraham HA 1992 Mutation of the POU-specific domain of Pit-1 and hypopituitarism without pituitary hypoplasia. Science 257:1118-1121

43. Radovick S, Nations M, Du Y, Berg LA, Weintraub BD, Wondisford FE 1992 A mutation in the POU-homeodomain of Pit-1 responsible for combined pituitary hormone deficiency. Science 257:1115-1118

44. Li S, Crenshaw EB, III, Rawson EJ, Simmons DM, Swanson LW, Rosenfeld MG 1990 Dwarf locus mutants lacking three pituitary cell types result from mutations in the POU-domain gene pit-1. Nature (London) 347:528-533

45. Roessmann U 1985 Duplication of the pituitary gland and spinal cord. Archives of Pathology and Laboratory Medicine 109:518-520

46. Priesel A 1927 Uber die dystopie der neurohyophyse. Virchows Archiv fur Pathologische Anatomie und Physiologie und fur Klinische Medizin (Berlin) 266:407-415

47. Jordan RM, Kendall JW, Kerber CW 1977 The primary empty sella syndrome. Analysis of the clinical characteristics, radiographic features, pituitary function and cerebrospinal fluid adenohypophysial hormone concentrations. American Journal of Medicine 62:569-580

48. Bergeron C, Kovacs K, Bilbao JM 1979 Primary empty sella. A histologic and immunocytologic study. Archives of Internal Medicine 139:248-249

49. Gharib H, Frey HM, Laws ER, Jr., Randall RV, Scheithauer BW 1983 Coexistent primary empty sella syndrome and hyperprolactinemia. Report of 11 cases. Archives of Internal Medicine 143:1383-1386

50. Shin JL, Asa SL, Woodhouse LJ, Smyth HS, Ezzat S 1999 Cystic lesions of the pituitary: clinicopathological features distinguishing craniopharyngioma, Rathke's cleft cyst, and arachnoid cyst. J Clin Endocrinol Metab 84:3972-3982

51. Obenchain TG, Becker DP 1972 Abscess formation in a Rathke's cleft cyst. Case report. Journal of Neurosurgery (Baltimore) 36:359-362

52. Kucharczyk W, Peck WW, Kelly WM, Norman D, Newton TH 1987 Rathke's cleft cysts: CT, MR imaging, and pathologic features. Radiology (Easton, PA) 165:491-495

53. Yamakawa K, Shitara N, Genka S, Manaka S, Takakura K 1989 Clinical course and surgical prognosis of 33 cases of intracranial epidermoid tumors. Neurosurgery (Baltimore) 24:568-573

54. Abramson RC, Morawetz RB, Schlitt M 1989 Multiple complications from an intracranial epidermoid cyst: case report and literature review. Neurosurgery (Baltimore) 24:574-578

55. Lewis AJ, Cooper PW, Kassel EE, Schwartz ML 1983 Squamous cell carcinoma arising in a suprasellar epidermoid cyst. Case report. Journal of Neurosurgery (Baltimore) 59:538-541

56. Cheung CC, Ezzat S, Smyth HS, Asa SL 2001 The spectrum and significance of primary hypophysitis. J Clin Endocrinol Metab 86:1048-1053

57. Thodou E, Asa SL, Kontogeorgos G, Kovacs K, Horvath E, Ezzat S 1995 Lymphocytic hypophysitis: Clinicopathological findings. Journal of Clinical Endocrinology and Metabolism (Baltimore) 80:2302-2311

58. O'Dwyer DT, Clifton V, Hall A, Smith R, Robinson PJ, Crock PA 2002 Pituitary autoantibodies in lymphocytic hypophysitis target both gamma- and alpha-Enolase – a link with pregnancy? 2002 Archives of Physiology and Biochemistry 110:94-98.

59. Imura H, Nakao K, Shimatsu A, Ogawa Y, Sando T, Fujisawa I, Yamabe H 1993 Lymphocytic infundibuloneurohypophysitis as a cause of central diabetes insipidus. New England Journal of Medicine 329:683-689

60. Hasimoto K, Takao T, Makino S 1997 Lymphocytic and enohypophysitis and lymphocytic infundibuloneurohypophysitis. Endocrine Journal 44:1-10

61. Kamel N, Dagci Illgin S, Corapicioglu D, Deda H, Gullu S 1998 Lymphocytic infundibuloneurohypophysitis preesnting as diabetes insipidus in a man. Journal of Endocrinological Investigation (Milano) 21:537-540

62. Del Pozo JM, Roda JE, Montoya JG, Iglesias JR, Hurtado A 1980 Intrasellar granuloma. Case report. Journal of Neurosurgery (Baltimore) 53:717-719

63. Oeckler RCT, Bise K 1991 Non-specific granulomas of the pituitary: Report of six cases treated surgically. Neurosurg Rev 14:185-190

64. Rickards AD, Harvey PW 1989 Giant-cell granuloma and the other pituitary granulomata. Quarterly Journal of Medicine 23:425-439

