

OBESITEXT Chapter 3.1:– The regulation of energy intake

Andrea L. Tracy¹, PhD, Grace E. Hazeltine¹, BA, Colin J.M. Wee¹, BA, & Stephen C. Benoit², PhD

¹Dept. of Psychology, Grinnell College, Grinnell, IA

²Dept. of Psychiatry, University of Cincinnati, Cincinnati, OH

Take-Home Points

- Regulation of food intake occurs homeostatically via activation of peripheral signals in the GI tract and adipose tissue, which directly monitor incoming nutrient and nutrient stores, and central systems (e.g., the hypothalamus), which receive these signals and alter behavioral and metabolic output to balance energy intake with need
- These signals engage in significant interaction with one another, often by making other signals more or less effective (e.g., increased levels of the adipose hormone leptin due to greater fat stores makes the satiety signal CCK more effective decreasing meal size).
- In addition to the hypothalamus, the hippocampus is involved in regulating food intake by cognitive mechanisms including explicit recall of prior meals and associative learning about the relationship of feelings of hunger/satiety to the consequences of eating.
- These signals act via numerous behavioral mechanisms, including altering meal initiation, termination and size, increasing or decreasing the motivation to seek and consume food, causing subjective feelings of hunger or satiety, or changing the perceived reward value of food.
- Obesity, or chronic consumption of a high-fat, energy-dense diet, can lead to changes in the sensitivity to these signals resulting in decreased ability to regulate and may contribute to the persistence of obesity. Cognitive function and sensitivity to the rewarding aspects of food are also affected by chronic obesity.

Introduction

Energy balance requires that an organism match caloric intake relatively precisely with caloric expenditure. In humans, an error of only +11 kcal/day results in a one pound weight gain over the course of a year. Over the past 40 years, the average body weight of American adults has increased at rate of less than that one pound per year, but the steady increase has yielded an increase of an average of 3 BMI points, bringing the average adult from a healthy weight into the overweight category (1). This increase brings with it a significantly increased risk of a number of health problems, including type 2 diabetes, high blood pressure, and cardiovascular disease and has a total financial cost estimated at \$139 billion per year (2). In attempting to identify potential biological causes and treatments for this widely-occurring disorder, it is critical to understand the mechanisms which regulate energy homeostasis. In this chapter, we will review both peripheral and central signaling mechanisms relating to the food intake side of the equation, including how these signals function with respect to specific aspects of food intake-related behavior, and a brief overview of how this system may become dysregulated during states of chronic overconsumption and obesity.

Environmental Signals

There are a variety of external factors that play a significant role in food intake, including social situations, time cues, food-related stimuli (e.g., sight, smell) and other learned information. While it is evident that these types of signals can have a definite impact on when to consume a meal, what foods to choose and how much to eat, the focus of this chapter will be on the molecular mechanisms involved in controlling these ingestive behaviors.

Peripheral Signals

Gastric Mechanoreceptors

After food is ingested, it moves into the gastrointestinal tract where the volume and the nutritive content of the meal is detected via mechanical and chemosensory mechanisms. The results of sham feeding experiments indicate clearly that detection of food in the gastrointestinal (GI) tract plays a large role in determining the amount consumed. In these studies, animals with open gastric fistulas which allow food to drain out of the stomach consume much larger volumes than animals consuming food normally, an effect which can be overcome by concurrently infusing nutritive solutions directly into the duodenum (3, 4). Gastric mechanoreceptors are located on vagal afferent and splanchnic nerve fibers and detect food volume by responding to stretch or pressure in the walls of the stomach (5, 6). Experiments in rats using pyloric occlusion to prevent contents from emptying into the intestines have demonstrated that satiety, as indicated by reduction in subsequent food intake, can occur based on gastric signals, that this is due predominantly to food volume, rather than caloric content, and that this effect is dependent on an intact vagus (7-9). However, it appears that the volumes required to reduce food intake are substantially greater than the volumes generally consumed in a single meal. Further, under the majority of self-controlled feeding conditions in rodents, intake was not significantly altered by pyloric occlusion (10, 11), indicating that, while gastric distension can act as a satiety signal, it may not be an important regulator during normal feeding situations. Although the idea that gastric distension may contribute to meal termination is consistent with data supporting a volumetric control of food intake, the data on gastric mechanoreceptors and satiety suggest that other mechanisms are likely at work in this phenomenon.

Gastrointestinal Satiety Signals

Although the stomach is thought to be primarily responsive to food volume, nutrient entry into the stomach also induces the release of gastrin releasing peptide (GRP), a member of the bombesin-like peptide family. GRPs (including GRP-10, -27 and -29) in humans and animals act as satiety signals by reducing meal size, prolonging time to begin the next meal and enhancing the satiating effects of a meal (12,13).

The intestinal tract is highly sensitive to the caloric content of ingested foods. Beginning in the duodenum, the detection of nutrients activates the release of a number of peptides, often termed “satiety signals”, which act primarily to terminate consumption of a meal. The most well known of these is cholecystokinin (CCK), an octapeptide that is released from the duodenum and, to a lesser extent, the ileum in response to nutrients (14). CCK activates receptors on the vagus nerve which terminates in the hindbrain at the nucleus of the solitary tract (NTS). As nutrients enter into and move through the GI tract, peptide YY (PYY) and glucagon-like peptide-1 (GLP-1) are secreted from the ileum and colon, while apolipoprotein A-IV (Apo A-IV) is synthesized in response to intestinal fat absorption (15-17). CCK and other GI peptides are differentially responsive to the macronutrient composition of a meal. CCK and gastrin are released more readily by protein ingestion, which is confirmed by the finding that protein hydrolysates directly stimulate CCK receptors in enteroendocrine cells (18,19). Consumption of carbohydrates and fats yields greater GLP-1 release, whereas Apo A-IV synthesis is induced exclusively by fat absorption and PYY is most responsive to protein and fats (20-22). In obese individuals, PYY is more responsive to fats than carbohydrates (23) and, in humans, exogenous administration of PYY₃₋₃₆ increases both ratings of satiety and plasma free fatty acids (24). Since most meals consist of multiple nutrient components, this allows for the integration of nutrient and caloric information by GI signals, and these varying profiles of satiety signal activation may contribute to differences in the relative satiety value of the macronutrients (25,26), as well as possibly affecting macronutrient selection and intake at later meals.

All of these GI “satiety signals” reduce food intake when administered to animals either systemically or centrally, and when administered peripherally in humans (24,27-34), although it should be noted that there is some controversy surrounding the efficacy of the active form of PYY₃₋₃₆ to reduce food intake

and body weight in humans and non-human animal models (35,36). These peptides effect reductions in food intake primarily by acting to terminate the current meal, although longer-term effects have been suggested for GLP-1 and PYY (37). The observation that antagonists of CCK and GLP-1 receptors results in increased food intake (38-40) more strongly indicates that these hormones function endogenously to control feelings of satiety and meal termination. The idea that these hormones are the natural mechanism for ending a meal is also supported by the observation that meal ingestion, infusion of calories into the GI tract and exogenous administration of several of these peptides, including CCK and bombesin/GRP, all result in animals displaying a similar set of behaviors, termed the "behavioral satiety sequence" (41-43). However, genetic deletion of these peptides or their receptors yields mixed results. For example, PYY knockout mice and OLETF rats, which lack the CCK-1 receptor (CCK-1R) due to a spontaneous mutation, display marked hyperphagia and obesity (44,45). However, rats genetically manipulated to possess the specific CCK-1R-null gene that characterizes the spontaneous OLETF phenotype do not display the same hyperphagia and obesity, suggesting that the genetic mutation affecting CCK-1R is not the sole contributor to these characteristics in OLETF rats (46). Additionally, GLP-1R-null mice display normal feeding behaviors and body weight (47). Of course, the majority of these genetic studies employ techniques that typically result in absence of the peptide or receptor throughout development allowing the function to be taken over by other systems. Overall, we can conclude that GI peptides play an important role in meal size and meal termination, with the endogenous function of some of these peptides being critical regulation of energy intake under normal ingestive conditions, while the role of others may be redundant with or compensated for by other systems.

Pancreatic Satiety Hormones

Outside of the GI tract, the pancreas also secretes peripheral meal-related hormones that act to reduce food intake. Insulin and amylin are co-secreted from pancreatic β -cells in proportion to the amount of food consumed (48,49). While the function of insulin seems to relate more to the long-term regulation of body adiposity (see below), amylin serves as a short-term signal that acts to reduce food intake by decreasing meal size (50). Exogenous, peripherally administered amylin reduces food intake, while systemic antagonists have the opposite effect, again indicating an endogenous role for this hormone in satiety (51-53). Glucagon is secreted from pancreatic A-cells very rapidly following meal onset, particularly meals high in protein, and acts via the liver to limit meal size in rodents and humans, an effect which can be reversed by administration of a glucagon-specific antibody (54-59).

Ghrelin

To date there is only one identified orexigenic, or appetite stimulating, gut peptide. Ghrelin is an endogenous ligand for the growth hormone secretagogue receptor (GHSR) that is synthesized in and secreted from gastric epithelium (60). Ghrelin produced by neurons in the hypothalamus also exerts orexigenic effects on appetite (61). Administration of exogenous ghrelin increases food intake in both humans and non-human animals (62-65), while GHSR antagonism *increases* food intake and body weight in rodents (66). Further supporting the notion that this hormone plays an endogenous role in food intake, peripheral ghrelin levels rise when fasting and prior to either scheduled or spontaneous meal ingestion and are reduced following nutrient consumption (62,67-74), with a greater suppression in response to carbohydrate or protein ingestion compared to fat (75-79). Genetic ghrelin deletion initially suggested that this peptide may have more critical effects on metabolic functions than on food intake and body adiposity, as the latter measures are normal in ghrelin knockout mice (80,81). However, mice lacking the GHSR display resistance to high-fat diet-induced obesity via mechanisms that include reduced food intake and reduced body adiposity, in addition to metabolic changes (82).

In 2005, another peptide was identified from the gene encoding ghrelin, dubbed obestatin or ghrelin-associated peptide (83). While ghrelin stimulates food intake and gastroduodenal motility, obestatin reduces food intake and inhibits gastrointestinal motility in fed, but not fasted, animals (83,84). Subsequent research on obestatin produced conflicting results of its effect on hypophagia, interaction

with ghrelin, gastrointestinal motility and its ability to activate GPR39, the putative obestatin receptor (85-89). Though there is some potential with respect to the role of obestatin (or obestatin/ghrelin balance) in pathological eating (e.g., anorexia nervosa), based on the current, conflicting state of the findings on this peptide, categorizing obestatin as an important peripheral signal for food intake and body weight regulation seems unjustified.

Adiposity Signals

Signals from the GI tract are acutely sensitive to nutrients entering the system and function primarily to regulate short-term intake on a meal-to-meal basis. However, the body also stores fuel for times of food shortage, mainly in the form of fat. One early hypothesis for the long-term regulation of body weight was that food intake and metabolic rate was adjusted based on the detection and regulation of the amount of adipose tissue present in the body (i.e., the “lipostatic hypothesis”; 90,91). To date, two major hormones have been identified and found to meet the criteria qualifying them as adiposity signals: insulin, which is produced in pancreatic β -cells, and leptin, which is secreted directly from adipocytes. These two peptides are secreted in proportion to the amount of body fat and have access to the brain where they act via central effector systems in the hypothalamus, as well as the hindbrain, to reduce food intake (92,93). Receptors for both insulin and leptin are found in the arcuate nucleus of the hypothalamus, a critical region for the control of energy homeostasis and central administration of both hormones potently reduces food intake (94-102). Hypothalamic administration of insulin antibodies has the opposite effect, increasing food intake and body weight (102,103) and rodents which are genetically incapable of leptin production (*ob/ob* mice) or have a dysfunctional leptin receptor (Zucker fatty or *fa/fa* rats and *db/db* mice) display an obese, hyperphagic phenotype (104-106), as do mice with a brain-specific deletion of the insulin receptor or disruption of pancreatic beta-cell and hypothalamic insulin receptor substrate 2 (107,108). Similarly, congenitally leptin-deficient humans are morbidly obese and markedly hyperphagic, as are those with leptin receptor mutations, although the symptoms are less severe (109,110).

In the hypothalamus, leptin receptors are located on both orexigenic neurons (i.e., neuropeptide Y (NPY)/agouti-related peptide (AgRP); 97) and some anorexigenic neurons (i.e., proopiomelanocortin (POMC)/cocaine- and amphetamine-related transcript (CART); 111). Insulin receptor expression is high in the arcuate nucleus with insulin receptor substrate-2, a key element mediating insulin effects on food intake, co-localizing there with NPY and α -melanocyte stimulating hormone (α -MSH) (112,113), as well as POMC, though insulin activates a different population of these cells than leptin, allowing for “cross-talk” between these signals (114). Broadly speaking, increases in the amount of stored fat increase circulating levels of insulin and leptin which, in turn, act via central regulatory mechanisms to reduce orexigenic signaling and increase anorexigenic signaling, allowing these peripheral adiposity signals to influence food intake.

Central Regulation

Hypothalamus

The primary forebrain regulation of food intake behavior is thought to occur in the hypothalamus. Early evidence indicated that lesions in this area had profound effects on ingestive behavior. Lesions of the ventromedial hypothalamus (VMH) result in drastically increased food intake and obesity, while lateral hypothalamic area (LHA) lesions yield hypophagia and reduced growth. These findings led to the hypothesis that these two areas controlled food intake by acting as the “satiety” and “feeding” centers, respectively, in the brain (115). Although this is now acknowledged to be a vast oversimplification of the regulation of food intake and body weight, the hypothalamus is still considered the key region for central control of energy homeostasis. A good deal more is now known regarding the molecular mechanisms at work in this area that act to control energy intake. The arcuate nucleus contains two populations of neurons that seem to be the first-order relay neurons in responding to adiposity signals from the periphery. The first arcuate neuronal population co-expresses the peptides NPY and the melanocortin

receptor antagonist AgRP, while the second population of neurons contains POMC, the pre-cursor to the melanocortin receptor agonist α -MSH, and CART. Central infusion of NPY or AgRP potently stimulates food intake (116-118), while icv administration of α -MSH or CART inhibits food intake (119,120), suggesting that these two neuronal populations represent a primary orexigenic and its opposing anorexigenic pathway, respectively, in the central regulation of energy homeostasis.

In support of the endogenous function of these peptides, food deprivation increases expression of AgRP and NPY mRNA, while decreasing POMC and CART gene expression (121-124). Overexpression of agouti or AgRP yields hyperphagia and obesity, as does disruption of the genes encoding the melanocortin-4 receptor, POMC or CART (125-129). Recently developed optogenetic techniques demonstrated that specific stimulation of AGRP neurons invoked cell number-dependent, light-frequency and duration dependent feeding response in well-fed mice, indicating that independent activation of AGRP cells could cause short-term feeding (130). Finally, while there is compensation for developmental deletion of the NPY or AgRP genes or neonatal destruction of NPY/AgRP neurons, ablation of these neurons in adult mice yields dramatic reductions in food intake and bodyweight, while the reverse occurs with ablation of POMC neurons (131-135). In humans, through relatively rare, genetic POMC deficiency also leads to an obese phenotype, as do a number of mutations of the MC-4 receptor. In addition, it has been suggested that variations in the POMC gene may be a contributing factor in obesity in the larger population (135).

