

# VON HIPPEL-LINDAU DISEASE

**Sven Gläsker, MD**, Professor, Department of Neurosurgery, VUB University Hospital Brussels, Belgium. Sven.glaesker@uzbrussel.be

**Hartmut P.H. Neumann, MD**, Professor, Department of Internal Medicine, Freiburg University Medical Center, Germany. hartmut.neumann@uniklinik-freiburg.de

**Christian A. Koch, MD, FACP, MACE**, Professor, Department of Internal Medicine, University of Louisville, KY; Oldenburg University and Medicover MVZ, Germany. Christian.koch65@gmail.com

**Alexander Vortmeyer, MD**, Professor and Director, Neuropathology Training Program, Department of Pathology, Indiana University School of Medicine, Indianapolis, IN. avortmey@iu.edu

**Updated September 12, 2018**

## ABSTRACT

Von Hippel-Lindau (VHL) disease is an autosomal dominantly inherited tumor syndrome. The incidence of VHL disease is about one in 36,000 livebirths and the penetrance is higher than 90%. Similar to other tumor suppressor gene disorders, VHL disease is characterized by frequent development of specific types of tumors in selective organs. The disease is named after the German ophthalmologist Eugen von Hippel, who identified and described characteristic retinal manifestations, and the Swedish pathologist Arvid Lindau, who discovered the frequent co-occurrence of retinal and cerebellar hemangioblastoma with tumors and cysts in visceral organs. He described the clinical spectrum of VHL disease in detail in a large series of cases from Sweden but also from other European countries. A clinical classification system divides individuals who are affected by VHL disease into two groups: Those predominantly without pheochromocytoma are classified as VHL type 1, and those predominantly with pheochromocytoma are classified as VHL type 2. VHL type 2 is further subdivided into type 2A (with renal cancer) and type 2B (without renal cancer). In type 2C affected patients develop solely pheochromocytomas. Tumorigenesis in patients with VHL disease shares fundamental principles in the different affected organ systems: germline VHL inactivation leads to persistence of microscopic developmentally arrested structures. These microscopic cell clusters already reveal biallelic VHL inactivation and consequent up-regulation of hypoxia inducible factor (HIF) and down-stream targets like VEGF, EPO. Current research for pharmacotherapy of VHL disease targets proteins of the HIF cascade and compounds that lead to upregulation and re-functionalization of mutated VHL protein. For complete coverage of all related areas of Endocrinology, please visit our on-line FREE web-text, [WWW.ENDOTEXT.ORG](http://WWW.ENDOTEXT.ORG).

## INTRODUCTION

Von Hippel-Lindau (VHL) disease is an autosomal dominantly inherited tumor syndrome. The incidence of VHL disease is about one in 36,000 livebirths [1, 2] and the penetrance is higher than 90 % [3]. Similar to other tumor suppressor gene disorders, VHL disease is characterized by frequent development of specific types of tumors in selective organs. The disease is named after the German ophthalmologist Eugen von Hippel, who identified and described characteristic retinal manifestations [4], and the Swedish pathologist Arvid Lindau, who discovered the frequent co-occurrence of retinal and cerebellar hemangioblastoma with tumors and cysts in visceral organs. He described the clinical spectrum of VHL disease in detail in a large series of cases from Sweden but also from other European countries [5, 6].

A clinical classification system divides individuals who are affected by VHL disease into two groups [7]: Those predominantly without pheochromocytoma are classified as VHL type 1, and those predominantly with pheochromocytoma are classified as VHL type 2. VHL type 2 is further subdivided into type 2A (with renal cancer) and type 2B (without renal cancer). In type 2C affected patients develop solely pheochromocytomas [8].

## **VON HIPPEL-LINDAU DISEASE MANIFESTATIONS**

VHL disease targets a highly selective subset of organs by the frequent development of specific types of heavily vascularized tumors. Multiple and bilateral tumors occur frequently. Affected organs and tumors are listed in table 1 [9-12].