65. Scanarini M, d'Ercole AJ, Rotilio A, Kitromilis N, Mingrino S 1989 Giant-cell granulomatous hypophysitis: a distinct clinicopathological entity. Journal of Neurosurgery (Baltimore) 71:681-686

66. Inoue T, Kaneko Y, Mannoji H, Fukui M 1997 Giant Cell granulomatous hypophysitis manifesting as an intrasellar mass with unilateral ophthalmoplegia. Neurol Med Chir (Tokyo) 37:766-770

67. Vasile M, Marsot-Dupuch K, Kujas M, Brunereau L, Braun BS, Cooper C, Tubiana JM 1997 Idiopathic granulomatous hypophysitis: Clinical and imaging features. Neuroradiology 39:7-11

68. Hassoun P, Anayssi E, Salti I 1985 A case of granulomatous hypophysitis with hypopituitarism and minimal pituitary enlargement. Journal of Neurology, Neurosurgery and Psychiatry 48:949-951

69. Holck S, Laursen H 1983 Prolactinoma coexistent with granulomatous hypophysitis. Acta Neuropathologica (Berlin) 61:253-257

70. Yoshioka M, Yamakawa N, Sarro H, Yoneda M, Nakayama T, Kuroki M, Tsuchida T, Sekiya M 1992 Granulomatous hypophysitis with meningitis and hypopituitarism. Internal Medicine 31:1147-1150

71. Rossi GP, Pavan E, Chiesura-Corona M, Rea F, Poletti A, Pessina AC 1994 Bronchocentric granulomatosis and central diabetes insipidus successfully treated with corticosteroids. European Respirology Journal 7:1893-1898

72. Folkerth RD, Price DL, Schwartz M, Black PM, De Girolami U 1998 Xanthomatous hypophysitis. American Journal of Surgical Pathology 22:736-741

73. Glauber HS, Brown BM 1992 Pituitary macroadenoma associated with intrasellar abscess: a case report and review. The Endocrinologist 2:169-172

74. Berger SA, Edberg SC, David G 1986 Infectious disease in the sella turcica. Review of Infectious Diseases 5:747-755

75. Veseley DL, Maldonodo A, Levey GS 1977 Partial hypopituitarism and possible hypothalamic involvement in sarcoidosis. Report of a case and review of the literature. American Journal of Medicine 62:425-431

76. Toth M, Szabo P, Racz K, Szende B, Balogh I, Czirjak S, Slowik F, Glaz E 1996 Granulomatous hypophysitis associated with Takayasu's disease. Clinical Endocrinology 45:499-503

77. Lohr KM, Ryan.L.M., Toohill RJ, Anderson T 1988 Anterior pituitary involvement in Wegener's granulomatosis. J Rheumatol 15:855-861

78. De Bruin WI, van't Verlaat JW, Graamans K, De Bruin TWA 1991 Sellar granulomatous mass in a pregnant woman with active Crohn's disease. Neth J Med 39:136-141

79. Albini CH, MacGillvray MHFJE, Woorhess ML, Klein DM 1988 Triad of hypopituitarism, granulomatous hypophysitis, and ruptured Rathke's cleft cyst. Neurosurg 22:133-136

80. Cannavo S, Romaon C, Calbucci F, Faglia G 1997 Granulomatous sarcoidotic lesion of hypothalamic-pituitary region associated with Rathke's cleft cyst. J Enocrinol Invest 20:77-81

81. Sano T, Kovacs K, Scheithauer BW, Rosenblum MK, Petito CK, Greco CM 1989 Pituitary pathology in acquired immunodeficiency syndrome. Archives of Pathology and Laboratory Medicine 113:1066-1070

82. Sheehan HL, Davis JC 1968 Pituitary necrosis. British Medical Bulletin (London) 24:59-70

83. Kovacs K 1969 Necrosis of anterior pituitary in humans. Neuroendocrinology (Basel) 4:179-199

84. Kovacs K 1972 Adenohypophysial necrosis in routine autopsies. Endokrinologie 60:309-316

85. Kovacs K, Bilbao JM 1974 Adenohypophysial necrosis in respirator maintained patients. Pathol Microbiol 41:275-282

86. Sheehan HL 1937 Post-partum necrosis of the anterior pituitary. Journal of Pathology and Bacteriology (London) 45:189-214

87. Cardoso ER, Peterson EW 1984 Pituitary apoplexy: A review. Neurosurgery (Baltimore) 14:363-373

88. Voigt C, Saeger W, Gerigk Ch, Lüdecke DK 1988 Amyloid in pituitary adenomas. Pathology, Research and Practice (Stuttgart) 183:555-557

89. Landolt AM, Kleihues P, Heitz PhU 1987 Amyloid deposits in pituitary adenomas. Differentiation of two types. Archives of Pathology and Laboratory Medicine 111:453-458

90. Bilbao JM, Horvath E, Hudson AR, Kovacs K 1975 Pituitary adenoma producing amyloidlike substance. Archives of Pathology and Laboratory Medicine 99:411-415

91. Bilbao JM, Kovacs K, Horvath E, et al. 1975 Pituitary melanocorticotrophinoma with amyloid deposition. Journal of Canadian Scientific Neurology 2:199-202