As discussed above, leptin and insulin receptors are located on both orexigenic and anorexigenic cell types, suggesting that these neurons are responsive to circulating levels of adiposity signals and act as effectors for altering food intake in response to shifts in energy balance (at least those reflected in body adiposity). Leptin and insulin both cross the blood-brain barrier via independent, saturable transport mechanisms (136,137), indicating that peripheral production of these hormones can have central action. Indeed, as predicted, central insulin and leptin increase hypothalamic POMC expression, leptin increases activity in POMC neurons and melanocortin antagonists can block leptin-induced anorexia (138-141). These melanocortinergic neurons project from the arcuate to other areas of the hypothalamus, such as the paraventricular nucleus (PVN) and the LHA where several additional peptides that influence food intake and body weight are synthesized. The second-order neurons acting to regulate energy homeostasis in the PVN synthesize and release anorexigenic compounds, such as corticotrophin-releasing hormone (CRH), TRH, and oxytocin (142-147), while those in the LHA and adjacent perifornical area (PFA) are orexigenic, such as melanin-concentrating hormone (MCH) and orexin A and B (aka hypocretin 1 and 2) (148-151).

While leptin and insulin appear to control energy balance over a longer time scale, short-term peripheral signals, such as ghrelin and nesfatin also appear to act via the hypothalamic system to regulate intake. Receptors for ghrelin are also located on arcuate AgRP/NPY neurons, which are activated by central ghrelin administration to increase food intake (152-154). Administration of peripheral ghrelin activates neurons in the ARC as well, and AgRP and NPY have been demonstrated to be requisite mediators of the hyperphagia induced by systemic ghrelin (155,156). On the opposite side of the intake equation, nesfatin-1 appears to be a central satiety signal that acts in the hypothalamus independent of leptin function (likely via melanocortin pathways). Central injection of this peptide reduced food intake and body weight, while infusion of targeted antisense oligonucleotides produce the opposite effect, and fasting decreased nesfatin expression in the PVN (157). Following the identification of nesfatin-1 in the brain, it was found that this peptide is also secreted peripherally from endocrine cells in the stomach, where it is co-localized with ghrelin, and has been shown to cross the blood-brain barrier where it acts on neurons located in the PVN and arcuate nucleus of the hypothalamus (158-161). Similar to central administration, peripherally delivered nesfatin-1 reduces food intake and this anorexigenic effect was abolished in mice pre-treated with capsaicin suggesting a vagally-mediated pathway between peripheral nesfatin-1 and central sites of action (162). Not surprisingly, like other vagally mediated GI satiety signals (e.g., CCK), nesfatin-1 influences food intake by reducing the size of individual meals and increasing the length of inter-meal intervals (163).

Hindbrain

In contrast to ghrelin and nesfatin-1, which are secreted from the stomach, most short-term signals arising from lower in the GI tract tend to be received and integrated in the hindbrain. Receptors for mechanical and chemical signals in the gut are found on afferent terminals of the vagus nerve, which transduces these sensory signals and relays information to the nucleus of the solitary tract (NTS) (164). Studies using a chronic decerebrate rat model have demonstrated that these signals from the periphery (e.g., gastric preloads, CCK) can act to reduce meal size in the absence of hypothalamic input (165,166), however, there are a number of reciprocal connections between hypothalamic and hindbrain nuclei which suggest an integration of information from both sites acts to control food intake and energy balance (167-169). In addition, there are neurons expressing receptors for leptin, melanocortins and NPY, as well as POMC/ α -MSH found in brainstem nuclei (170-175). Administration of synthetic MC receptor agonists and antagonists, AgRP, or NPY or its receptor agonists to the fourth ventricle all yield similar effects on food intake as when they are delivered to hypothalamic sites via the third ventricle (176-179). Leptin appears to exert a modulatory influence on brainstem controls of feeding, as well, as its administration alters the responsiveness of NTS neurons to gastric distension and CCK, as well as mediating the effects of these and other peripheral factors on food intake (180-184). Selective leptin-R ablation in GLP-1 expressing neurons in the hindbrain produced an obese phenotype characterized by hyperphagia and increased metabolic rate in mice (185). Although the evidence clearly demonstrates a role for both hypothalamic and hindbrain sites in the regulation of food intake, it is still unclear which aspects are controlled by each of these areas and how these regions interact to ultimately regulate energy balance.

“Gut” Peptides in the CNS

In addition to acting through the vagus, a number of the peptides identified primarily as peripheral signals also function directly at CNS sites to affect energy homeostasis. Receptors for these peptides are often found in hindbrain areas, such as the NTS and DMH as well as in the circumventricular organs, allowing for central monitoring of the status of these circulating hormones (186). Whereas CCK-A receptors predominate in the GI tract, CCK-B receptors are found largely in the CNS. It appears that CCK-A receptors are the primary mediators of food intake and satiety both peripherally and centrally, however under some conditions, central CCK-B receptors may be involved (187,188). Central CCK receptors, however, do not appear to be necessary for the intake suppressive effects of peripheral CCK and observed “central” feeding effects may be due to stimulation of peripheral receptors (189-191), indicating an independent role for centrally-produced CCK in food intake. CCK is produced and released in a number of brain regions, including the hypothalamus, but also the caudate nucleus, hippocampus, striatum, and cortical areas, and in addition to ingestive behavior, central CCK is implicated in anxiety, pain, and cognition, functions which seem predominantly related to CCK-B receptor activity (192-197).

Beyond CCK, peripheral PYY 3-36 may cross the blood-brain barrier to act at Y_2 receptors in the ARC, inhibiting NPY activity and stimulating POMC neurons (29,198). Direct central administration of PYY, however, produces a strong hyperphagic effect, likely acting via Y-family receptors, which also mediate NPY actions, in other brain regions, such as the PVN and hindbrain (178,199,200). GLP-1 and its receptors are also located in the hypothalamus as well as the brainstem, and icv infusion of GLP-1 reduces food intake in fasted rats, likely acting in the PVN, while the receptor antagonist exendin₉₋₃₉ increased feeding behavior (201,202).

Hippocampus

Just as conventional thinking about the localization of brain function firmly seats control of ingestive behavior in the hypothalamus, the hippocampus is typically considered the region responsible for memory. This dates back to the observation that Patient HM was unable to create new long-term memories after surgery that lesioned the hippocampus and associated structures (203). In addition to broad-scope anterograde amnesia, HM repeatedly reported the same level of hunger/fullness when asked by researchers, and would continue to eat as long as food was placed in front of him (204).

Studies of hippocampal lesions in animal models have yielded somewhat contradictory outcomes in terms of food intake and body weight, with the most common result being a pattern of increased number of meals/food contacts without an increase in either food intake or body weight (205,206), though other studies found increases in food intake that did not result in body weight changes and increases in both food intake and body weight relative to controls (207-209). The most recent study, employing a more specific lesioning technique and controlling for the effects of surgical recovery, found that hippocampal lesioned rats showed increased body weight, which could be attributed both to increases in food intake and to alterations in metabolic function (210).

Furthermore, like HM, rats with selective hippocampal lesions appear unable to detect or interpret signals of hunger and satiety, as they are unable to learn a discrimination based on this internal state or to perform this discrimination when it is learned prior to the lesion, indicating it is an impairment in using hunger states, rather than a learning deficit (206,211). In humans, activity in the hippocampus during periods of food restriction and negative energy balance is supportive of this region playing a role in response to these states (212).

With respect to physiology, the hippocampus has bidirectional connections with traditional “feeding centers” including the hypothalamus and the brainstem, which may serve to communicate signals of energy balance to this region (213,214). Administration of either leptin or ghrelin alters food intake when injected directly into the ventral hippocampus, with leptin reducing and ghrelin increasing consummatory and appetitive behaviors (215,216) and inactivation of the dorsal hippocampal after a meal inhibits the onset of the next meal, suggesting a role for this region in meal initiation (217).

Estrogens

There are a number of other peptides that affect food intake outside of those listed above. Estrogen has inhibitory effects on food intake that are observed both as cyclic changes in caloric consumption with hormonal fluctuations in females and as a dramatic increase in food intake following ovariectomy (218). Further, mice lacking estrogen receptor alpha (ER α) have increased body adiposity (219). Recent experiments show that centrally administered estradiol acts in the medial preoptic area, arcuate nucleus and dorsal raphe to decrease food intake (220). Experiments utilizing peripherally administered estrogens suggest a modulatory effect on the function of a number of other food intake-related peptides, including leptin, insulin, ghrelin, CCK, and the melanocortins (218,221). Sensitivity to central leptin and insulin differ between female and male rats, with females being more sensitive than males to leptin, while the reverse is true for insulin, and estrogen is responsible for this sensitivity (222,223). ER β is found on subcutaneous adipose tissue and likely plays a role in the observation that subcutaneous fat deposition is increased by the administration of estradiol (223,224).

Other Neurotransmitter Systems

Serotonin

Monoamine systems, including dopamine (DA) and serotonin (5-HT) have also been shown to be involved in food intake. Activation of serotonin receptors using subtype-specific agonists or reuptake inhibitors (SSRIs) has been clearly demonstrated to reduce food intake and has been strongly targeted as an effective weight-loss treatment (225). It is likely that serotonin has its effects on body weight regulation through interaction with both hypothalamic and extra-hypothalamic systems, as receptors for this transmitter are widespread and co-localized with a number of key feeding peptides (e.g., melanocortins, orexins, ghrelin and leptin) (226). Dexfenfluramine, a broad serotonergic agonist, was one component of the weight loss treatment Fen-Phen, which was highly effective at producing weight loss in animal subjects and overweight/obese human patients (227,228). However, this treatment was removed from the market due to dangerous cardiovascular side effects (229). The weight loss success of these drugs, however, led to the search for more targeted serotonergic compounds for this purpose. The

selective 5-HT_{2C} receptor agonist lorcaserin has been shown to produce effective weight loss in short- and long-term clinical trials and in 2012 was approved by the FDA to treat obesity (230,231), in spite of a still very general understanding of the mechanism by which serotonin influences food intake and body weight.

Dopamine

Animals treated with DA receptor agonists also exhibit reductions in food intake with activation of D₁, D₂ and D₃ receptor subtypes seeming to alter different aspects of appetitive and consummatory behaviors with DA₁ and DA₃ receptors appearing to play the most specific roles in ingestion (232-234). The mesolimbic DA system in particular is posited to be robustly involved in food anticipation and learned appetitive behaviors, particularly those related to highly palatable foods, as indicated by increases in dopamine release in these regions in response to food and food stimuli (235-237). Reductions in short-term tests of sucrose licking and sham feeding in rats treated with dopamine antagonists supports the notion that the effects of dopamine are primarily on taste hedonics, rather than post-ingestive effects (238,239). In particular, the mesolimbic DA system is posited to be robustly involved in food anticipation and learned appetitive behaviors, particularly those related to highly palatable foods, as indicated by increases in dopamine activity in these regions in response to food and food stimuli in both humans and non-human animals (235,236).

Acetylcholine

Although not given as much attention as other neurotransmitter systems, recent studies also indicate a role for cholinergic systems in food intake, as well, particularly within the nucleus accumbens-ventral tegmental pathway. Specifically, injections of muscarinic acetylcholine (mACh) receptor antagonists into the accumbens reduces consummatory, but not appetitive behaviors, in rats, and this effect appears to interact with the opioid, but not dopaminergic systems (240-242). Antagonism of nACh receptors in the VTA reduces ghrelin-induced food intake, likely through a reduction in the reward value of the food, as indicated by impaired development of a food-conditioned place preference (243). Outside of mesolimbic regions, nicotine, an nACh receptor agonist, has been shown to have a suppressive effect on food intake via binding to acetylcholine receptors on POMC neurons, activating anorexigenic MC4 pathways (244).

Opioids

The opioid system also has a potent effect on food intake, although, like dopaminergic effects, this system seems to reflect the hedonic impact of food with greater responsiveness to the palatability of the food than the energy status of the organism (245,246). Opioid agonists typically stimulate intake of preferred foods, while antagonists have the opposite effect (246). More recent evidence finds that opioid effects on feeding are particularly potent when acting in conjunction with endocannabinoids (247,248).

Endocannabinoids

Activation of cannabinoid (specifically, CB₁) receptors by either exogenous or endogenous ligands (e.g., Δ^9 -THC, anandamide) stimulates food intake, while pharmacological CB₁ antagonists (e.g., rimonabant) reduce food intake generally in fasted animals. Endocannabinoid levels are elevated in the hypothalamus during food deprivation and are reduced by food consumption and by leptin, indicating they are likely also involved in homeostatic control of food intake (249,250). Rimonabant has also been demonstrated to be effective at producing weight loss in humans (251). The presence of endocannabinoids in limbic regions and their interaction with opioids to modulate food intake suggests that this system also functions to affect intake of palatable foods (252).

Behavioral Effects of Molecular Genetics

There are a number of ways by which the numerous peptides and neural systems described here may act to alter food intake. They may alter meal initiation (i.e., the likelihood of beginning an eating bout), which is generally observed as a change in meal frequency, or they may alter meal termination (i.e., how much is consumed prior to ending a meal), which is generally observed as a change in meal size. They may also affect the subjective feelings that an individual interprets as “hunger” or “fullness” and uses to determine when to begin or end a meal or the subjective palatability or reward value associated with eating particular foods.

Meal Initiation, Meal Termination and Food Selection

Meal pattern analysis indicates that size of individual meals is the mechanism by which total food intake is generally altered. Increased meal size is the primary response to fasting and is almost exclusively responsible for the elevated caloric intake in rats bred for their susceptibility to diet-induced obesity (253-255). Furthermore, although some reports show overweight and obese humans consume both larger and more frequent meals, there is evidence that it is meal size that differs most significantly between those gaining weight and those maintaining their current weights (256,257). When analyzing the component of food intake that is influenced by peptides found either peripherally or centrally, it is meal size that is most often found to be affected, leading some to suggest that meal termination is more strongly controlled by biological processes, while there a vast number of environmental influences that are more likely to be involved in meal initiation (i.e., availability of food, time of day, cognitive factors, learned associations/signals) (258). The majority of the peripheral satiety hormones, including CCK, gastrin releasing peptide/bombesin, GLP-1 agonist exendin-4, amylin and leptin, appear to act by reducing meal size with little or no effect on meal frequency (27,28,50,259-263). However, one study found that an obesogenic phenotype characterized by high meal frequency was significantly correlated with polymorphism and haplotype variants in leptin and leptin-R genes (264). Not surprisingly, as central leptin effectors, melanocortin agonists have been shown to reduce meal size, while MC antagonists have the opposite effect (265,266). Similarly, icv administration of CART decreased and NPY increased meal size (267,268). In contrast to these signaling molecules, ghrelin acts to increase intake by playing a role in meal initiation and food anticipation (67,71,269,270), which is consistent with its role as a “hunger” signal (see below). Recent data indicates that there may be a meal anticipatory effect for GLP-1, as well (271).