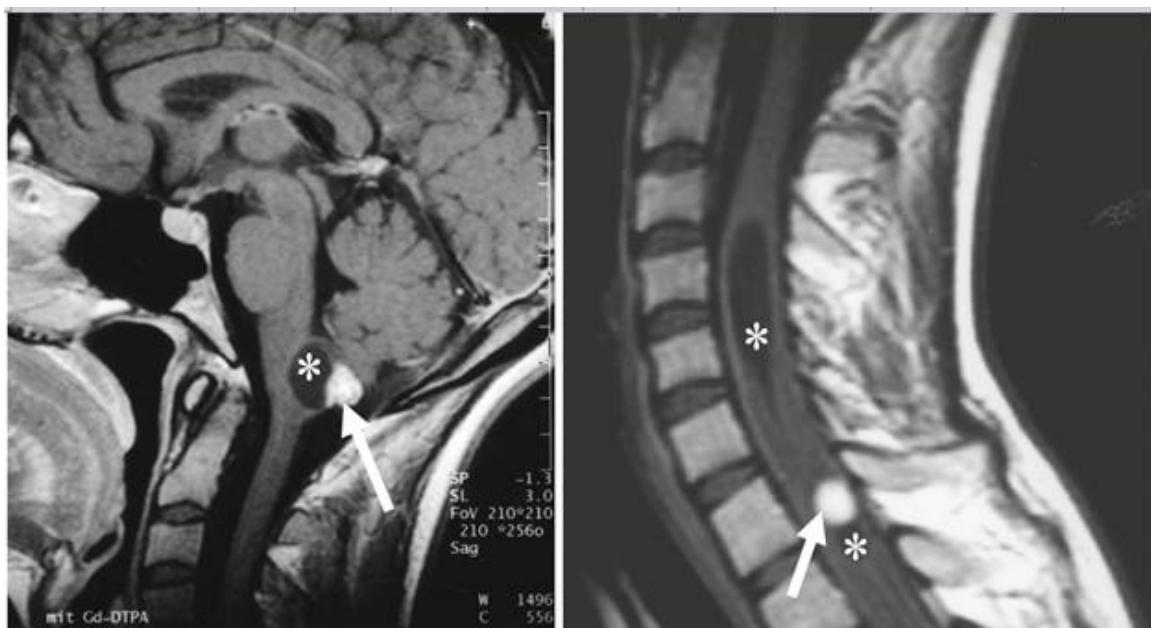
<b>Table 1: Manifestations of VHL disease [9-12]</b>		
<b>Tumor</b>	<b>Frequency</b>	<b>Mean age of onset</b>
Retinal hemangioblastomas	60%	25y
Cerebellar and spinal hemangioblastomas	65%	33y
Endolymphatic sac tumors	10%	22y
Renal clear cell carcinomas and cysts	45%	39y
Pheochromocytomas	20%	30y
Pancreatic cysts, microcystic serous adenomas, neuroendocrine tumors	35-70%	36y
Epididymal and broad ligament cystadenomas	> 50% of male	unknown

## **CNS HEMANGIOBLASTOMAS IN VHL SYNDROME**

Hemangioblastomas are the index tumors of VHL disease and are frequently the earliest manifestation of the disease. Multiple hemangioblastomas occur frequently in patients with VHL disease. In one major study 127 of 160 patients with VHL-associated hemangioblastomas had multiple tumors [13]. The central nervous system hemangioblastoma burden in VHL disease is associated with partial germline deletions and male sex [14]. The benign slow growing highly vascularized tumors cause neurologic symptoms depending on their size and location [13, 14]. A spinal

hemangioblastoma may lead to focal neurologic deficits such as weakness and paresthesias. Posterior fossa tumors may present with ataxia, dysmetria, nystagmus, and slurred speech. Symptoms may also evolve due to CSF obstruction causing signs of increased intracranial pressure or brain stem herniation [3, 11, 15]. Symptoms are usually not caused by the tumor itself but rather by an associated pseudocyst or syrinx, which is caused by vascular leakage from the tumor [16] and which is usually larger than the tumor itself [13]. Polyglobulia has been reported in patients with hemangioblastomas [17]. It occurs in 10% of patients [18] and may cause thrombosis. Removal of the largest hemangioblastoma usually resolves polyglobulia [18].