92. Mori H, Mori S, Saitoh Y, Moriwaki K, Iida S, Matsumoto K 1985 Growth hormone-producing pituitary adenoma with crystal-like amyloid immunohistochemically positive for growth hormone. Cancer (Philadelphia) 55:96-102

93. Livadas DP, Sofroniadou K, Souvatzoglou A, Boulanger V, Siafaka L, Koutras DA 1979 Pituitary and thyroid insufficiency in thalassaemic haemosiderosis. Clinical Endocrinology 20:435-443

94. Kletzky OA, Costin G, Marrs RP, Bernstein G, March CM, Mishell DRJr 1979 Gonadotropin insufficiency in patients with thalAssemia major. Journal of Clinical Endocrinology and Metabolism (Baltimore) 48:901-905

95. Horvath E 1988 Pituitary hyperplasia. Pathology, Research and Practice (Stuttgart) 183:623-625

96. Saeger W, Lüdecke DK 1983 Pituitary hyperplasia. Definition, light and electron microscopical structures and significance in surgical specimens. Virchows Archiv A,Pathological Anatomy and Histopathology (Berlin) 399:277-287

97. Zimmerman D, Young WF, Jr., Ebersold MJ, Scheithauer BW, Kovacs K, Horvath E, Whitaker MD, Eberhardt NL, Downs TR, Frohman LA 1993 Congenital gigantism due to growth hormone-releasing hormone excess and pituitary hyperplasia with adenomatous transformation. Journal of Clinical Endocrinology and Metabolism (Baltimore) 76:216-222

98. Kubota T, Hayashi M, Kabuto M, shirasaki N, Aradachi H, Miyanaga K, Miyabo S 1992 Corticotroph cell hyperplasia in a patient with Addison disease: case report. Surgical Neurology 37:441-447 99. McKeever PE, Koppelman MCS, Metcalf D, Quindlen E, Kornblith PL, Strott CA, Howard R, Smith BH 1982 Refractory Cushing's disease caused by multinodular ACTH-cell hyperplasia. Journal of Neuropathology and Experimental Neurology 41:490-499

100. McNicol AM 1981 Patterns of corticotropic cells in the adult human pituitary in Cushing's disease. Diagnostic Histopathology 4:335-341

101. Kovacs K, Horvath E, Rewcastle NB, Ezrin C 1980 Gonadotroph cell adenoma of the pituitary in a woman with long-standing hypogonadism. Archives of Gynecology (Berlin) 229:57-65

102. Nicolis G, Shimshi M, Allen C, Halmi NS, Kourides IA 1988 Gonadotropin-producing pituitary adenoma in a man with long-standing primary hypogonadism. Journal of Clinical Endocrinology and Metabolism (Baltimore) 66:237-241

103. Sano T, Asa SL, Kovacs K 1988 Growth hormone-releasing hormone-producing tumors: clinical, biochemical, and morphological manifestations. Endocrine Reviews 9:357-373

104. Ezzat S, Asa SL, Stefaneanu L, Whittom R, Smyth HS, Horvath E, Kovacs K, Frohman LA 1994 Somatotroph hyperplasia without pituitary adenoma associated with a long standing growth hormone-releasing hormone-producing bronchial carcinoid. Journal of Clinical Endocrinology and Metabolism (Baltimore) 78:555-560

105. Asa SL, Scheithauer BW, Bilbao JM, Horvath E, Ryan N, Kovacs K, Randall RV, Laws ER, Jr., Singer W, Linfoot JA, Thorner MO, Vale W 1984 A case for hypothalamic acromegaly: a clinicopathological study of six patients with hypothalamic gangliocytomas producing growth hormone-releasing factor. Journal of Clinical Endocrinology and Metabolism (Baltimore) 58:796-803

106. Kovacs K, Horvath E, Thorner MO, Rogol AD 1984 Mammosomatotroph hyperplasia associated with acromegaly and hyperprolactinemia in a patient with the McCune-Albright syndrome. Virchows Archiv A, Pathological Anatomy and Histopathology (Berlin) 403:77-86

107. Weinstein LS, Shenker A, Gejman PV, Merino MJ, Friedman E, Spiegel AM 1991 Activating mutations of the stimulatory G protein in the McCune-Albright syndrome. New England Journal of Medicine 325:1688-1695

108. Moran A, Asa SL, Kovacs K, Horvath E, Singer W, Sagman U, Reubi JC, Wilson CB, Larson R, Pescovitz OH 1990 Gigantism due to pituitary mammosomatotroph hyperplasia. New England Journal of Medicine 323:322-327

109. Peillon F, Dupuy M, Li JY, Kujas M, Vincens M, Mowszowicz I, Derome P 1991 Pituitary enlargement with suprasellar extension in functional hyperprolactinemia due to lactotroph hyperplasia: A pseudotumoral disease. Journal of Clinical Endocrinology and Metabolism (Baltimore) 73:1008-1015