In addition to influencing total energy intake via changes in these basic meal parameters when a constant test diet is used, some systems also differentially affect food selection or intake based on the macronutrient compositions of the diet. While NPY and AgRP both increase total caloric intake, NPY appears to induce greater appetitive and consummatory behaviors for foods high in carbohydrates, while the melanocortin system selectively affects fat intake and responding for fat-associated stimuli (199,272-275). Not surprisingly, leptin, acting to inhibit both NPY and AgRP, reduces intake of both carbohydrates and fats (276).

Interoceptive States: “Hunger” and “Satiety”

A number of these peripheral signals seem to be responsible for the subjective feelings of “hunger” or “satiety”. These states can be assessed in a rodent model using a experimental design known as the “deprivation intensity discrimination paradigm”, in which rats are trained to discriminate between internal cues associated with 24 hours or 1 hour of food deprivation by receiving a reinforcer in a specific environment under only one of these conditions (277,278). The generalization between these deprivation states and those of a variety of potential hunger- or satiety-inducing peptides is tested by administering an exogenous dose of the peptide of interest and measuring the animal’s behavior in the training environment. These types of experiments have suggested that ghrelin produces interoceptive cues similar to that of 24-hr food deprivation (i.e., “hunger”), while CCK and leptin produce cues similar to 1-hr food deprivation (i.e., “fullness”) (279-281). Other peptides that influence food intake, such as NPY,

bombesin, and MC-R agonists and antagonists do not appear to produce cues that generalize to either deprivation state, suggesting that their mechanism of action is independent of inducing a subjective feeling of “hunger” or “satiety” (280, 282-284). In humans, rating scales are often used to measure the subjective sensations perceived by subjects following administration of pharmacological agents associated with food intake regulation. These ratings are frequently, but not always, correlated with or predictive of consummatory behaviors, suggesting that, as in animals, these reported sensory stimuli may represent only one mechanism of altering food intake (285,286). Mechanical distention of the stomach and systemic infusion of CCK seem to produce consistent increases in self-reported ratings of “fullness” and decreases in ratings of “hunger”, while the effects of infusions of GLP-1 and PYY on these sensations are not as clear (33,287-291). Conversely, increased “hunger” ratings have been observed following treatment with peripheral ghrelin and CCK-A receptor antagonists (63,292). Ghrelin levels are also correlated with reported hunger levels and meal initiation in humans in the absence of external cues associated with meals, including time and food-related stimuli (71).

Reward Value, “Hedonic” Eating and Food Motivation

While the peripheral and hypothalamic systems have largely been viewed as involved in the homeostatic aspects of food intake based on energy balance, other systems, primarily the dopaminergic, opioidergic and, more recently, cannabinoid systems, seem to influence intake based on palatability, or subjective “reward value” of the food being consumed (245,246,252,293). Extensive work has characterized the roles of dopamine and opioids in reward-related eating as associated with “wanting” and “liking” palatable foods, respectively (245,294,295). However, the involvement of cannabinoids in metabolic processes, the interconnection of hypothalamic circuits with those in mesolimbic and striatal regions involved in reward, the effect of “feeding peptides” (such as leptin, ghrelin, and GLP-1) in the nucleus accumbens to alter appetitive and consummatory behaviors, and evidence of interaction between these systems is further blurring the lines (249,296-301,212).

Food motivation is often measured by the use of operant conditioning procedures – most commonly, a progressive ratio (PR) test in which the number of responses required increases for each reinforcer earned. The maximum number of responses that an animal is willing to engage in to earn a reinforcer is termed the “break point” and considered a measure of motivation. A number of studies have tested the administration of feeding peptides and hormones in this paradigm. As would be predicted, increases in PR break point have been observed in response to ghrelin, orexin-A, NPY, a CB1 receptor agonist and deletion of melanocortin-4 receptors, while decreases were observed after administration of leptin, insulin, a CB1 receptor antagonist, a GLP-1 agonist or deletion of CB1 receptors (272,302-306). While reductions in appetitively motivated behavior may stem from a number of underlying, modulatory mechanisms (e.g., shifts in hunger state, recall of the food value, etc.), this measure is a reliable indicator of the effort that will be expended to obtain food, which is both necessary for consumption and appears to be correlated with future weight gain in humans (307).

Cognitive Function

Learning and memory processes are critical to the regulation of food intake behaviors. Flavor-flavor and flavor-nutrient learning contribute significantly to learned food preferences and engaging explicit memories of eating episodes or manipulating attention to food during an eating episode can alter food intake (308,309). The hippocampus appears to be critically involved in the ability to use hunger and satiety as discriminative cues to control behavior, as described above (206,211) – and the ability to use those cues to determine eating, or not, in the presence of food cues has been posited as a model that describes the way in which we regulate energy balance (310). Several of the molecules discussed here as “feeding” peptides have been shown to play roles in various cognitive functions, both food-related and non-food-related (311,312). In particular, there is substantial evidence for ghrelin and leptin acting in the hippocampus to alter physiological function associated with memory, such as altering long-term potentiation and other measures of synaptic plasticity (313,314), as well as to influence a number of

food-motivated and food-seeking behaviors (215,216), which are necessary precursors to intake in animals and humans in environments with readily available food..

Regulatory Disturbances Associated with Obesity

Prolonged high-fat diet consumption, leading to obesity, in both humans and non-human animal models alters the endogenous profiles and the efficacy of a number of these energy balance-related signaling molecules. As would be expected, leptin and insulin levels are elevated in overweight subjects, as these adiposity signals circulate in levels proportional to the amount of body fat (315-317). Obese humans and rodents consuming diets high in fat also display peripheral reductions in circulating ghrelin and fasting PYY levels (318-321), although diet-induced obesity in rodents does not alter the effectiveness of PYY (322). Rodents who have been maintained on a diet high in saturated fat exhibit reduced expression of NPY and AgRP mRNA in the hypothalamus (323,324), although levels of these peptides measured in the cerebrospinal fluid of humans did not differ significantly between lean and obese subjects (325). These increases in the anorexigenic signals leptin and insulin and decreases in orexigens, such as ghrelin, AgRP and NPY would be predicted based on the state of positive energy balance in obese individuals and would be expected to reduce food intake and body weight. However, these individuals tend to remain at elevated weights and, frequently continue to gain weight, suggesting that these systems become dysfunctional in obesity. In fact, there is a large body of evidence supporting just that notion. Studies have clearly demonstrated that diet-induced obese humans and animals become resistant to the anorectic effects of both peripheral and central leptin, as well as central insulin (316,326-331). A number of experiments have demonstrated that obesity and consumption of high-fat diet impair the transport of leptin across the blood-brain barrier, reduce the sensitivity of intracellular signaling pathways activated by insulin and leptin, reduce the capacity of these hormones to act through central effector systems such as NPY and melanocortin pathways and decrease hypothalamic NPY and CCK levels (332-339). Diet-induced obese rodents also appear to be less sensitive to the food intake-reducing effects of centrally administered melanocortin agonists (316).

Although peripheral satiety signals tend to maintain normal basal levels in obese subjects, several of these peptides, including CCK, PYY and Apo A-IV are less responsive to nutrient influx into the gut, with decreased release following meals and reduced satiety effects. Postprandial ghrelin levels tend to remain high, which may contribute further to reduced feelings of satiety (340-343). However, there appears to be reduced responsiveness (i.e., "resistance") to administration of both ghrelin and GLP-1 in obese animals (319,344,345). Chronic high-fat diet consumption in rats leads to reduced operant responding for sucrose, as well as impaired conditioned place preference and altered mesolimbic dopamine function (346). Correspondingly, neuroimaging assays have indicated a reduction in central dopamine D2 receptors in obese humans, as well as increases in neural activity in reward areas during food anticipation, but decreased activity during consumption, leading to speculation that this may indicate decreased reward sensitivity and result in compensatory overeating (347-349).

In addition to changes in sensitivity in regulatory and reward systems, changes in memory structures, primarily the hippocampal formation, have also been observed following diet-induced obesity. These changes include reductions in BDNF levels, reductions in the capacity for LTP, decreases in neurogenesis, and loss of blood-brain barrier integrity (350-353), along with behavioral deficits that mimic the impairments observed following hippocampal damage, such as poor performance in spatial memory tasks and inhibitory learning (350,353). It has been proposed that these impairments are part of a "vicious cycle" that contributes to the maintenance of obesity by further impairing the ability to use hunger/satiety signals to regulate eating based on prior associative learning (310).

However, a major difficulty with these types of studies, in which comparisons are made between obese and lean populations, is the inability to dissociate whether these alterations cause obesity or are a consequence of chronic overeating and increased body adiposity (e.g., 356). Prospective studies in humans have been minimal and interpretation complex, with attempts to predict weight gain or future BMI based on neural activity in reward regions in response to palatable food cues yielding inconsistent

results (357,358). A particularly useful tool in addressing this issue has been the development of selectively bred rats either prone or resistant to obesity induced by the consumption of high-fat, calorically dense diets (359). Studies of these animals on standard low-energy-density diets prior to divergence in weight gain and body adiposity have demonstrated that animals prone to diet-induced obesity also have a pre-disposition to insulin and leptin resistance, reduced central leptin signaling, reduced central insulin and leptin receptor binding, and increased expression of hypothalamic NPY, indicating that these factors may play a role in the development of obesity (328,331,360-369). On the other hand, no differences were observed in melanocortin binding and ghrelin and GHSR expression were reduced in obesity-prone relative to obesity-resistant rats, suggesting that these systems are not implicated in the onset of hyperphagia and weight gain (360,362).

Conclusion

The control of food intake involves the detection and integration of many different stimuli beginning with gastrointestinal detection of food volume and nutrient content acting to limit consumption within meals and peripheral adiposity signals acting via central sites and interacting with other peptides and hormones to balance short-term intake with long-term energy stores. In addition, there are a number of systems which act to select particular foods based on their nutrient content or, outside of homeostatic regulation, based on their palatability or reward value. These signals can alter a range of behaviors, including appetitive behavior, consumption, food selection, the interoceptive states labeled “hunger” and “satiety”, and even memory and other cognitive functions, to ultimately control energy intake. In spite of this complex regulatory system, a large proportion of the population of the United States is currently overweight or obese, suggesting a dysfunction. In fact, obesity and the consumption of high-fat diets appear to induce resistance to a number of the signals designed to limit intake in the face of positive energy balance, complicating the search for effective treatments. However, furthering a careful understanding of ingestive behaviors and the biological mechanisms that underlie them is the best hope for identifying a way to reverse the current obesity epidemic.

References

1. Ogden CL, Carroll MD, Curtin LR, McDowell MA, Tabak CJ, Flegal KM. Prevalence of overweight and obesity in the United States, 1999-2004. *JAMA* 295:1549-1555, 2006.
2. Finkelstein EA, Ruhm CJ, Kosa KM. Economic causes and consequences of obesity. *Annu Rev Public Health* 26:239-257, 2005.
3. Young RC, Gibbs J, Antin J, Holt J, Smith GP. Absence of satiety during sham feeding in the rat. *J Comp Physiol Psychol* 87:795-800, 1974.
4. Liebling DS, Eisner JD, Gibbs J, Smith GP. Intestinal satiety in rats. *J Comp Physiol Psychol* 89:955-965, 1975.
5. Prechtl JC, Powley TL. The fiber composition of the abdominal vagus of the rat. *Anat Embryol* 181:101-115, 1990.
6. Berthoud HR, Powley TL. Vagal afferent innervation of the rat fundic stomach: morphological characterization of the gastric tension receptor. *J Comp Neurol* 319:261-276, 1992.
7. Eisen S, Davis JD, Rauhofer E, Smith GP. Gastric negative feedback produced by volume and nutrient during a meal in rats. *Am J Physiol Regul Comp Physiol* 281:R1201-R1214, 2001.
8. Phillips RJ, Powley TL. Gastric volume rather than nutrient content inhibits food intake. *Am J Physiol* 271:R766-R769, 1996.
9. Phillips RJ, Powley TL. Gastric volume detection after selective vagotomies in rats. *Am J Physiol* 274:R1626-R1638, 1998.
10. Seeley RJ, Kaplan JM, Grill HJ. Effect of occluding the pylorus on intraoral intake: a test of the gastric hypothesis of meal termination. *Physiol Behav* 58:245-249, 1995.
11. Rauhofer EA, Smith GP, Gibbs J. Acute blockade of gastric emptying and meal size in rats. *Physiol Behav* 54:881-884, 1993.
12. Merali Z, McIntosh J, Anisman H. Role of bombesin-related peptides in the control of food intake. *Neuropeptides* 33:376-386, 1999.
13. Washington MC, Wright SA, Sayegh SI. Gastrin releasing peptide-29 evokes feeding responses in the rat. *Peptides* 32:241-245, 2011.
14. Liddle RA. Cholecystokinin cells. *Annu Rev Physiol* 59:221-242, 1997.
15. Orskov C, Rabenhøj L, Wettergren A, Kofod H, Holst JJ. Tissue and plasma concentrations of amidated and glycine-extended glucagon-like peptide I in humans. *Diabetes* 43:535-539, 1994.
16. Adrian TE, Ferri GL, Bacarese-Hamilton AJ, Fuessl HS, Polak JM, Bloom SR. Human distribution and release of a putative new gut hormone, peptide YY. *Gastroenterology* 89:1070-1077, 1985.
17. Liu M, Doi T, Tso P. Regulation of intestinal and hypothalamic apolipoprotein A-IV. *Exp Biol Med* 228:1181-1189, 2003.
18. Foltz M, Ansems P, Schwarz J, Tasker MC, Loubakos A, Gerhardt CC. Protein hydrolysates induce CCK release from enteroendocrine cells and act as partial agonists of the CCK1 receptor. *J Agric Food Chem* 56:837-843, 2008.
19. Geraedts MC, Troost FJ, Fischer MA, Edens L, Saris WH. Direct induction of CCK and GLP-1 release from murine endocrine cells by intact dietary proteins. *Mol Nutr Food Res* 55:476-484, 2011.
20. Liddle RA, Green GM, Conrad CK, Williams JA. Proteins but not amino acids, carbohydrates, or fats stimulate cholecystokinin release in the rat. *Am J Physiol* 251:G243-G248, 1986.
21. Buchan AMJ. Nutrient tasting and signaling mechanisms in the gut III. Endocrine cell recognition of luminal nutrients. *Am J Physiol* 277:G1103-G1107, 1999.
22. Orr J, Davy B. Dietary influences on peripheral hormones regulating energy intake: potential applications for weight management. *J Am Diet Assoc* 105