Diagnosis is established by contrast enhanced MRI of the head and spine [19, 20], which typically identifies a solid enhancing nodule associated with a pseudocyst or syrinx (figure 1).



**Figure 1: On contrast-enhanced MRI, hemangioblastoma is identified as solid enhancing nodule; the tumor is typically associated with extra-tumoral cyst (left) or syrinx (right).**

Patients with VHL disease frequently develop several CNS hemangioblastomas but not all of the tumors need to be removed. Surgery is needed for all symptomatic tumors and for all tumors causing CSF obstruction [13, 21, 22]. Once symptoms occur, they can usually not be reversed by surgery; therefore, surgery may be recommended in a subset of asymptomatic tumors with radiographic progression [22]. Complete surgical removal of the solid tumor is the treatment of choice. If surgery is not possible, radiation therapy may be an alternative. [23-27]. There is currently no established chemotherapy for hemangioblastomas. Substances studied include different tyrosine kinase inhibitors such as semaxanib [28-32], Vatalanib (ongoing trial), Sunitinib [33-36] and Dovitinib [37] as well as pazopanib [38-40].

Surgical resection of hemangioblastomas requires careful dissection in the plane between tumor and CNS tissue. If the tumor is entered, major bleeding will occur. Hemangioblastoma patients harbor a risk for spontaneous or perioperative hemorrhage [41] and preoperative embolization has been recommended for large solid tumors [42, 43]. The tumors can usually be completely removed and permanent new neurologic deficit due to surgery is rare in experienced hands. Intraoperative ultrasound allows for identification of small tumors and control of complete resection of the tumors [44]. Recently intraoperative ICG angiography has been used by different groups. Some of them find it helpful in understanding the vascular anatomy of the lesions, whereas others find it of limited value [45-50]

Hemangioblastomas are composed of a dense capillary network with “stromal” cells in between. Microdissection and genetic deletion analysis revealed the “stromal” cells to represent the neoplastic component of these tumors [51], while the capillary network predominantly represents reactive VEGF-driven angiogenesis. The histogenesis of the neoplastic “stromal” cells has been controversial. Tumor cells stain consistently positive for neuron specific enolase [52-54], neural cell adhesion molecule [55, 56] and vimentin [54-58]. The “stromal” cells appears to represent developmentally arrested VHL-deficient hemangioblast progenitor cells with hematopoietic differentiation potential [59-63], based on the observation of expression of pre-hemangioblastic markers like scl and brachyury [63-66] and the ability of primitive blood island formation and differentiation into early red blood cells [65, 67-69]. Furthermore, anatomic studies on spinal cord and cerebellum of VHL patients revealed numerous developmentally arrested structural elements [70] that serve as potential precursor material for hemangioblastic tumors [71]. In addition to hematopoietic differentiation capacity the tumor cells are also capable of primitive vasculogenesis[72, 73]. It has remained unresolved, however, whether the tumor cells are of mesodermal or neuroectodermal origin.