110. Jay V, Kovacs K, Horvath E, Lloyd RV, Smyth HS 1991 Idiopathic prolactin cell hyperplasia of the pituitary mimicking prolactin cell adenoma: a morphological study including immunocytochemistry, electron microscopy, and in situ hybridization. Acta Neuropathologica (Berlin) 82:147-151

111. Khalil A, Kovacs K, Sima AAF, Burrow GN, Horvath E 1984 Pituitary thyrotroph hyperplasia mimicking prolactin-secreting adenoma. Journal of Endocrinological Investigation (Milano) 7:399-404

112. Chan AW, MacFarlane IA, Foy PM, Miles JB 1990 Pituitary enlargement and hyperprolactinaemia due to primary hypothyroidism: errors and delays in diagnosis. British Journal of Neurosurgery 4:107-112

1. Grubb MR, Chakeres D, Malarkey WB 1987 Patients with primary hypothyroidism presenting as prolactinomas. American Journal of Medicine 83:765-769

114.Ezzat S, Asa SL, Couldwell WT, Barr CE, Dodge WE, Vance ML, McCutcheon IE 2004 The prevalence of pituitary adenomas: a systematic review. Cancer 101:613-9

115. Asa SL, Ezzat S 2002 The pathogenesis of pituitary tumours. Nature Reviews Cancer 2:836-49.

116. Ezzat S, Asa SL 2006 Mechanisms of Disease: the pathogenesis of pituitary tumors. Nature Clinical Practice Endocrinology and Metabolism 2:220-230

117. Ezzat S, Asa SL 1996 Syndromes of multiple endocrine neoplasia and hyperplasia. In: Kovacs K, Asa SL (eds). Functional Endocrine Pathology.Blackwell Science, Boston:

118. Scheithauer BW, Laws ER, Jr., Kovacs K, Horvath E, Randall RV, Carney JA 1987 Pituitary adenomas of the multiple endocrine neoplasia type I syndrome. Seminars in Diagnostic Pathology 4:205-211

119. Rasmussen P, Lindholm J 1979 Ectopic pituitary adenomas. Clinical Endocrinology (Oxf) 11:69-74

120. Dyer EH, Civit T, Abecassis J-P, Derome PJ 1994 Functioning ectopic supradiaphragmatic pituitary adenomas. Neurosurgery (Baltimore) 43:529-532

121. Anand NK, Osborne CM, Harkey HLI 1993 Infiltrative clival pituitary adenoma of ectopic origin. Head and Neck Surgery 108:178-183

122. Coire CI, Horvath E, Kovacs K, Smyth HS, Ezzat S 1997 Cushing's syndrome from an ectopic pituitary adenoma with peliosis: A histological, immunohistochemical and ultrastructural study and review of the literature. Endocrine Pathology 8:65-74

123. Kleinschmidt-De Masters BK, Winston KR, Rubinstein D, Samuels MH 1990 Ectopic

pituitary adenomas of the third ventricle. Case report. Journal of Neurosurgery (Baltimore) 72:139-142

124. Lloyd RV, Chandler WF, Kovacs K, Ryan N 1986 Ectopic pituitary adenomas with normal anterior pituitary glands. American Journal of Surgical Pathology 108:546-552

125. Matsumura A, Meguro K, Doi M, Tsurushima H, Tomono Y 1990 Suprasellar ectopic pituitary adenoma: case report and review of the literature. Neurosurgery (Baltimore) 26:681-685

126. Shenker Y, Lloyd RV, Weatherbee L, Port FK, Grekin RJ, Barkan AL 1986 Ectopic prolactinoma in a patient with hyperparathyroidism and abnormal sellar radiography. Journal of Clinical Endocrinology and Metabolism (Baltimore) 62:1065-1069

127. Slonim SM, Haykal HA, Cushing GW, Freidberg SR, Lee AK 1993 MRI appearances of an ectopic pituitary adenoma: case report and review of the literature. Neuroradiology 35:546-548

128. Hardy J 1969 Transphenoidal microsurgery of the normal and pathological pituitary. Clinical Neurosurgery. Proceedings of the Congress of Neurological Surgeons, 1968. Williams and Wilkins, Baltimore: 185-217

129.DeLellis RA, Lloyd RV, Heitz PU, Eng C (Editors) 2004 Pathology and Genetics of Tumours of Endocrine Organs. World Health Organization Classification of Tumours, IARC Press, Lyon

130. Ezzat S, Forster MJ, Berchtold P, Redelmeier DA, Boerlin V, Harris AG 1994 Acromegaly. Clinical and biochemical features in 500 patients. Medicine 73:233-240

131. Ezzat S, Snyder PJ, Young WF, Boyajy LD, Newman C, Klibanski A, Molitch ME, Boyd AE, Sheeler L, Cook DM, Malarkey WB, Jackson I, Vance ML, Thorner MO, Barkan AL, Frohman LA, Melmed S 1992 Octreotide treatment of acromegaly. A randomized, multicenter study. Annals of Internal Medicine 117:711-718

132. Bhayana S, Booth G, Asa SL, Kovacs K, Ezzat S 2005 The implication of somatotroph adenoma phenotype to somatostatin analog responsiveness in acromegaly. Journal of Clinical Endocrinology and Metabolism (Baltimore) 90:6290-5.