23. Essah PA, Levy JR, Sistrun SN, Kelly SM, Nestler JE. Effect of macronutrient composition on postprandial peptide YY levels. *J Clin Endocrinol Metab* 92:4052-4055, 2007.
24. Sloth B, Holst JJ, Flint A, Gregersen NT, Astrup A. Effects of PYY1-36 and PYY3-36 on appetite, energy intake, energy expenditure, glucose and fat metabolism in obese and lean subjects. *Am J Physiol Endocrinol Metab* 292:E1062-E1068, 2007.
25. Rolls BJ. Carbohydrates, fats, and satiety. *Am J Clin Nutr* 61:960S-967S, 1995.
26. Gerstein DE, Woodward-Lopez G, Evans AE, Kelsey K, Drewnowski A. Clarifying concepts about macronutrients' effects on satiation and satiety. *J Am Diet Assoc* 104:1151-1153, 2004.
27. Gibbs J, Young RC, Smith GP. Cholecystokinin decreases food intake in rats. *J Comp Physiol Psychol* 84:488-495, 1973.
28. Kissileff HR, Pi-Sunyer FX, Thornton J, Smith GP. C-terminal octapeptide of cholecystokinin decreases food intake in man. *Am J Clin Nutr* 34:154-160, 1981.
29. Batterham RL, Cowley MA, Small CJ, Herzog H, Cohen MA, Dakin CL, Wren AM, Brynes AE, Low MJ, Ghatei MA, Cone RD, Bloom SR. Gut hormone PYY(3-36) physiologically inhibits food intake. *Nature* 418:650-654, 2002.
30. Fujimoto K, Machidori H, Iwakiri R, Yamamoto K, Fujisaki J, Sakata T, Tso P. Effect of intravenous administration of apolipoprotein A-IV on patterns of feeding, drinking and ambulatory activity of rats. *Brain Res* 608:233-237, 1993.
31. Chelikani PK, Haver AC, Reidelberger RD. Intravenous infusion of peptide YY(3-36) potently inhibits food intake in rats. *Endocrinology* 146:879-888, 2005.
32. Ritter RC. Gastrointestinal mechanisms of satiation for food. *Physiol Behav* 81:249-273, 2004.
33. Degen L, Oesch S, Casanova M, Graf S, Ketterer S, Drewe J, Beglinger C. Effect of peptide YY3-36 on food intake in humans. *Gastroenterology* 129:1430-1436, 2006.
34. Chaudhri O, Small C, Bloom S. Gastrointestinal hormones regulating appetite. *Philos Trans R Soc Lond B Biol Sci* 361:1187-1209, 2006.
35. Sloth B, Davidsen L, Holst JJ, Flint A, Astrup A. Effect of subcutaneous injections of PYY1-36 and PYY3-36 on appetite, ad libitum energy intake, and plasma free fatty acid concentration in obese males. *Am J Physiol Endocrinol Metab* 293:E604-E609, 2007.
36. Tschöp M, Castaneda TR, Joost HG, Thone-Reineke C, Ortmann S, Klaus S, Hagan MM, Chandler PC, Oswald KD, Benoit SC, Seeley RJ, Kinzig KP, Moran TH, Beck-Sickinger AG, Koglin N, Rodgers RJ, Blundell JE, Ishii Y, Beattie AH, Holch P, Allison DB, Raun K, Madsen K, Wulff BS, Stidsen CE, Birringer M, Kreuzer OJ, Schindler M, Arndt K, Rudolf K, Mark M, Deng XY, Whitcomb DC, Halem H, Taylor J, Dong J, Datta R, Culler M, Craney S, Flora D, Smiley D, Heiman ML. Physiology: does gut hormone PYY3-36 decrease food intake in rodents? *Nature* 430:165-166, 2004.
37. Moran TH. Gut peptide signaling in the controls of food intake. *Obesity* 14:250S-253S, 2006.
38. Brenner L, Ritter RC. Peptide cholecystokinin receptor antagonist increases food intake in rats. *Appetite* 24:1-9, 1995.
39. Corwin RL, Gibbs J, Smith GP. Increased food intake after type A but not type B cholecystokinin receptor blockade. *Physiol Behav* 50:255-258, 1991.
40. Williams DL, Baskin DG, Schwartz MW. Evidence that intestinal glucagon-like peptide-1 plays a physiological role in satiety. *Endocrinology* 150:1680-1687, 2009.
41. Gibbs J, Smith GP. Gut peptides and food in the gut produce similar satiety effects. *Peptides* 3:553-557, 1982.
42. Antin J, Gibbs J, Holt J, Young RC, Smith GP. Cholecystokinin elicits the complete behavioral sequence of satiety in rats. *J Comp Physiol Psychol* 89:784-790, 1975.
43. Martin CF, Gibbs J. Bombesin elicits satiety in sham feeding rats. *Peptides* 1:131-134, 1980.
44. Moran TH, Bi S. Hyperphagia and obesity in OLETF rats lacking CCK-1 receptors. *Philos Trans R Soc Lond B Biol Sci* 361:1211-1218, 2006.
45. Batterham RL, Heffron H, Kapoor S, Chivers JE, Chandarana K, Herzog H, LeRoux CW, Thomas EL, Bell JD, Withers DJ. Critical role for peptide YY in protein-mediated satiation and body-weight regulation. *Cell Metab* 4:223-233, 2006.

46. Blevins JE, Overduin J, Fuller JM, Cummings DE, Matsumoto K, Moralejo DH. Normal feeding and body weight in Fischer 344 rats lacking the cholecystokinin-1 receptor gene. *Am J Physiol Regul Integr Comp Physiol* 303:R1231-R1240, 2009.
47. Scrocchi LA, Drucker DJ. Effects of aging and a high fat diet on body weight and glucose tolerance in glucagon-like peptide-1 receptor -/- mice. *Endocrinology* 139:3127-3132, 1998.
48. Lukinius A, Wilander E, Westermark GT, Engstrom U, Westermark P. Co-localization of islet amyloid polypeptide and insulin in the B cell secretory granules of the human pancreatic islets. *Diabetologia* 32:240-244, 1989.
49. Butler PC, Chou J, Carter WB, Wang YN, Bu BH, Chang D, Chang JK, Rizza RA. Effects of meal ingestion on plasma amylin concentration in NIDDM and nondiabetic humans. *Diabetes* 39:752-756, 1990.
50. Lutz TA, Geary N, Szabady MM, Del Prete E, Scharrer E. Amylin decreases meal size in rats. *Physiol Behav* 58:1197-1202, 1995.
51. Morley JE, Flood JF. Amylin decreases food intake in mice. *Peptides* 12:865-869, 1991.
52. Lutz TA, Del Prete E, Scharrer E. Reduction of food intake in rats by intraperitoneal injection of low doses of amylin. *Physiol Behav* 55:891-895, 1994.
53. Reidelberger RD, Haver AC, Arnelo U, Smith DD, Schaffert CS, Permert J. Amylin receptor blockade stimulates food intake in rats. *Am J Physiol Regul Integr Comp Physiol* 287:R568-R574, 2004.
54. de Jong A, Strubbe JH, Steffens AB. Hypothalamic influence on insulin and glucagon release in the rat. *Am J Physiol* 233:E380-E388, 1977.
55. Langhans W, Pantel K, Muller-Schell W, Eggenberger E, Scharrer E. Hepatic handling of pancreatic glucagon and glucose during meals in rats. *Am J Physiol* 247:R827-R832, 1984.
56. LeSauter J, Geary N. Hepatic portal glucagon infusion decreases spontaneous meal size in rats. *Am J Physiol* 261:R154-R161, 1991.
57. LeSauter J, Noh U, Geary N. Hepatic portal infusion of glucagon antibodies increases spontaneous meal size in rats. *Am J Physiol* 261:R162-R165, 1991.
58. Geary N, Kissileff HR, Pi-Sunyer FX, Hinton V. Individual, but not simultaneous, glucagon and cholecystokinin infusions inhibit feeding in men. *Am J Physiol* 262:R975-R980, 1992.
59. Langhans W, Zeiger U, Scharrer E, Geary N. Stimulation of feeding in rats by intraperitoneal injection of antibodies to glucagon. *Science* 218:894-896, 1982.
60. Kojima M, Hosoda H, Date Y, Nakazato M, Matsuo H, Kangawa K. Ghrelin is a growth-hormone-releasing acylated peptide from stomach. *Nature* 402:656-660, 1999.
61. Cowley MA, Smith RG, Diano S, Tschop M, Pronchuk N, Grove KL, Strasburger CJ, Bidlingmaier M, Esterman M, Heiman ML, Garcia-Segura LM, Nilini EA, Mendez P, Low MJ, Sotonyi P, Friedman JM, Liu H, Pinto S, Colmers WF, Cone RD, Horvath TL. The distribution and mechanisms of action of ghrelin in the CNS demonstrates a novel hypothalamic circuit regulating energy homeostasis. *Neuron* 37:649-661, 2003.
62. Tschop M, Smiley DL, Heiman ML. Ghrelin induces adiposity in rodents. *Nature* 407:908-913, 2000.
63. Wren AM, Seal LJ, Cohen MA, Brynes AE, Frost GS, Murphy KG, Dhillo WS, Ghatei MA, Bloom SR. Ghrelin enhances appetite and increases food intake in humans. *J Clin Endocrinol Metab* 86:5992, 2001.
64. Wren AM, Small CJ, Abbott CR, Dhillo WS, Seal LJ, Cohen MA, Batterham RL, Taheri S, Stanley SA, Ghatei MA, Bloom SR. Ghrelin causes hyperphagia and obesity in rats. *Diabetes* 50:2540-2547, 2001.
65. Druce MR, Wren AM, Park AJ, Milton JE, Patterson M, Frost G, Ghatei MA, Small C, Bloom SR. Ghrelin increases food intake in obese as well as lean subjects. *Int J Obes* 29:1130-1136, 2005.
66. Asakawa A, Inui A, Kaga T, Katsuura G, Fujimiya M, Fujino MA, Kasuga M. Antagonism of ghrelin receptor reduces food intake and body weight gain in mice. *Gut* 52:947-952, 2003.
67. Cummings DE, Purnell JQ, Frayo RS, Schmidova K, Wisse BE, Weigle DS. A preprandial rise in plasma ghrelin levels suggests a role in meal initiation in humans. *Diabetes* 50:1714-1719, 2001.

68. Tschop M, Wawarta R, Riepl RL, Friedrich S, Bidlingmaier M, Landgraf R, Folwaczny C. Post-prandial decrease of circulating human ghrelin levels. *J Endocrinol Invest* 24:RC19-RC21, 2001.
69. Crowley WR, Ramoz G, Keefe KA, Torto R, Kalra SP, Hanson GR. Differential effects of methamphetamine on expression of neuropeptide Y mRNA in hypothalamus and on serum leptin and ghrelin concentrations in ad libitum-fed and schedule-fed rats. *Neuroscience* 132:167-173, 2005.
70. Drazen DL, Vahl TP, D'Alessio DA, Seeley RJ, Woods SC. Effects of a fixed meal pattern on ghrelin secretion: evidence for a learned response independent of nutrient status. *Endocrinology* 147:23-30, 2006.
71. Cummings DE, Frayo RS, Marmonier C, Aubert R, Chapelot D. Plasma ghrelin levels and hunger scores in humans initiating meals voluntarily without time- and food-related cues. *Am J Physiol Endocrinol Metab* 287:E297-E304, 2004.
72. Murakami N, Hayashida T, Kuroiwa T, Nakahara K, Ida T, Mondal MS, Nakazato M, Kojima M, Kangawa K. Role for central ghrelin in food intake and secretion profile of stomach ghrelin in rats. *J Endocrinol* 174:283-288, 2002
73. Sanchez J, Oliver P, Pico C, Palou A. Diurnal rhythms of leptin and ghrelin in the systemic circulation and in the gastric mucosa are related to food intake in rats. *Pflugers Arch* 448:500-506, 2004.
74. Williams DL, Cummings DE, Grill HJ, Kaplan JM. Meal-related ghrelin suppression requires postgastric feedback. *Endocrinology* 144:2765-2767, 2003.
75. Sanchez J, Oliver P, Palou A, Pico C. The inhibition of gastric ghrelin production by food intake in rats is dependent on the type of macronutrient. *Endocrinology* 145:5049-5055, 2004.
76. Al Awar R, Obeid O, Hwalla N, Azar S. Postprandial acylated ghrelin status following fat and protein manipulation of meals in healthy young women. *Clin Sci* 109:405-411, 2005.
77. Tannous dit El Khoury D, Obeid O, Azar ST, Hwalla N. Variations in postprandial ghrelin status following ingestion of high-carbohydrate, high-fat, and high-protein meals in males. *Ann Nutr Metab* 50:260-269, 2006.
78. Murray CD, le Roux CW, Gouveia C, Bassett P, Ghatei MA, Bloom SR, Emmanuel AV, Gabe SM. The effect of different macronutrient infusions on appetite, ghrelin and peptide YY in parenterally fed patients. *Clin Nutr* 25:626-633, 2006.
79. Overduin J, Frayo RS, Grill HJ, Kaplan JM, Cummings DE. Role of the duodenum and macronutrient type in ghrelin regulation. *Endocrinology* 146:845-850, 2005.
80. Wortley KE, Anderson KD, Garcia K, Murray JD, Malinova L, Liu R, Moncrieffe M, Thabet K, Cox HJ, Yancopoulos GD, Wiegand SJ, Sleeman MW. Genetic deletion of ghrelin does not decrease food intake but influences metabolic fuel preference. *Proc Natl Acad Sci USA* 101:8227-8232, 2004.
81. De Smet B, Depoortere I, Moechars D, Swennen Q, Moreaux B, Cryns K, Tack J, Buyse J, Coulie B, Peeters TL. Energy homeostasis and gastric emptying in ghrelin knockout mice. *J Pharmacol Exp Ther* 316:431-439, 2006.
82. Zigman JM, Nakano Y, Coppari R, Balthasar N, Marcus JN, Lee CE, Jones JE, Deysher AE, Waxman AR, White RD, Williams TD, Lachey JL, Seeley RJ, Lowell BB, Elmquist JK. Mice lacking ghrelin receptors resist the development of diet-induced obesity. *J Clin Invest* 115:3564-3572, 2005.
83. Zhang JV, Ren PG, Avsian-Kretchmer O, Luo CW, Rauch R, Klein C, Hsueh AJ. Obestatin, a peptide encoded by the ghrelin gene, opposes ghrelin's effects on food intake. *Science* 310:996-999, 2005.
84. Lagaud GJ, Young A, Acena A, Morton MF, Barrett TD, Shankley NP. Obestatin reduces food intake and suppresses body weight gain in rodents. *Biochem Biophys Res Commun* 357:264-269, 2007.
85. Lauwers E, Landuyt B, Arckens L, Schoofs L, Luyten W. Obestatin does not activate orphan G protein-coupled receptor GPR39. *Biochem Biophys Res Commun* 351:21-25, 2006.
86. Gourcerol G, Tache Y. Obestatin – a ghrelin-associated peptide that does not hold its promise to suppress food intake and motility. *Neurogastroenterol Motil* 19:161-165, 2007.
87. Noguieras R, Pfluger P, Tovar S, Arnold M, Mitchell S, Morris A, Perez-Tilve D, Vasquez MJ, Wiedmer P, Casteneda TR, DiMarchi R, Tschop M, Schurmann A, Joost HG, Williams LM, Langhans W, Dieguez C. Effects of obestatin on energy balance and growth hormone secretion in rodents. *Endocrinology* 148:21-26, 2007.
88. Bassil AK, Haglund Y, Brown J, Rudholm T, Hellstrom PM, Naslund E, Lee K, Sanger GJ. Little or no ability of obestatin to interact with ghrelin or modify motility in the rat gastrointestinal tract. *Br J Pharmacol* 150:58-64, 2007.