133. Ezzat S, Horvath E, Harris AG, Kovacs K 1994 Morphological effects of octreotide on growth hormone-producing pituitary adenomas. Journal of Clinical Endocrinology and Metabolism (Baltimore) 79:113-118

- 1. Gillam MP. Molitch ME, Lombardi G, Colao A 2006 Advances in the treatment of prolactinomas. Endocrine Reviews 27:485-534
- 2. Grossman A, Besser GM 1985 Prolactinomas. British Medical Journal 290:182-184

136. Burrow GN, Wortzman G, Rewcastle NB, Holgate RC, Kovacs K 1981 Microadenomas of the pituitary and abnormal sellar tomograms in an unselected autopsy series. New England

Journal of Medicine 304:156-158

137. Grisoli F, Vincentelli F, Jaquet P, Guibout M, Hassoun J, Farnarier P 1980 Prolactin secreting adenoma in 22 men. Surgical Neurology 13:241-247

138. Delgrange E, Trouillas J, Maiter D, Donckier J, Tourniaire J 1997 Sex-related difference in the growth of prolactinomas: a clinical and proliferation marker study. J Clin Endocrinol Metab 82:2102-2107

139. Tindall GT, Kovacs K, Horvath E, Thorner MO 1982 Human prolactin-producing adenomas and bromocriptine: a histological, immunocytochemical, ultrastructural and morphometric study. Journal of Clinical Endocrinology and Metabolism (Baltimore) 55:1178-1183

140. Asa SL, Kovacs K, Horvath E, Singer W, Smyth HS 1992 Hormone secretion in vitro by plurihormonal pituitary adenomas of the acidophil cell line. Journal of Clinical Endocrinology and Metabolism (Baltimore) 75:68-75

141. Beck-Peccoz P, Brucker-Davis F, Persani L, Smallridge RC, Weintraub BD 1996 Thyrotropin-secreting pituitary tumors. Endocrine Reviews 17:610-638

142. Felix IA, Horvath E, Kovacs K 1981 Massive Crooke's hyalinization in corticotroph cell adenomas of the human pituitary. A histological, immunocytological and electron microscopic study of three cases. Acta Neurochirurgica (Wien) 58:235-243

143. Franscella S, Favod-Coune C-A, Pizzolato G, Asa SL, Gaillard R, Berney J, Philippe J 1991 Pituitary corticotroph adenoma with Crooke's hyalinization. Endocrine Pathology 2:111-116

144. Neumann PE, Horoupian DS, Goldman JE, Hess MA 1984 Cytoplasmic filaments of Crooke's hyaline change belong to the cytokeratin class. An immunocytochemical and ultrastructural study. American Journal of Pathology 116:214-222

145. Snyder PJ 1985 Gonadotroph cell adenomas of the pituitary. Endocrine Reviews 6:552-563

146. Daneshdoost L, Gennarelli TA, Bashey HM, Savino PJ, Sergott RC, Bosley TM, Snyder PJ 1991 Recognition of gonadotroph adenomas in women. New England Journal of Medicine 324:589-594

147. Asa SL, Gerrie BM, Kovacs K, Horvath E, Singer W, Killinger DW, Smyth HS 1988 Structure-function correlations of human pituitary gonadotroph adenomas in vitro. Laboratory Investigation (Baltimore) 58:403-410

148. Asa SL, Gerrie BM, Singer W, Horvath E, Kovacs K, Smyth HS 1986 Gonadotropin secretion in vitro by human pituitary null cell adenomas and oncocytomas. Journal of Clinical Endocrinology and Metabolism (Baltimore) 62:1011-1019

1. Horvath E, Kovacs K 1984 Gonadotroph adenomas of the human pituitary: sex-related fine-structural dichotomy. A histologic, immunocytochemical, and electron-microscopic study of 30 tumors. American Journal of Pathology 117:429-440

149a.Sano T, Mader R, Asa SL, Rong QZ, Hino A, Yamada S 2003 The "honeycomb Golgi" in pituitary adenomas: Not a marker of gonadotroph adenomas. Endocrine Pathology 14:363-368.

150. Apel RL, Wilson RJ, Asa SL 1994 A composite somatotroph-corticotroph pituitary adenoma. Endocrine Pathology 5:240-246

151. Syro LV, Horvath E, Kovacs K 2000 Double adenoma of the pituitary: a somatotroph adenoma colliding with a gonadotroph adenoma. Journal of Endocrinological Investigation (Milano) 23:37-41

152. Horvath E, Kovacs K, Smyth HS, Cusimano M, Singer W 2005 Silent adenoma subtype 3 of the pituitary–immunohistochemical and ultrastructural classification: a review of 29 cases. Ultrastructural Patholology 29:511-524.