89. Gourcerol G, St-Pierre DH, Tache Y. Lack of obestatin effects on food intake: should obestatin be renamed ghrelin-associated peptide (GAP)? *Regul Pept* 141:1-7, 2007.
90. Mayer J. Regulation of energy intake and the body weight: the glucostatic theory and the lipostatic hypothesis. *Ann NY Acad Sci* 63:15-43, 1955.
91. Nishizawa Y, Bray GA. Evidence for a circulating ergostatic factor: studies on parabiotic rats. *Am J Physiol* 239:R344-R351, 1980.
92. Benoit SC, Clegg DJ, Seeley RJ, Woods SC. Insulin and leptin as adiposity signals. *Recent Prog Horm Res* 59:267-285, 2004.
93. Baskin DG, Figlewicz Latteman D, Seeley RJ, Woods SC, Porte D Jr, Schwartz MW. Insulin and leptin: dual adiposity signals to the brain for the regulation of food intake and body weight. *Brain Res* 848:114-123, 1999.
94. Havrankova J, Schmechel D, Roth J, Brownstein M. Identification of insulin in rat brain. *Proc Natl Acad Sci USA* 75:5737-5741, 1978.
95. Corp ES, Woods SC, Porte D Jr, Dorsa DM, Figlewicz DP, Baskin DG. Localization of 125I-insulin binding sites in the rat hypothalamus by quantitative autoradiography. *Neurosci Lett* 70:17-22, 1986.
96. Schwartz MW, Seeley RJ, Campfield LA, Burn P, Baskin DG. Identification of targets of leptin action in rat hypothalamus. *J Clin Invest* 98:1101-1106, 1996.
97. Baskin DG, Hahn TM, Schwartz MW. Leptin sensitive neurons in the hypothalamus. *Horm Metab Res* 31:345-350, 1999.
98. Woods SC, Lotter EC, McKay LD, Porte D Jr. Chronic intracerebroventricular infusion of insulin reduces food intake and body weight of baboons. *Nature* 282:503-505, 1979.
99. Air EL, Benoit SC, Blake Smith KA, Clegg DJ, Woods SC. Acute third ventricular administration of insulin decreases food intake in two paradigms. *Pharmacol Biochem Behav* 72:423-429, 2002.
100. Air EL, Benoit SC, Clegg DJ, Seeley RJ, Woods SC. Insulin and leptin combine additively to reduce food intake in rats. *Endocrinology* 143:2449-2452, 2002.
101. Campfield LA, Smith FJ, Guisez Y, Devos R, Burn P. Recombinant mouse OB protein: evidence for a peripheral signal linking adiposity and central neural networks. *Science* 269:546-549, 1995.
102. McGowan MK, Andrews KM, Grossman SP. Chronic intrahypothalamic infusions of insulin or insulin antibodies alter body weight and food intake in the rat. *Physiol Behav* 51:753-766, 1992.
103. Strubbe JH, Mein CG. Increased feeding in response to bilateral injection of insulin antibodies in the VMH. *Physiol Behav* 19:309-313, 1977.
104. Coleman DL. Obese and diabetes: two mutant genes causing diabetes-obesity syndromes in mice. *Diabetologia* 14:141-148, 1978.
105. Zucker LM, Zucker TF. Fatty, a new mutation in the rat. *J Hered* 52:275-278, 1961
106. Chua SC, Chung WK, Wu-Peng XS, Zhang Y, Liu SM, Tartaglia L, Leibel RL. Phenotypes of mouse diabetes and rat fatty due to mutations in the OB (leptin) receptor. *Science* 271:994-996, 1996.
107. Bruning JC, Gautam D, Burks DJ, Gillette J, Schubert M, Orban PC, Klein R, Krone W, Muller-Wieland D, Kahn CR. Role of brain insulin receptor in control of body weight and reproduction. *Science* 289:2122-2125, 2000.
108. Lin X, Taguchi A, Park S, Kushner JA, Li F, Li Y, White MF. Dysregulation of insulin receptor substrate 2 in beta cells and brain causes obesity and diabetes. *J Clin Invest* 114:908-916, 2004.
109. Montague CT, Farooqi IS, Whitehead JP, Soos MA, Rau H, Wareham NJ, Sewter CP, Digby JE, Mohammed SN, Hurst JA, Cheetham CH, Earley AR, Barnett AH, Prieur JB, O'Rahilly S. Congenital leptin deficiency is associated with severe early-onset obesity in humans. *Nature* 387:903-908, 1997.
110. Farooqi IS, Wangenstein T, Collins S, Kimber W, Matarese G, Keogh JM, Lank E, Bottomley B, Lopez-Fernandez J, Ferraz-Amaro I, Dattani MT, Ercan O, Myhre AG, Retterstol L, Stanhope R, Edge JA, McKenzie S, Lessan N, Ghodsi M, De

Rosa V, Perna F, Fontana S, Barrosos I, Undlien DE, O'Rahilly S. Clinical and molecular genetic spectrum of congenital deficiency of the leptin receptor. *N Engl J Med* 356:237-247, 2007.

111. Cheung CC, Clifton DK, Steiner RA. Proopiomelanocortin neurons are direct targets for leptin in the hypothalamus. *Endocrinology* 138:4489-4492, 1997.

112. Marks JL, Porte D Jr, Stahl WL, Baskin DG. Localization of insulin receptor mRNA in rat brain by in situ hybridization. *Endocrinology* 127:3234-3236, 1990.

113. Pardini AW, Nguyen HT, Figlewicz DP, Baskin DG, Williams DL, Kim F, Schwartz MW. Distribution of insulin receptor substrate-2 in brain areas involved in energy homeostasis. *Brain Res* 1112:169-178, 2006.

114. Williams KW, Margatho LO, Lee CE, Choi M, Lee S, Scott MM, Elias CF, Elmquist JK. Segregation of acute leptin and insulin effects in distinct populations of arcuate proopiomelanocortin neurons. *J Neurosci* 30:2472-2479, 2010.

115. Stellar E. The physiology of motivation. *Psychol Rev* 61:5-22, 1954.

116. Stanley BG, Leibowitz SF. Neuropeptide Y: stimulation of feeding and drinking by injection into the paraventricular nucleus. *Life Sci* 35:2635-2642, 1984.

117. Rossi M, Kim MS, Morgan DG, Small CJ, Edwards CM, Sunter D, Abusnana S, Goldstone AP, Russell SH, Stanely SA, Smith DM, Yagaloff K, Ghatel MA, Bloom SR. A C-terminal fragment of Agouti-related protein increases feeding and antagonizes the effect of alpha-melanocyte stimulating hormone in vivo. *Endocrinology* 139:4428-4431, 1998.

118. Ollmann M, Wilson B, Yang Y, Kerns J, Chen Y, Gantz I, Barsh G. Antagonism of central melanocortin receptors in vitro and in vivo by agouti-related protein. *Science* 278:135-138, 1997.

119. McMinn JE, Wilkinson CW, Havel PJ, Woods SC, Schwartz MW. Effect of intracerebroventricular alpha-MSH on food intake, adiposity, c-Fos induction, and neuropeptide expression. *Am J Physiol Regul Integr Comp Physiol* 279:R695-R703, 2000.

120. Lambert PD, Couceyro PR, McGirr KM, Dall Vechia SE, Smith Y, Kuhar MJ. CART peptides in the central control of feeding and interactions with neuropeptide Y. *Synapse* 29:293-298, 1998.

121. Hahn TM, Breininger JF, Baskin DG, Schwartz MW. Coexpression of AgRP and NPY in fasting-activated hypothalamic neurons. *Nat Neurosci* 1:271-272, 1998.

122. Brady LS, Smith MA, Gold PW, Herkenham M. Altered expression of hypothalamic neuropeptide mRNAs in food-restricted and food-deprived rats. *Neuroendocrinology* 52:441-447, 1990.

123. Kim EM, Welch CC, Grace MK, Billington CJ, Levine AS. Chronic food restriction and acute food deprivation decrease mRNA levels of opioid peptides in arcuate nucleus. *Am J Physiol* 270:R1019-R1024, 1996.

124. Li HY, Hwang HW, Hu YH. Functional characterization of cocaine- and amphetamine-regulated transcript mRNA expression in rat hypothalamus. *Neurosci Lett* 323:203-206, 2002.

125. Miltenberger RJ, Mynatt RL, Wilkinson JE, Woychik RP. The role of the agouti gene in the yellow obese syndrome. *J Nutr* 127:1902S-1907S, 1997.

126. Graham M, Shutter JR, Sarmiento U, Sarosi I, Stark KL. Overexpression of AgRP leads to obesity in transgenic mice. *Nat Genet* 17:273-274, 1997.

127. Huszar D, Lynch C, Fairchild-Huntress V, Dunmore J, Fang Q, Berkemeier L, Gu W, Kesterson R, Boston B, Cone R, Smith F, Campfield L, Burn P, Lee F. Targeted disruption of the melanocortin-4 receptor results in obesity in mice. *Cell* 88:131-141, 1997.

128. Yaswen L, Diehl N, Brennan MB, Hochgeschwender U. Obesity in the mouse model of proopiomelanocortin deficiency responds to peripheral melanocortin. *Nat Med* 5:1066-1070, 1999.

129. Wierup N, Richards WG, Bannon AW, Kuhar MJ, Ahren B, Sundler F. CART knock out mice have impaired insulin secretion and glucose intolerance, altered beta cell morphology and increased body weight. *Regul Pept* 129:203-211, 2005.

130. Aponte Y, Atasoy D, Sternson SM. AGRP neurons are sufficient to orchestrate feeding behavior rapidly and without training. *Nat Neurosci* 14:351-355, 2011.

131. Palmiter RD, Erickson JC, Hollopeter G, Baraban SC, Schwarz MW. Life without neuropeptide Y. *Recent Prog Horm Res* 53:163-199, 1998.
132. Qian S, Chen H, Weingarh D, Trumbauer ME, Novi DE, Guan X, Yu H, Shen Z, Feng Y, Frazier E, Chen A, Camacho RE, Shearman LP, Gopal-Truter S, MacNeil DJ, Van der Ploeg LH, Marsh DJ. Neither agouti-related protein nor neuropeptide Y is critically required for the regulation of energy homeostasis in mice. *Mol Cell Biol* 22:5027-5035, 2002.
133. Gropp E, Shanabrough M, Borok E, Xu AW, Janoschek R, Buch T, Plum L, Balthasar N, Hampel B, Waisman A, Barsh GS, Horvath TL, Bruning JC. Agouti-related peptide-expressing neurons are mandatory for feeding. *Nat Neurosci* 8:1289-1291, 2005.
134. Luguët S, Perez FA, Hnasko TS, Palmiter RD. NPY/AgRP neurons are essential for feeding in adult mice but can be ablated in neonates. *Science* 310:683-685, 2005.
135. Farooqi IS, O'Rahilly S. Genetics of obesity in humans. *Endocr Rev* 27:710-718, 2006.
136. Banks WA, Kastin AJ, Huang W, Jaspan JB, Maness LM. Leptin enters the brain by a saturable system independent of insulin. *Peptides* 17:305-311, 1996.
137. Banks WA, Jaspan JB, Huang W, Kastin AJ. Transport of insulin across the blood-brain barrier: saturability at euglycemic doses of insulin. *Peptides* 18:1423-1429, 1997.
138. Benoit SC, Air EL, Coolen LM, Strauss R, Jackman A, Clegg DJ, Seeley RJ, Woods SC. The catabolic action of insulin in the brain is mediated by melanocortins. *J Neurosci* 22: 9048-9052, 2002.
139. Schwartz MW, Seeley RJ, Weigle DS, Burn P, Campfield LA, Baskin DG. Leptin increases hypothalamic proopiomelanocortin (POMC) mRNA expression in the rostral arcuate nucleus. *Diabetes* 46:2119-2123, 1997.
140. Cowley MA, Smart JL, Rubenstein M, Cerdan MG, Diano S, Horvath TL, Cone RD, Low MJ. Leptin activates anorexigenic POMC neurons through a neural network in the arcuate nucleus. *Nature* 411:480-484, 2001.
141. Seeley RJ, Yagaloff K, Fisher S, Burn P, Thiele T, van Dijk G, Baskin DG, Schwartz MW. Melanocortin receptors in leptin effects. *Nature* 390:349, 1997.
142. Heinrichs SC, Menzaghi F, Pich EM, Hauger RL, Koob GF. Corticotropin-releasing factor in the paraventricular nucleus modulates feeding induced by neuropeptide Y. *Brain Res* 611:18-24, 1993.
143. Uehara Y, Shimizu H, Ohtani K, Sato N, Mori M. Hypothalamic corticotrophin-releasing hormone is a mediator of the anorexigenic effects of leptin. *Diabetes* 47:890-893, 1998.
144. Mihaly E, Fekete C, Tatro JB, Liposits Z, Stopa EG, Lechan RM. Hypophysiotropic thyrotropin-releasing hormone-synthesizing neurons in human hypothalamus are innervated by neuropeptide Y, agouti-related protein, and alpha-melanocyte-stimulating hormone. *J Clin Endocrinol Metab* 85:2596-2603, 2000.
145. Vijayan E, McCann SM. Suppression of feeding and drinking activity in rats following intraventricular injection of thyrotropin releasing hormone (TRH). *Endocrinology* 100:1727-1730, 1977.
146. Olson BR, Drutarosky MD, Chow MS, Hruby VJ, Stricker EM, Verbalis JG. Oxytocin and an oxytocin agonist administered centrally decrease food intake in rats. *Peptides* 12:113-118, 1991.
147. Arletti R, Benelli A, Bertolini A. Influence of oxytocin on feeding behavior in the rat. *Peptides* 10:89-93, 1989.
148. Qu D, Ludwig DS, Gammeltoft S, Piper M, Pelleymounter MA, Cullen MJ, Mathes WF, Przyspek R, Kanarek R, Maratos-Flier JS. A role for melanin-concentrating hormone in the central regulation of feeding behavior. *Nature* 380:243-247, 1996.
149. de Lecea L, Kilduff TS, Peyton C, Gao X, Foye PE, Danielson PE, Fukuhara C, Battenberg EL, Gautvik VT, Bartlett FS 2nd, Frankel WN, van den Pol AN, Bloom FE, Gautvik KM, Sutcliffe JG. The hypocretins: hypothalamus-specific peptides with neuroexcitatory activity. *Proc Natl Acad Sci USA* 95:322-327, 1998.
150. Broberger C, de Lecea L, Sutcliffe JG, Hokfelt T. Hypocretin/orexin- and melanin-concentrating hormone-expressing cells form distinct populations in the rodent lateral hypothalamus: relationship to the neuropeptide Y and agouti gene-related protein synthesis. *J Comp Neurol* 402:460-474, 1998.
151. Sakurai T, Amemiya A, Ishii M, Matsuzaki I, Chemelli RM, Tanaka H, Williams SC, Richardson JA, Kozlowski GP, Wilson S, Arch JR, Buckingham RE, Haynes AC, Carr SA, Annan RS, McNulty DE, Liu WS, Terrett JA, Elshourbagy NA, Bergsma DJ,

Yanagisawa M. Orexin and orexin receptors: a family of hypothalamic neuropeptides and G protein-coupled receptors that regulate feeding behavior. *Cell* 92:572-585, 1998.

152. Nakazato M, Murakami N, Date Y, Kojima M, Matsuo H, Kanagawa K, Matsukura S. A role for ghrelin in the central regulation of feeding. *Nature* 409:194-198, 2001.

153. Seoane LM, Lopez M, Tovar S, Casanueva FF, Senaris R, Dieguez C. Agouti-related peptide, neuropeptide Y, and somatostatin-producing neurons are targets for ghrelin actions in the rat hypothalamus. *Endocrinology* 144:544-551, 2003.

154. Mondal MS, Date Y, Yamaguchi H, Toshinai K, Tsuruta T, Kangawa K, Nakazato M. Identification of ghrelin and its receptor in neurons of the rat arcuate nucleus. *Regul Pept* 126:55-59, 2005.

155. Hewson AK, Dickson SL. Systemic administration of ghrelin induces Fos and Egr-1 proteins in the hypothalamic arcuate nucleus of fasted and fed rats. *J Endocrinol* 12:1047-1049, 2000.

156. Chen HY, Trumbauer ME, Chen AS, Weingarth DT, Adams JR, Frazier EG, Shen Z, Marsh DJ, Feighner SD, Guan XM, Ye Z, Nargund RP, Smith RG, Van der Ploeg LH, Howard AD, MacNeil DJ, Qian S. Orexigenic action of peripheral ghrelin is mediated by neuropeptide Y and agouti-related protein. *Endocrinology* 144:2607-2612, 2004.

157. Oh-I S, Shimizu H, Satoh T, Okada S, Adachi S, Inoue K, Eguchi H, Yamamoto M, Imake T, Hashimoto K, Tsuchiya T, Monden T, Horiguchi K, Yamada M, Mori M. Identification of nesfatin-1 as a satiety molecule in the hypothalamus. *Nature* 443:709-712, 2006.

158. Stengel A, Goebel M, Yakubov I, Wang L, Witcher D, Coskun T, Tache Y, Sachs G, Lambrecht NW. Identification and characterization of nesfatin-1 immunoreactivity in endocrine cell types of the rat gastric oxyntic mucosa. *Endocrinology* 150:232-238, 2009.

159. Price TO, Samson WK, Niehoff ML, Banks WA. Permeability of the blood-brain barrier to a novel satiety molecule nesfatin-1. *Peptides* 28:2372-2381, 2007.

160. Price CJ, Hoyda TD, Samson WK, Ferguson AV. Nesfatin-1 influences the excitability of paraventricular nucleus neurones. *J Endocrinol* 20:245-250, 2008.

161. Price CJ, Samson WK, Ferguson AV. Nesfatin-1 inhibits NPY neurons in the arcuate nucleus. *Brain Res* 1230:99-106, 2008.

162. Shimizu H, Ohsaki A, Oh-I S, Okada S, Mori M. A new anorexigenic protein, nesfatin-1. *Peptides* 30:995-998, 2009.

163. Goebel M, Stengel A, Wang L, Tache Y. Central nesfatin-1 reduces the nocturnal food intake in mice by reducing meal size and increasing inter-meal intervals. *Peptides* 32:36-43, 2011.

164. Schwartz GJ. The role of gastrointestinal vagal afferents in the control of food intake: current prospects. *Nutrition* 16:866-873, 2000.

165. Grill HJ, Smith GP. Cholecystokinin decreases sucrose intake in chronic decerebrate rats. *Am J Physiol* 254:R853-R856, 1988.

166. Seeley RJ, Grill HJ, Kaplan JM. Neurological dissociation of gastrointestinal and metabolic contributions to meal size control. *Behav Neurosci* 108:347-352, 1994.

167. Berthoud HR. Multiple neural systems controlling food intake and body weight. *Neurosci Biobehav Rev* 26:393-428, 2002.

168. Schwartz GJ. Integrative capacity of the caudal brainstem in the control of food intake. *Philos Trans R Soc Lond B Biol Sci* 361:1275-1280, 2006.

169. Grill HJ. Distributed neural control of energy balance: contributions from hindbrain and hypothalamus. *Obesity* 14:216S-221S, 2006.

170. Grill HJ, Schwartz MW, Kaplan JM, Foxhall JS, Breininger J, Baskin DG. Evidence that the caudal brainstem is a target for the inhibitory effect of leptin on food intake. *Endocrinology* 143:239-246, 2002.

171. Jacobowitz DM, O'Donohue TL. Alpha-Melanocyte stimulating hormone: immunohistochemical identification and mapping in neurons of rat brain. *Proc Natl Acad Sci USA* 75:6300-6304, 1978.

172. Palkovits M, Mezey E, Eskay RL. Pro-opiomelanocortin-derived peptides (ACTH/beta-endorphin/alpha-MSH) in brainstem baroreceptor areas of the rat. *Brain Res* 436:323-338, 1987.

173. Mountjoy KG, Mortrud MT, Low MJ, Simerly RB, Cone RD. Localization of the melanocortin-4 receptor (MC4-R) in neuroendocrine and autonomic control circuits in the brain. *Mol Endocrinol* 8:503-508, 1994.
174. Kishi T, Aschkenasi CJ, Choi BJ, Lopez ME, Lee CE, Liu H, Hollenberg AN, Friedman JM, Elmquist JK. Neuropeptide Y Y1 receptor mRNA in rodent brain: distribution and colocalization with melanocortin-4 receptor. *J Comp Neurol* 482:217-243, 2005.
175. Kishi T, Aschkenasi CJ, Lee CE, Mountjoy KG, Saper CB, Elmquist JK. Expression of melanocortin 4 receptor mRNA in the central nervous system of the rat. *J Comp Neurol* 457:213-235, 2003.
176. Grill HJ, Ginsberg AB, Seeley RJ, Kaplan JM. Brainstem application of melanocortin receptor ligands produces long-lasting effects on feeding and body weight. *J Neurosci* 18:10128-10135, 1998.
177. Taylor K, Lester E, Hudson B, Ritter S. Hypothalamic and hindbrain NPY, AGRP and NE increase consummatory feeding responses. *Physiol Behav* 90:744-750, 2007.
178. Corp ES, Melville LD, Greenberg D, Gibbs J, Smith GP. Effect of fourth ventricular neuropeptide Y and peptide YY on ingestive and other behaviors. *Am J Physiol* 259:R317-R323, 1990.
179. Corp ES, McQuade J, Krasnicki S, Conze DB. Feeding after fourth ventricular administration of neuropeptide Y receptor agonists in rats. *Peptides* 22:493-499, 2001.
180. Moran TH, Aja S, Ladenheim EE. Leptin modulation of peripheral controls of meal size. *Physiol Behav* 89:511-516, 2006.
181. Schwartz GJ, Moran TH. Leptin and neuropeptide Y have opposing modulatory effects on nucleus of the solitary tract neurophysiological responses to gastric loads: implications for the control of food intake. *Endocrinology* 143:3779-3784, 2002.
182. Huo L, Maeng L, Bjorbaek C, Grill HJ. Leptin and the control of food intake: neurons in the nucleus of the solitary tract are activated by both gastric distension and leptin. *Endocrinology* 148:2189-2197, 2007.
183. Emond M, Ladenheim EE, Schwartz GJ, Moran TH. Leptin amplifies the feeding inhibition and neural activation arising from a gastric nutrient preload. *Physiol Behav* 72:123-128, 2001.
184. Emond M, Schwartz GJ, Ladenheim EE, Moran TH. Central leptin modulates behavioral and neural responsivity to CCK. *Am J Physiol* 276:R1545-R1549, 1999.
185. Scott MM, Williams KW, Rossi J, Lee CE, Elmquist JK. Leptin receptor expression in hindbrain Glp-1 neurons regulates food intake and energy balance in mice. *J Clin Invest* 121:2413-2421, 2011.
186. Fry M, Ferguson AV. The sensory circumventricular organs: brain targets for circulating signals controlling ingestive behavior. *Physiol Behav* 91:413-423, 2007.
187. Asin KE, Gore PA Jr, Bednarz L, Holladay M, Nadzan AM. Effects of selective CCK receptor agonists on food intake after central or peripheral administration in rats. *Brain Res* 571:169-174, 1992.
188. Ebenezer IS. Effects of intracerebroventricular administration of the CCK(1) receptor antagonist devazepide on food intake in rats. *Eur J Pharmacol* 441:79-82, 2002.
189. Corp ES, Curcio M, Gibbs J, Smith GP. The effect of centrally administered CCK-receptor antagonists on food intake in rats. *Physiol Behav* 61:823-827, 1997.
190. Brenner LA, Ritter RC. Intracerebroventricular cholecystokinin A-receptor antagonist does not reduce satiation by endogenous CCK. *Physiol Behav* 63:711-716, 1998.
191. Crawley JN, Fiske SM, Durieux C, Derrien M, Roques BP. Centrally administered cholecystokinin suppresses feeding through a peripheral-type receptor mechanism. *J Pharmacol Exp Ther* 257:1076-1080, 1991.
192. Lindefors N, Linden A, Brene S, Sedvall G, Persson H. CCK peptides and mRNA in the human brain. *Prog Neurobiol* 40:671-690, 1993.
193. Crawley JN, Corwin RL. Biological actions of cholecystokinin. *Peptides* 15:731-755, 1994.
194. Moran TH, Schwartz GJ. Neurobiology of cholecystokinin. *Crit Rev Neurobiol* 9:1-28, 1994.
195. Beinfeld MC, Meyer DK, Eskay RL, Jensen RT, Brownstein MJ. The distribution of cholecystokinin immunoreactivity in the central nervous system of the rat as determined by radioimmunoassay. *Brain Res* 212:51-57, 1981.

196. Dauge V, Lena I. CCK in anxiety and cognitive processes. *Neurosci Biobehav Rev* 22:815-825, 1998.
197. Hebb AL, Poulin JF, Roach SP, Zacharko RM, Drolet G. Cholecystokinin and endogenous opioid peptides: interactive influence on pain, cognition and emotion. *Prog Neuropsychopharmacol Biol Psychiatry* 29:1225-1238, 2005.
198. Challis BG, Pinnock SB, Coll AP, Carter RN, Dickson SL, O'Rahilly S. Acute effects of PYY3-36 on food intake and hypothalamic neuropeptide expression in the mouse. *Biochem Biophys Res Commun* 311:915-919, 2003.
199. Stanley BG, Daniel DR, Chin AS, Leibowitz SF. Paraventricular nucleus injections of peptide YY and neuropeptide Y preferentially enhance carbohydrate ingestion. *Peptides* 6:1205-1211, 1985.
200. Morley JE, Levine AS, Grace M, Kneip J. Peptide YY (PYY), a potent orexigenic agent. *Brain Res* 341:200-203, 1985.
201. Uttenthal LO, Toledano A, Blazquez E. Autoradiographic localization of receptors for glucagon-like peptide-1 (7-36) amide in rat brain. *Neuropeptides* 21:143-146, 1992.
202. Turton MD, O'Shea D, Gunn I, Beak SA, Edwards CM, Meeran K, Choi SJ, Taylor GM, Heath MM, Lambert PD, Wilding JP, Smith DM, Ghatei MA, Herbert J, Bloom SR. A role for glucagon-like peptide-1 in the central regulation of feeding. *Nature* 379:68-72, 1996.
203. Scoville WB, Milner B. Loss of recent memory after bilateral hippocampal lesions. *J Neurol Neurosurg Psychiatry* 20:11-21, 1957.
204. Hebden N, Corkin S, Eichenbaum H, Shedlack K. Diminished ability to interpret and report internal states after bilateral medial temporal resection: case H.M. *Behav Neurosci* 99:1031-1039.
205. Clifton PG, Vickers SP, Somerville EM. Little and often: ingestive behavior patterns following hippocampal lesions in rats. *Behav Neurosci* 112:502-511, 1998.
206. Davidson TL, Jarrard LE. A role for the hippocampus in the utilization of hunger signals. *Behav Neural Biol* 59:167-171, 1993.
207. King BM, Hoan S, Arceneaux ER, Kass JM. Effect on food intake and body weight of lesions in and adjacent to the posterodorsal amygdala in rats. *Physiol Behav* 55:963-966, 1994.
208. King BM, Arceneaux ER, Cook JT, Benjamin AL, Alheid GF. Temporal lobe lesion-induced obesity in rats: an anatomical investigation of the posterior amygdala and hippocampal formation. *Physiol Behav* 59:843-848, 1996.
209. Forloni G, Fisone G, Guaitani A, Ladinsky H, Consolo S. Role of the hippocampus in the sex-dependent regulation of eating behavior: studies with kainic acid. *Physiol Behav* 38:321-326, 1986.
210. Davidson TL, Chan K, Jarrard LE, Kanoski SE, Clegg DJ, Benoit SC. Contributions of the hippocampus and medial prefrontal cortex to energy and body weight regulation. *Hippocampus* 19:235-252, 2009.
211. Davidson TL, Kanoski SE, Chan K, Clegg DJ, Benoit SC, Jarrard LE. Hippocampal lesions impair retention of discriminative responding based on energy state cues. *Behav Neurosci* 124:97-105, 2010.
212. Stice E, Burger K, Yokum S. Caloric deprivation increases responsivity of attention and reward brain regions to intake, anticipated intake and images of palatable foods. *Neuroimage* 67:322-330, 2013.
213. Moser MB, Moser EI. Functional differentiation in the hippocampus. *8:608-619*, 1998.
214. Cenquizca LA, Swanson LW. Analysis of direct hippocampal cortical field CA1 axonal projections to diencephalon in the rat. *J Comp Neurol* 497:101-114, 2006.
215. Kanoski SE, Hayes MR, Greenwald HS, Fortin SM, Gianessi CA, Gilbert JR, Grill HJ. Hippocampal leptin signaling reduces food intake and modulates food-related memory processing. *Neuropsychopharmacology* 36:1859-1870, 2011.
216. Kanoski SE, Fortin SM, Ricks KM, Grill HJ. Ghrelin signaling in the ventral hippocampus stimulates learned and motivational aspects of feeding via PI3K-Akt signaling. *Biol Psychiatry* 73:915-923, 2013.
217. Henderson YO, Smith GP, Parent MB. Hippocampal neurons inhibit meal onset. *Hippocampus* 23:100-107, 2013.
218. Asarian L, Geary N. Modulation of appetite by gonadal steroid hormones. *Philos Trans R Soc Lond B Biol Sci* 361:1251-1263, 2006.