153. Saeger W, Lübke D 1996 Pituitary carcinomas. Endocrine Pathology 7:21-35

154. Cai WY, Alexander JM, Hedley-Whyte ET, Scheithauer BW, Jameson JL, Zervas NT, Klibanski A 1994 Ras mutations in human prolactinomas and pituitary carcinomas. Journal of Clinical Endocrinology and Metabolism (Baltimore) 78:89-93

155. Pei L, Melmed S, Scheithauer B, Kovacs K, Prager D 1994 H-ras mutations in human pituitary carcinoma metastases. Journal of Clinical Endocrinology and Metabolism (Baltimore) 78:842-846

156. Thapar K, Scheithauer BW, Kovacs K, Pernicone PJ, Laws ER, Jr. 1996 p53 expression in pituitary adenomas and carcinomas: Correlation with invasiveness and tumor growth fractions. Neurosurgery (Baltimore) 38:765-771

157. Frost AR, Tenner S, Tenner M, Rollhauser C, Tabbara SO 1995 ACTH-producing pituitary carcinoma presenting as the cauda equina syndrome. Archives of Pathology and Laboratory Medicine 119:93-96

158. Kouhara H, Tatekawa T, Koga M, Hiraga S, Arita N, Mori H, Sato B 1992 Intracranial and intraspinal dissemination of an ACTH-secreting pituitary tumor. Endocrinologia Japonica (Tokyo) 39:177-184

159. Mixson AJ, Friedman TC, Katz DA, Feuerstein IM, Taubenberger JK, Colandrea JM, Doppman JL, Oldfield EH, Weintraub BD 1993 Thyrotropin-secreting pituitary carcinoma. Journal of Clinical Endocrinology and Metabolism (Baltimore) 76:529-533

160. Petterson T, MacFarlane IA, MacKenzie JM, Shaw MDM 1992 Prolactin secreting pituitary carcinoma. Journal of Neurology, Neurosurgery and Psychiatry 55:1205-1206

161. Stewart PM, Carey MP, Graham CT, Wright AD, London DR 1992 Growth hormone secreting pituitary carcinoma: a case report and literature review. Clinical Endocrinology (Oxf) 37:189-195

162. Walker JD, Grossman A, Anderson JV, Ur E, Trainer PJ, Benn J, Lowy C, Sönksen PH, Plowman PN, Lowe DG, Doniach I, Wass JAH, Besser GM 1993 Malignant prolactinoma with extracranial metastases: a report of three cases. Clinical Endocrinology (Oxf) 38:411-419

163. Luzi P, Miracco C, Lio R, Malandrini A, Piovani S, Venezia SG, Tosi P 1987 Endocrine inactive pituitary carcinoma metastasizing to cervical lymph nodes: a case report. Human Pathology (Philadelphia) 18:90-92

164. Beauchesne P, Trouillas J, Barral F, Brunon J 1995 Gonadotropic pituitary carcinoma: case report. Neurosurg 37:810-815

165. Zahedi A, Booth GL, Smyth HS, Farrell WE, Clayton RN, Asa SL, Ezzat S 2001 Distinct clonal composition of primary and metastatic adrencorticotrophic hormone-producing pituitary carcinoma. Clinical Endocrinology (Oxf) 55:549-556

166. Banna M 1976 Cranopharyngioma: based on 160 cases. British Journal of Radiology 49:206-223

167. Cusimano MD, Kovacs K, Bilbao JM, Tucker WS, Singer W 1988 Suprasellar craniopharyngioma associated with hyperprolactinemia, pituitary lactotroph hyperplasia, and microprolactinoma. Case report. Journal of Neurosurgery (Baltimore) 69:620-623

168. Wheatley T, Clark JDA, Stewart S 1986 Craniopharyngioma with hyperprolactinaemia due to a prolactinoma. Journal of Neurology, Neurosurgery and Psychiatry 49:1305-1307

169. Laws ER, Jr. 1992 Craniopharyngioma: Diagnosis and treatment. The Endocrinologist 2:184-188

170. Scheithauer BW 1998 The hypothalamus and neurohypophysis. In: Kovacs K, Asa SL (eds). Functional Endocrine Pathology.Blackwell Scientific Publications, Inc., Boston:171-246

171. Puchner MJA, Lüdecke DK, Saeger W, Riedel M, Asa SL 1995 Gangliocytomas of the sellar region – a review. Experimental and Clinical Endocrinology 103:129-149

171a Harris PE, Perry L, Chard T, Chaundry L, Crooke BA, Touzel R, Coates P, Lowe DG, Afshar F, Wass JAH, Besser GM 1988. Immunoreactive chrionic gonadotrophin from the cyst fluid and CSF of patients with craniopharyngioma. Clinical Endocrinology 29: 5-3-508.