219. Heine PA, Taylor JA, Iwamoto GA, Lubahn DB, Cooke PS. Increased adipose tissue in male and female estrogen receptor- α knockout mice. *Proc Natl Acad Sci USA* 97:12729-12734, 2000.
220. Santollo J, Torregrossa AM, Eckel LA. Estradiol acts in the medial preoptic area, arcuate nucleus and dorsal raphe nucleus to reduce food intake in ovariectomized rats. *Horm Behav* 60:86-93, 2011.
221. Brown LM, Clegg DJ. Central effects of estradiol in the regulation of food intake, body weight, and adiposity. *J Steroid Biochem Mol Biol* 122:65-73, 2010.
222. Clegg DJ, Riedy CA, Blake Smith KA, Benoit SC, Woods SC. Differential sensitivity to central leptin and insulin in male and female rats. *Diabetes* 52:682-687, 2003.
223. Clegg DJ, Brown LM, Woods SC, Benoit SC. Gonadal hormones determine sensitivity to central leptin and insulin. *Diabetes* 55:978-987, 2006.
224. Crandall DL, Busler DE, Novak TJ, Weber RV, Kral JG. Identification of estrogen receptor β in human breast and abdominal subcutaneous adipose tissue. *Biochem Biophys Res Commun* 248:523-526, 1998.
225. Halford JC, Harrold JA, Boyland EJ, Lawton CL, Blundell JE. Serotonergic drugs: effects on appetite expression and use for the treatment of obesity. *Drugs* 67:27-55, 2007.
226. Donovan MH, Tecott LH. Serotonin and the regulation of mammalian energy balance. *Front Neurosci* 7:36, 2013.
227. Roth JD, Rowland NE. Efficacy of administration of dexfenfluramine and phentermine, alone or in combination on ingestive behavior and body weight in rats. *Psychopharmacology* 137:99-106, 1998.
228. Weintraub M, Hasday JD, Mushlin AI, Lockwood DH. A double-blind clinical trial in weight control. Use of fenfluramine and phentermine alone and in combination. *Arch Intern Med* 144:1143-1148, 1984.
229. Connolly HM, Crary JL, McGoon MD, Hensrud DD, Edwards BS, Edwards WD, Schaff HV. Valvular heart disease associated with fenfluramine-phentermine. *N Engl J Med* 337:581-588, 1997.
230. Smith SR, Prosser WA, Donahue DJ, Morgan ME, Anderson CM, Shanahan WR, APD356-004 Study Group. Lorcaserin (APD356), a selective 5-HT(2C) agonist, reduces body weight in obese men and women. *Obesity* 17:494-503, 2009.
231. Fidler MC, Sanchez M, Raether B, Weissman NJ, Shanahan WR, Anderson CM, BLOSSOM Clinical Trial Group. A one-year randomized trial of lorcaserin for weight loss in obese and overweight adults: the BLOSSOM trial. *J Clin Endocrinol Metab* 96:3067-3077, 2011.
232. Terry P, Gilbert DB, Cooper SJ. Dopamine receptor subtype agonists and feeding behavior. *Obes Res* 3:515S-523S, 1995.
233. McQuade JA, Benoit SC, Woods SC, Seeley RJ. 7-OH-DPAT selectively reduces intake of both chow and high fat diets in different food intake regimens. *Pharmacol Biochem Behav* 76:517-523, 2003.
234. Ball KT, Combs TA, Beyer DN. Opposing roles for dopamine D1- and D2-like receptors in discrete cue-induced reinstatement of food seeking. *Behav Brain Res* 222:390-393, 2011.
235. Baldo BA, Kelley AE. Discrete neurochemical coding of distinguishable motivational processes: insights from nucleus accumbens control of feeding. *Psychopharmacology* 191:439-459, 2007.
236. Wise RA. Role of brain dopamine in food reward and reinforcement. *Philos Trans R Soc Lond B Biol Sci* 361:1149-1158, 2006.
237. Mogg K, Bradley BP, O'Neill B, Bani M, Merlo-Pich E, Koch A, Bullmore ET, Nathan PJ. Effect of dopamine D₃ receptor antagonism on approach responses to food cues in overweight and obese individuals.
238. Smith GP, Smith JC. The inhibitory potency of SCH 23390 and raclopride on licking for sucrose increases across brief-access tests. *Phys Behav* 101:315-319, 2010.
239. Geary N, Smith GP. Pimozide decreases the positive reinforcing effect of sham feeding sucrose in the rat. *Pharmacol Biochem Behav* 22:787-790, 1985.
240. Perry ML, Baldo BA, Andrzejewski ME, Kelley AE. Muscarinic receptor antagonism causes a functional alteration in nucleus accumbens mu-opiate-mediated feeding behavior. *Behav Brain Res* 197:225-229, 2009.

241. Perry ML, Andrzejewski ME, Bushek SM, Baldo BA. Intra-accumbens infusion of a muscarinic antagonist reduces food intake without altering the incentive properties of food-associated cues. *Behav Neurosci* 124:44-54, 2010.
242. Pratt WE, Blackstone K. Nucleus accumbens acetylcholine and food intake: decreased muscarinic tone reduces feeding but not food seeking. *Behav Brain Res* 198:252-257, 2009.
243. Dickson SL, Hrabovszky E, Hansson C, Jerlhag E, Alvarez-Crespo M, Skibicka KP, Molnar CS, Liposits Z, Engel JA, Egecioglu E. Blockade of central nicotine acetylcholine receptor signaling attenuate ghrelin-induced food intake in rodents. *Neuroscience* 171:1180-1186, 2010.
244. Mineur YS, Abizaid A, Rao Y, Salas R, DiLeone RJ, Gundisch D, Diano S, De Biasi M, Horvath TL, Gao X-B, Picciotto MR. Nicotine decreases food intake through activation of POMC neurons. *Science* 332:1330-1332, 2011.
245. Barbano MF, Cador M. Opioids for hedonic experience and dopamine to get ready for it. *Psychopharmacology* 191:497-506, 2007.
246. Gosnell BA, Levine AS. Reward systems and food intake: role of opioids. *Int J Obes* 33:S54-S58, 2009.
247. Kirkham TC, Williams CM. Synergistic effects of opioid and cannabinoid antagonists on food intake. *Psychopharmacology* 153:267-270, 2001.
248. Solinas M, Goldberg SR. Motivational effects of cannabinoids and opioids on food reinforcement depend on simultaneous activation of cannabinoid and opioid systems. *Neuropsychopharmacology* 30:2035-2045, 2005.
249. DiMarzo V, Matias I. Endocannabinoid control of food intake and energy balance. *Nat Neurosci* 8:585-589, 2005.
250. Cota D, Marsicano G, Lutz B, Vicennati V, Stalla GK, Pasquali R, Pagotto U. Endogenous cannabinoid system as a modulator of food intake. *Int J Obes Relat Metab Disord* 27:289-301, 2003.
251. Patel PN, Pathak R. Rimonabant: a novel selective cannabinoid-1 receptor antagonist for treatment of obesity. *Am J Health Syst Pharm* 64:481-489, 2007.
252. Cota D, Tschop MH, Horvath TL, Levine AS. Cannabinoids, opioids and eating behavior: the molecular face of hedonism? *Brain Res Rev* 51:85-107, 2006.
253. Levitsky DA. Feeding patterns of rats in response to fasts and changes in environmental conditions. *Physiol Behav* 5:291-300, 1970.
254. Tagliaferro AR, Levitsky DA. Overcompensation of food intake following brief periods of food restriction. *Physiol Behav* 29:747-750, 1982.
255. Farley C, Cook JA, Spar BD, Austin TM, Kowalski TJ. Meal pattern analysis of diet-induced obesity in susceptible and resistant rats. *Obes Res* 11:845-851, 2003.
256. Howarth NC, Huang TT, Roberts SB, Lin BH, McCrory MA. Eating patterns and dietary composition in relation to BMI in younger and older adults. *Int J Obes* 31:675-684, 2007.
257. Pearcey SM, de Castro JM. Food intake and meal patterns of weight-stable and weight-gaining persons. *Am J Clin Nutr* 76:107-112, 2002.
258. Woods SC, Seeley RJ. Understanding the physiology of obesity: review of recent developments in obesity research. *Int J Obes Relat Metab Disord* 26:S8-S10, 2002.
259. Stein LJ, Woods SC. Gastrin releasing peptide reduces meal size in rats. *Peptides* 3:833-835, 1982.
260. Scott KA, Moran TH. The GLP-1 agonist exendin-4 reduces food intake in nonhuman primates through changes in meal size. *Am J Physiol Regul Integr Comp Physiol* 293:R983-R987, 2007.
261. Flynn MC, Plata-Salaman CR. Leptin (OB protein) and meal size. *Nutrition* 15:508-509, 1999.
262. Kahler A, Geary N, Eckel LA, Campfield LA, Smith FJ, Langhans W. Chronic administration of OB protein decreases food intake by selectively reducing meal size in male rats. *Am J Physiol* 275:R180-R185, 1998.
263. Eckel LA, Langhans W, Kahler A, Campfield LA, Smith FJ, Geary N. Chronic administration of OB protein decreases food intake by selectively reducing meal size in female rats. *Am J Physiol* 275:R186-R193, 1998.

264. de Krom M, van der Schouw YT, Hendrks J, Ophoff RA, van Gils CH, Stolk RP, Grobbee DE, Adan R. Common genetic variations in CCK, leptin, and leptin receptor genes are associated with specific human eating patterns. *Diabetes* 56:276-280, 2007.
265. Azzara AV, Sokolnicki JP, Schwartz GJ. Central melanocortin receptor agonist reduces spontaneous and scheduled meal size but does not augment duodenal preload-induced feeding inhibition. *Physiol Behav* 77:411-416, 2002.
266. Williams DL, Grill HJ, Weiss SM, Baird JP, Kaplan JM. Behavioral processes underlying the intake suppressive effects of melanocortin 3/4 receptor activation in the rat. *Psychopharmacology* 161:47-53, 2002.
267. Aja S, Schwartz GJ, Kuhar MJ, Moran TH. Intracerebroventricular CART peptide reduces rat ingestive behavior and alters licking microstructure. *Am J Physiol Regul Comp Integr Comp Physiol* 280:R1613-R1619, 2001.
268. Baird JP, Gray NE, Fischer SG. Effects of neuropeptide Y on feeding microstructure: dissociation of appetitive and consummatory actions. *Behav Neurosci* 120:937-951, 2006.
269. Davis JF, Choi DL, Clegg DJ, Benoit SC. Signaling through the ghrelin receptor modulates hippocampal function and meal anticipation in mice. *Physiol Behav* 103:39-43, 2011.
270. Ott V, Friedrich M, Zemlin J, Lehnert H, Schultes B, Born J, Hallschmid M. Meal anticipation potentiates postprandial ghrelin suppression in humans. *Psychoneuroendocrinology*, 37:1096-1100, 2012.
271. Vahl TP, Drazen DL, Seeley RJ, D'Alessio DA, Woods SC. Meal-anticipatory glucagon-like peptide-1 secretion in rats. *Endocrinology*, 151:569-575, 2010.
272. Brown CM, Fletcher PJ, Coscina DV. Neuropeptide Y-induced operant responding for sucrose is not mediated by dopamine. *Peptides* 19:1667-1673, 1998.
273. Altizer AM, Davidson TL. The effects of NPY and 5-TG on responding to cues for fats and carbohydrates. *Physiol Behav* 65:685-690, 1999.
274. Hagan MM, Rushing PA, Benoit SC, Woods SC, Seeley RJ. Opioid receptor involvement in the effect of AgRP- (83-132) on food intake and food selection. *Am J Physiol Regul Integr Comp Physiol* 280:R814-R821, 2001.
275. Tracy AL, Clegg DJ, Johnson JD, Davidson TL, Benoit SC. Effects of AgRP (83-132) on appetitive responding for fat and carbohydrate. *Pharmacol Biochem Behav*, under review.
276. Wetzler S, Dumaz V, Goubern M, Tome D, Larue-Achagiotis C. Intraperitoneal leptin modifies macronutrient choice in self-selecting rats. *Physiol Behav* 83:65-72, 2004.
277. Davidson TL. Learning about deprivation intensity stimuli. *Behav Neurosci* 101:198-208, 1987.
278. Davidson TL, Flynn FW, Jarrard LE. Potency of food deprivation intensity cues as discriminative stimuli. *J Exp Psychol Anim Behav Process* 18:174-181, 1992.
279. Davidson TL, Kanoski SE, Tracy AL, Walls EK, Clegg D, Benoit SC. The interoceptive cue properties of ghrelin generalize to food deprivation. *Peptides* 26:1602-1610, 2005.
280. Davidson TL, Carretta JC. Cholecystokinin, but not bombesin, has interoceptive sensory consequences like 1-h food deprivation. *Physiol Behav* 53:737-745, 1993.
281. Kanoski SE, Walls EK, Davidson TL. Interoceptive "satiety" signals produced by leptin and CCK. *Peptides* 28:988-1002, 2007.
282. Seeley RJ, Benoit SC, Davidson TL. Discriminative cues produced by NPY do not generalize to the interoceptive cues produced by food deprivation. *Physiol Behav* 58:1237-1241, 1995.
283. Jewett DC, Schaal DW, Cleary J, Thompson T, Levine AS. The discriminative stimulus effects of neuropeptide Y. *Brain Res* 561:165-168, 1991.
284. Benoit SC, Tracy AL, Air EL, Kinzig K, Seeley RJ, Davidson TL. The role of the hypothalamic melanocortin system in behavioral appetitive processes. *Pharmacol Biochem Behav* 69:603-609, 2001.
285. Mattes R. Hunger ratings are not a valid proxy measure of reported food intake in humans. *Appetite* 15:103-113, 1990.

286. Stubbs RJ, Hughes DA, Johnstone AM, Rowley E, Reid C, Elia M, Stratton R, Delargy H, King N, Blundell JE. The use of visual analogue scales to assess motivation to eat in human subjects: a review of their reliability and validity with an evaluation of new hand-held computerized systems for temporal tracking of appetite ratings. *Br J Nutr* 84:405-415, 2000.
287. Oesch S, Ruegg C, Fischer B, Degen L, Beglinger C. Effect of gastric distension prior to eating on food intake and feelings of satiety in humans. *Physiol Behav* 87:903-910, 2006.
288. Brennan IM, Feltrin KL, Horowitz M, Smout AJ, Meyer JH, Wishart J, Feinle-Bisset C. Evaluation of interactions between CCK and GLP-1 in their effects on appetite, energy intake, and antropyloroduodenal motility in healthy men. *Am J Physiol Regul Integr Comp Physiol* 288:R1477-R1485, 2005.
289. Lieveverse RJ, Jansen JB, Masclee AM, Lamers CB. Satiety effects of cholecystokinin in humans. *Gastroenterology* 106:1451-1454, 1994.
290. Gutzwiller JP, Goke B, Drewe J, Hildebrand P, Ketterer S, Handschin D, Winterhalder R, Conen D, Beglinger C. Glucagon-like peptide-1: a potent regulator of food intake in humans. *Gut* 44:81-86, 1999.
291. Batterham RL, Bloom SR. The gut hormone peptide YY regulates appetite. *Ann NY Acad Sci* 994:162-168, 2003.
292. Beglinger C, Degen L, Matzinger D, D'Amato M, Drewe J. Loxiglumide, a CCK-A receptor antagonist, stimulates calorie intake and hunger feelings in humans. *Am J Physiol Regul Integr Comp Physiol* 280:R1149-R1154, 2001.
293. Giraudo SQ, Grace MK, Billington CJ, Levine AS. Differential effects of neuropeptide Y and the mu-agonist DAMGO on 'palatability' vs. 'energy'. *Brain Res* 834:160-163, 1999.
294. Berridge KC. The debate over dopamine's role in reward: the case for incentive salience. *Psychopharmacology* 191:391-431, 2007.
295. Berridge KC. 'Liking' and 'wanting' food rewards: brain substrates and roles in eating disorders. *Physiol Behav* 97:537-550, 2009.
296. Kelley AE, Baldo BA, Pratt WE, Will MJ. Corticostriatal-hypothalamic circuitry and food motivation: integration of energy, action and reward. *Physiol Behav* 86:773-795, 2005.
297. Naleid AM, Grace MK, Cummings DE, Levine AS. Ghrelin induces feeding in the mesolimbic reward pathway between the ventral tegmental area and the nucleus accumbens. *Peptides* 26:2274-2279, 2005.
298. Figlewicz DP, Naleid AM, Sipols AJ. Modulation of food reward by adiposity signals. *Physiol Behav* 91:473-478, 2007.
299. Skibicka KP, Shirazi RH, Rabasa-Papio C, Alvarez-Crespo M, Neuber C, Vogel H, Dickson SL. Divergent circuitry underlying food reward and intake effects of ghrelin: dopaminergic VTA-accumbens projection mediates ghrelin's effect on food reward but not food intake. *Neuropharmacology* 73C:274-283, 2013.
300. Dossat AM, Lilly N, Kay K, Williams DL. Glucagon-like peptide 1 receptors in nucleus accumbens affect food intake. *J Neurosci* 31:14453-14457, 2011.
301. Romero-Pico A, Novelle MG, Folgueira C, Lopez M, Nogueiras R, Dieguez C. Central manipulation of dopamine receptors attenuates the orexigenic action of ghrelin. *Psychopharmacology* doi: 10.1007/s00213-013-3096-7, 2013.
302. Overduin J, Figlewicz DP, Bennett-Jay J, Kittleson S, Cummings DE. Ghrelin increases the motivation to eat, but does not alter food palatability. *Am J Regul Integr Comp Physiol* 303:R259-R269, 2012.
303. Thorpe AJ, Cleary JP, Levine AS, Kotz CM. Centrally administered orexin A increases motivation for sweet pellets in rats. *Psychopharmacology* 182:75-83, 2005.
304. Ward SJ, Dykstra LA. The role of CB1 receptors in sweet versus fat reinforcement: effect of CB1 receptor deletion, CB1 receptor antagonism (SR141716A) and CB1 receptor agonism (CP-55940). *Behav Pharmacol* 16:381-388, 2005.
305. Figlewicz DP, Bennett JL, Naleid AM, Davis C, Grimm JW. Intraventricular insulin and leptin decrease sucrose self-administration in rats. *Physiol Behav* 89:611-616, 2006.
306. Dickson SL, Shirazi RH, Hansson C, Bergquist F, Nissbrandt H, Skibicka KP. The glucagon-like peptide 1 (GLP-1) analogue, exendin-4, decreases the rewarding value of food: a new role for mesolimbic GLP-1 receptors. *J Neurosci* 32: 4812-20.