171b. Sekine S, Shibata T, Kokubu A, Morishita Y, Noguchi M, Nakanishi Y, Sakamoto M, Hirohashi S 2002 Craniopharyngiomas of adamantinomatous type harbor beta-catenin gene mutations. American Journal of Pathology 161:1997-2001

172. Rossi ML, Bevan JS, Esiri MM, Hughes JT, Adams CBT 1987 Pituicytoma (pilocytic astrocytoma). Journal of Neurosurgery (Baltimore) 67:768-772

173. Kitanaka C, Shitara N, Nakagomi T, Nakamura H, Genka S, Nakagawa K, Akanuma A, Aoyama H, Takakura K 1989 Postradiation astrocytoma. Report of two cases. Journal of Neurosurgery (Baltimore) 70:469-474

174. Zampieri P, Zorat PL, Mingrino S, Soattin GB 1989 Radiation-associated cerebral gliomas. A report of two cases and review of the literature. Journal of Neurological Sciences 33:271-279

175. Grisoli F, Vincentelli F, Raybaud C, Harter M, Guibout M, Baldini M 1983 Intrasellar meningioma. Surgical Neurology 20:36-41

176. Slavin MJ, Weintraub J 1987 Suprasellar meningioma with intrasellar extension simulating pituitary adenoma. Archives of Ophthalmology 105:1488-1489

177. Spallone A 1982 Meningioma as a sequel of radiotherapy for pituitary adenoma. Neurochirurgia (stuttgart) 25:68-72

178. Sridhar K, Ramamurthi B 1989 Intracranial meningioma subsequent to radiation for a pituitary tumor: case report. Neurosurgery (Baltimore) 25:643-645

179. Kasantikul V, Shuangshoti S, Phonprasert C 1988 Intrasellar meningioma after radiotherapy for prolactinoma. Journal of the Medical Association of Thailand 71:524-527

180. Buley ID, Gatter KC, Kelly PMA, Heryet A, Millard PR 1988 Granular cell tumours revisited. An immunohistological and ultrastructural study. Histopathology 12:263-274

181. Cone L, Srinivasan M, Romanul FCA 1990 Granular cell tumor (choristoma) of the neurohypophysis: Two cases and a review of the literature. American Journal of Neuroradiology 11:403-406

182. Nishioka H, Ii K, Llena JF, Hirano A 1991 Immunohistochemical study of granular cell tumors of the neurohypophysis. Virchows Archiv B Cell Pathology (Berlin) 60:413-417

183. Mathews W, Wilson CB 1974 Ectopic intrasellar chordoma. Journal of Neurosurgery (Baltimore) 39:260-263

184. Ishige N, Ito C, Saeki N, Oka N 1985 Neurinoma with intrasellar extension: a case report. Neurological Surgery 13:79-84

185. Perone TP, Robinson B, Holmes SM 1984 Intrasellar schwannoma: case report. Neurosurgery (Baltimore) 14:71-73

186. Wilberger JE, Jr. 1989 Primary intrasellar schwannoma: case report. Surgical Neurology 32:156-158

187. Kleihues P, Burger PC, Scheithauer BW 1993 Histological Typing of Tumours of the Central Nervous System. World Health Organization International Histological Classification of Tumours.Springer-Verlag, Berlin, 2 edn

188. Rueda-Pedraza ME, Heifetz SA, Sesterhenn IA, Clark GB 1987 Primary intracranial germ cell tumors in the first two decades of life. A clinical, light-microscopic, and immunohistochemical analysis of 54 cases. Perspectives in Pediatric Pathology 10:160-207

189. Jennings MT, Gelman R, Hochberg F 1985 Intracranial germ-cell tumors: natural history and pathogenesis. Journal of Neurosurgery (Baltimore) 63:155-167

190. Furukawa F, Haebara H, Hamashima Y 1986 Primary intracranial choriocarcinoma arising from the pituitary fossa. Report of an autopsy case with literature review. Acta Pathologica Japonica (Tokyo) 36:773-781

191. Kageyama N, Kobayashi T, Kida Y, Yoshida J, Kato K 1987 Intracranial germinal tumors. Progress in Experimental Tumor Research 30:255-267

192. Sheehan T, Cuthbert RJG, Parker AC 1989 Central nervous system involvement in haematological malignancies. Clinical and Laboratory Haematology 11:331-338

193. Singh VP, Mahapatra AK, Dinde AK 1993 Sellar-suprasellar primary malignant lymphoma: Case report. Indian Journal of Cancer 30:88-91

194. Samaratunga H, Perry-Keene D, Apel RL 1997 Primary lymphoma of the pituitary gland: a neoplasm of acquired MALT? Endocrine Pathology 8:335-341

195. Warnke R, Dorfman R, Weiss L, Cleary M, Chan J 1995 Tumors of the Lymphoid System. Atlas of Tumor Pathology, Third Series, Fascicle.Armed Forces Institute of Pathology, Washington, D.C.