307. Carr KA, Lin H, Fletcher KD, Epstein LH. Food reinforcement, dietary disinhibition and weight gain in nonobese adults. *Obesity* doi: 10.1002/oby.20392, 2013.
308. Meyers KP, Sclafani A. Development of learned flavor preferences. *Dev Psychobiol* 48:380-388, 2006.
309. Higgs S. Memory and its role in appetite regulation. *Physiol Behav* 85:67-72, 2005.
310. Kanoski SE, Davidson TL. Western diet consumption and cognitive impairment: links to hippocampal dysfunction and obesity. *Physiol Behav* 103:59-68, 2011.
311. Farr SA, Banks WA, Morley JE. Effects of leptin on memory processing. *Peptides* 27:1420-1425, 2006.
312. Harvey J. Leptin regulation of neuronal excitability and cognitive function. *Curr Opin Pharmacol* 7:643-647, 2007.
313. Diano S, Farr SA, Benoit SC, McNay EC, da Silva I, Horvath B, Gaskin FS, Nonaka N, Jaeger LB, Banks WA, Morley JE, Pinto S, Sherwin RS, Xu L, Yamada KA, Sleeman MW, Tschop MH, Horvath TL. Ghrelin controls hippocampal spine synapse density and memory performance. *Nat Neurosci* 9:381-388, 2006.
314. Morrison CD. Leptin signaling in brain: a link between nutrition and cognition. *Biochem Biophys Acta* 1792:401-408, 2009.
315. Polonsky KS. Dynamics of insulin secretion in obesity and diabetes. *Int J Obes Relat Metab Disord* 24:S29-S31, 2000.
316. Woods SC, D'Alessio DA, Tso P, Rushing PA, Clegg DJ, Benoit SC, Gotoh K, Liu M, Seeley RJ. Consumption of a high-fat diet alters the homeostatic regulation of energy balance. *Physiol Behav* 83:573-578, 2004.
317. Maffei M, Halaas J, Ravussin E, Pratley RE, Lee GH, Zhang Y, Fei H, Kim S, Lallone R, Ranganathan S, Kern PA, Friedman JM. Leptin levels in human and rodent: measurement of plasma leptin and ob mRNA in obese and weight-reduced subjects. *Nat Med* 1:1155-1161, 1995.
318. Tschop M, Weyer C, Tataranni PA, Devanarayan V, Ravussin E, Heiman ML. Circulating ghrelin levels are decreased in human obesity. *Diabetes* 50:707-709, 2001.
319. Perreault M, Istrate N, Wang L, Nichols AJ, Tozzo E, Stricker-Krongrad A. Resistance to the orexigenic effect of ghrelin in dietary-induced obesity in mice: reversal upon weight loss. *Int J Obes Relat Metab Disord* 28:879-885, 2004.
320. Beck B, Max JP, Fernet B, Richy S. Adaptation of ghrelin levels to limit body weight gain in the obese Zucker rat. *Biochem Biophys Res Commun* 318:846-851, 2004.
321. Williams DL, Grill HJ, Cummings DE, Kaplan JM. Overfeeding-induced weight gain suppresses plasma ghrelin levels in rats. *J Endocrinol Invest* 29:863-868, 2006.
322. Hyland NP, Pittman QJ, Sharkey KA. Peptide YY containing enteroendocrine cells and peripheral tissue sensitivity to PYY and PYY(3-36) are maintained in diet-induced obese and diet-resistant rats. *Peptides* 28:1185-1190, 2007.
323. Wang H, Storlien LH, Huang XF. Effects of dietary fat types on body fatness, leptin, and ARC leptin receptor, NPY, and AgRP mRNA expression. *Am J Physiol Endocrinol Metab* 282:E1352-E1359, 2002.
324. Lin S, Storlien LH, Huang XF. Leptin receptor, NPY, POMC mRNA expression in the diet-induced obese mouse brain. *Brain Res* 875:89-95, 2000.
325. Nam SY, Kratzsch J, Kim KW, Kim KR, Lim SK, Marcus C. Cerebrospinal fluid and plasma concentrations of leptin, NPY, and alpha-MSH in obese women and their relationship to negative energy balance. *J Clin Endocrinol Metab* 86:4849-4853, 2001.
326. Widdowson PS, Upton R, Buckingham R, Arch J, Williams G. Inhibition of food response to intracerebroventricular injection of leptin is attenuated in rats with diet-induced obesity. *Diabetes* 46:1782-1785, 1997.
327. Halaas JL, Boozer C, Blair-West J, Fidanhusein N, Denton DA, Friedman JM. Physiological responses to long-term peripheral and central leptin infusion in lean and obese mice. *Proc Natl Acad Sci USA* 94:8878-8883, 1997.
328. Levin BE, Dunn-Meynell AA. Reduced central leptin sensitivity in rats with diet-induced obesity. *Am J Physiol Regul Integr Comp Physiol* 283:R941-R948, 2002.
329. Heymsfield SB, Greenberg AS, Fujioka K, Dixon RM, Kushner R, Hunt T, Lubina JA, Patane J, Self B, Hunt P, McCamish M. Recombinant leptin for weight loss in obese and lean adults: a randomized, controlled, dose-escalation trial. *JAMA* 282:1568-1575, 2000.

330. Zelissen PM, Stenlof K, Lean ME, Fogteloo J, Keulen ET, Wilding J, Finer N, Rossner S, Lawrence E, Fletcher C, McCamish M. Effect of three treatment schedules of recombinant methionyl human leptin on body weight in obese adults: a randomized, placebo-controlled trial. *Diabetes Obes Metab* 7:755-761, 2005.
331. Clegg DJ, Benoit SC, Reed JA, Woods SC, Dunn-Meynell A, Levin BE. Reduced anorexic effects of insulin in obesity-prone rats fed a moderate-fat diet. *Am J Physiol Regul Integr Comp Physiol* 288:R981-R986, 2004.
332. Banks WA, DiPalma CR, Farrell CL. Impaired transport of leptin across the blood-brain barrier in obesity. *Peptides* 20:1341-1345, 1999.
333. Banks WA, Coon AB, Robinson SM, Moinuddin A, Shultz JM, Nakaoke R, Morley JE. Triglycerides induce leptin resistance at the blood-brain barrier. *Diabetes* 53:1253-1260, 2004.
334. Petersen KF, Shulman GI. Etiology of insulin resistance. *Am J Med* 119:S10-S16, 2006.
335. Munzberg H, Myers MG Jr. Molecular and anatomical determinants of central leptin resistance. *Nat Neurosci* 8:566-570, 2005.
336. Enriori PJ, Evans AE, Sinnayah P, Jobst EE, Tonelli-Lomos L, Billes SK, Glavas MM, Grayson BE, Perello M, Nillni EA, Grove KL, Cowley MA. Diet-induced obesity causes severe but reversible leptin resistance in arcuate melanocortin neurons. *Cell Metab* 5:181-194, 2007.
337. Enriori PJ, Evans AE, Sinnayah P, Cowley MA. Leptin resistance and obesity. *Obesity* 14:254S-258S, 2006.
338. Sahu A. Resistance to the satiety action of leptin following chronic central leptin infusion is associated with the development of leptin resistance in neuropeptide Y neurons. *J Neuroendocrinol* 14:796-804, 2002.
339. Morris DL, Rui L. Recent advances in understanding leptin signaling and leptin resistance. *Am J Physiol Endocrinol Metab* 297:E1247-E1259, 2009.
340. Pfluger PT, Kampe J, Castaneda TR, Vahl T, D'Alessio DA, Kruthaupt T, Benoit SC, Cuntz U, Rochlitz HJ, Moehlig M, Pfeiffer AF, Koebnick C, Weickert MO, Otto B, Spranger J, Tschop MH. Effect of human body weight changes on circulating levels of peptide YY and peptide Y3-36. *J Clin Endocrinol Metab* 92:583-588, 2007.
341. le Roux CW, Batterham RL, Aylwin SJ, Patterson M, Borg CM, Wynne KJ, Kent A, Vincent RP, Gardiner J, Ghatei MA, Bloom SR. Attenuated peptide YY release in obese subjects is associated with reduced satiety. *Endocrinology* 147:3-8, 2006.
342. Wisen O, Bjorvell H, Cantor P, Johansson C, Theodorsson E. Plasma concentrations of regulatory peptides in obesity following modified sham feeding (MSF) and a liquid test meal. *Regul Pept* 39:43-54, 1992.
343. Tso P, Liu M. Apolipoprotein A-IV, food intake, and obesity. *Physiol Behav* 83:631-643, 2004.
344. Briggs DI, Enriori PJ, Lemus MB, Cowley MA, Andrews ZB. Diet-induced obesity causes ghrelin resistance in arcuate NPY/AgRP neurons. *Endocrinology* 151:4745-4755, 2010.
345. Williams DL, Hyvarinen N, Lilly N, Kay K, Dossat A, Parise E, Torregrossa AM. Maintenance on a high-fat diet impairs the anorexic response to glucagon-like-peptide-1 receptor activation. *Physiol Behav* 103: 557-564, 2011.
346. Davis JF, Tracy AL, Schurdak JD, Tschop MH, Lipton JW, Clegg DJ, Benoit SC. Exposure to elevated levels of dietary fat attenuates psychostimulant reward and mesolimbic dopamine turnover in the rat. *Behav Neurosci* 122:1257-1263, 2008.
347. Wang GJ, Volkow ND, Logan J, Pappas NR, Wong CT, Zhu W, Netusil N, Fowler JS. Brain dopamine and obesity. *Lancet* 357:354-357, 2001.
348. Wang GJ, Volkow ND, Fowler JS. The role of dopamine in motivation for food in humans: implications for obesity. *Expert Opin Ther Targets* 6:601-609, 2002.
349. Stice E, Spoor S, Ng J, Zald DH. Relation of obesity to consummatory and anticipatory food reward. *Physiol Behav* 97:551-560, 2009.
350. Molteni R, Barnard RJ, Ying Z, Roberts CK, Gomez-Pinilla F. A high-fat, refined sugar diet reduces hippocampal brain-derived neurotrophic factor, neuronal plasticity and learning. *Neuroscience* 112:803-814, 2002.
351. Hwang LL, Wang CH, Li TL, Chang SD, Lin LC, Chen CP, Chen CT, Liang KC, Ho IK, Yang WS, Chiou LC. Sex differences in high-fat diet-induced obesity, metabolic alterations and learning, and synaptic plasticity deficits in mice. *Obesity* 18:463-469, 2010.

352. Lindqvist A, Mohapel P, Bouter B, Frielingsdorf H, Pizzo D, Brundin P, Erlanson-Albertsson C. High-fat diet impairs hippocampal neurogenesis in male rats. *Eur J Neurol* 13:1385-1388, 2006.
353. Kanoski SE, Zhang Y, Zheng W, Davidson TL. The effects of a high-energy diet on hippocampal function and blood-brain barrier integrity in the rat. *J Alzheimers Dis* 21:207-219, 2010.
354. Winocur G, Greenwood CE. Studies of the effects of high fat diets on cognitive function in a rat model. *Neurobiol Aging* 26:46-49, 2005.
355. Kanoski SE, Davidson TL. Different patterns of memory impairments accompany short- and long-term maintenance on a high-energy diet. *J Exp Psychol Anim Behav Process* 36:313-319, 2010.
356. Scarpace PJ, Zhang Y. Elevated leptin: consequence or cause of obesity? *Front Biosci* 12:3531-3544, 2007.
357. Yokum S, Ng J, Stice E. Attentional bias to food images associated with elevated weight and future weight gain: an fMRI study. *Obesity* 19:1775-1783, 2011.
358. Stice E, Yokum S, Burger KS. Elevated reward region responsivity predicts future substance use onset, but not overweight/obesity onset. *Biol Psychiatry* 73:869-876, 2013.
359. Levin BE, Dunn-Meynell AA, Balkan B, Keeseey RE. Selective breeding for diet-induced obesity and resistance in Sprague-Dawley rats. *Am J Physiol* 273:R725-R730, 1997.
360. Irani BG, Dunn-Meynell AA, Levin BE. Altered hypothalamic leptin, insulin, and melanocortin binding associated with moderate-fat diet and predisposition to obesity. *Endocrinology* 148:310-316, 2007.
361. Levin BE, Dunn-Meynell AA. Dysregulation of arcuate nucleus preproneuropeptide Y mRNA in diet-induced obese rats. *Am J Physiol* 272:R1365-1370, 1997.
362. Levin BE. Arcuate NPY neurons and energy homeostasis in diet-induced obese and resistant rats. *Am J Physiol* 276:R382-R387, 1999.
363. Levin BE, Dunn-Meynell AA, Banks WA. Obesity-prone rats have normal blood-brain barrier transport but defective central leptin signaling before obesity onset. *Am J Physiol Regul Integr Comp Physiol* 286:R143-R150, 2004.
364. Levin BE, Magnan C, Migrenne S, Chua SC Jr, Dunn-Meynell AA. F-DIO obesity-prone rat is insulin resistant before obesity onset. *Am J Physiol Regul Integr Comp Physiol* 289:R704-R711, 2005.