196. Ornvold K, Ralfkiaer E, Carstensen H 1990 Immunohistochemical study of the abnormal cells in Langerhans cell histiocytosis (Histiocytosis X). Virchows Archiv A, Pathological Anatomy and Histopathology (Berlin) 416:403-410

197. Chang WH, Khosla VK, Radotra BD, Kak VK 1991 Large cavernous hemangioma of the pituitary fossa: a case report. British Journal of Neurosurgery 5:627-629

198. Sansone ME, Liwnicz BH, Mandybur TI 1980 Giant pituitary cavernous hemangioma. Case report. Journal of Neurosurgery (Baltimore) 53:124-126

199. Asa SL, Kovacs K, Horvath E, Ezrin C, Weiss MH 1984 Sellar glomangioma. Ultrastructural Pathology (New York) 7:49-54

200. Dan NG, Smith DE 1975 Pituitary hemangioblastoma in a patient with von Hippel-Lindau disease. Journal of Neurosurgery (Baltimore) 42:232-235

201. Miki K, Kawamoto K, Kawamura Y, Matsumura H, Asada Y, Hamada A 1987 A rare case of Maffucci's syndrome combined with tuberculum sellae enchondroma, pituitary adenoma and thyroid adenoma. Acta Neurochirurgica (Wien) 87:79-85

202. Dutton J 1978 Intracranial solitary chondroma. Case report. Journal of Neurosurgery (Baltimore) 49:460-463

203. Angiari P, TTorcia E, Botticelli RA, villani M, Merli GA, Crisi G 1987 Ossifying parasellar chondroma. Case report. Journal of Neurological Sciences 31:59-63

204. Sindou M, Daher A, Vighetto A, Goutelle A 1989 Chondrosarcome parasellaire: rapport d'un cas opéré par voie ptériono-temporale et revue de la littérature. Neuro-Chirurgie 35:186-190

205. Viswanathan R, Jegathraman AR, Ganapathy K, Bharati AS, Govindan R 1987 Parasellar chondromyxofibroma with ipsilateral total internal carotid artery occlusion. Surgical Neurology 28:141-144

206. Wolfe JTI, Scheithauer BW, Dahlin DC 1983 Giant-cell tumor of the sphenoid bone. Review of 10 cases. Journal of Neurosurgery (Baltimore) 59:322-327

207. Bots GTAM, Tijssen CC, Wijnalda D, Teepen JLJM 1988 Alveolar soft part sarcoma of the pituitary gland with secondary involvement of the right cerebral ventricle. British Journal of Neurosurgery 2:101-107

208. Amine ARC, Sugar O 1976 Suprasellar osteogenic sarcoma following radiation for pituitary adenoma. Case report. Journal of Neurosurgery (Baltimore) 44:88-91

209. Ahmad K, Fayos JV 1978 Pituitary fibrosarcoma secondary to radiation therapy. Cancer (Philadelphia) 42:107-110

210. Piatt JH, Blue JM, Schold SC, Burger PC 1983 Glioblastoma multiforme after radiotherapy for acromegaly. Neurosurgery (Baltimore) 13:85-89

211. McCormick PC, Post KD, Kandji AD, Hays AP 1989 Metastatic carcinoma to the pituitary gland. British Journal of Neurosurgery 3:71-79

212. Roessmann U, Kaufman B, Friede RL 1970 Metastatic lesions in the sella turcica and pituitary gland. Cancer (Philadelphia) 25:478-480

213. van Seters AP, Bots GTAM, Van Dulken H, Luyendijk W, Vielvoye GJ 1985 Metastasis of an occult gastric carcinoma suggesting growth of a prolactinoma during bromocriptine therapy: a case report with a review of the literature. Neurosurgery (Baltimore) 16:813-817

214. de la Monte SM, Hutchins GM, Moore GW 1984 Endocrine organ metastases from breast carcinoma. American Journal of Pathology 114:131-136

215. Max MB, Deck MDF, Rottenberg DA 1981 Pituitary metastasis: incidence in cancer patients and clinical differentiation from pituitary adenoma. Neurology (Cleveland) 31:998-1002

216. Molinatti PA, Scheithauer BW, Randall RV, Laws ER, Jr. 1985 Metastasis to pituitary adenoma. Archives of Pathology and Laboratory Medicine 109:287-289

217. Post KD, McCormick PC, Hays AP, Kankji AD 1988 Metastatic carcinoma to pituitary adenoma. Report of two cases. Surgical Neurology 30:286-292

218. Ramsay JA, Kovacs K, Scheithauer BW, Ezrin C, Weiss MH 1988 Metastatic carcinoma to pituitary adenomas: a report of two cases. Experimental and Clinical Endocrinology 92:69-76

219. Gartman JJ, Jr., Powers SK, Fortune M 1989 Pseudotumor of the sellar and parasellar areas. Neurosurgery (Baltimore) 24:896-901

220. Dussault J, Plamondon C, Volpe R 1969 Aneurysms of the internal carotid artery simulating pituitary tumours. Canadian Medical Association Journal 101:51-56

221. Durham LH, Mackenzie IJ, Miles JB 1988 Transphenoidal meningohydroencephalocoele. British Journal of Neurosurgery 2:407-410

222. Schochet SS, Jr., McCormick WF, Halmi NS 1974 Salivary gland rests in the human pituitary. Light and electron microscopical study. Archives of Pathology 98:193-200

223. Kato T, Aida T, Abe H, Miyamachi K, Hida K, Taneda M, Ogata A 1988 Ectopic salivary gland within the pituitary gland. Case report. Neurologia Medico-Chirurgica 28:930-933