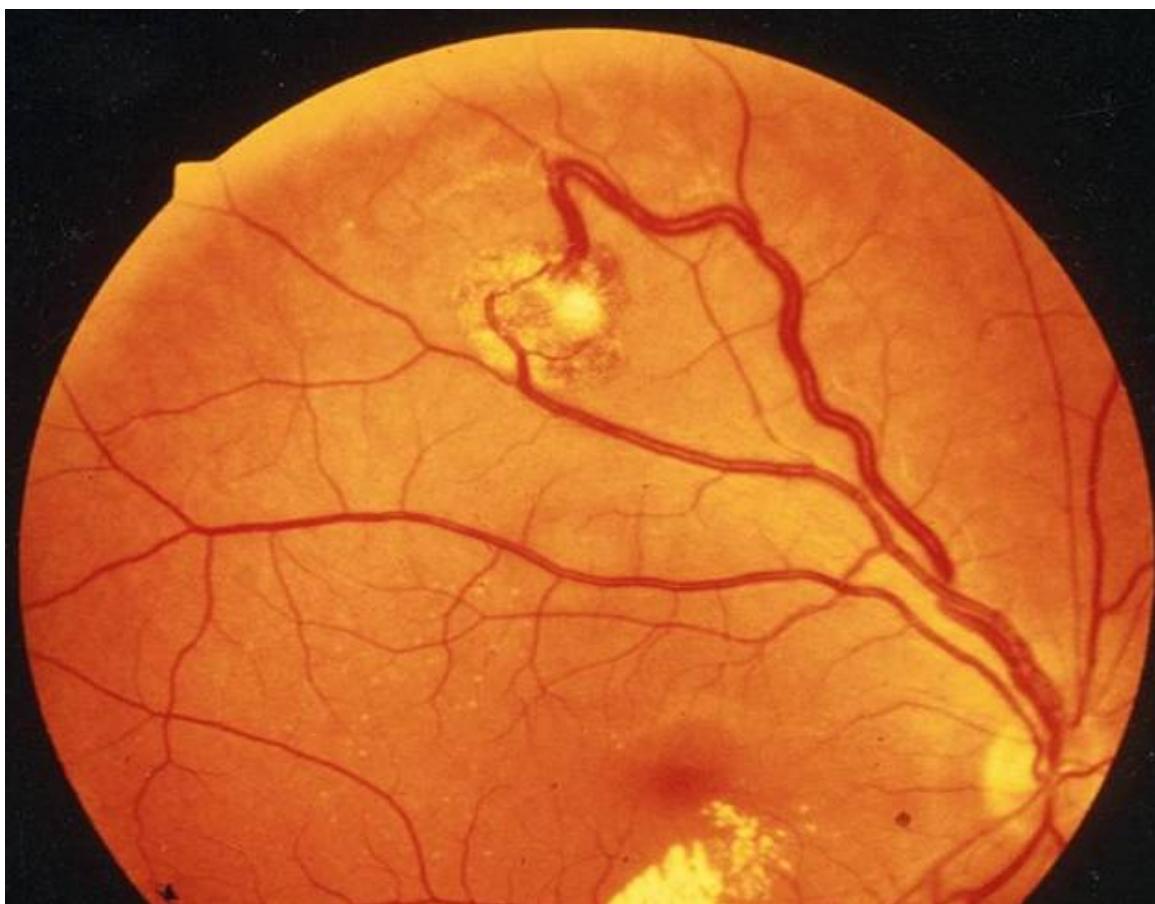
## **RETINAL HEMANGIOBLASTOMAS IN VHL SYNDROME**

Retinal hemangioblastomas are benign tumors that can occur as a sporadic entity as well as in patients with VHL disease. Tumors frequently occur in multiplicity and bilaterally (in about 50 %). Histologically, retinal hemangioblastomas are identical to CNS hemangioblastomas [74]. High expression of VEGF in these tumors causes vascularization, vascular leakage and exudation, and eventually retinal detachment in the eye.

Annual fundoscopic screening and early treatment of asymptomatic lesions is recommended for VHL patients starting at one year of age. Most peripheral retinal hemangioblastomas are well controlled by laser photocoagulation or cryotherapy [75, 76]. Vitrectomy may be performed in larger tumors [77]. Tumors in the optic disc should be monitored without treatment. Alternative treatments are in experimental stages and

include VEGF inhibition [30, 78, 79] and radiation therapy [80]. Recently propranolol has been suggested for treatment of retinal hemangioblastomas [81].

With screening and early treatment of detected lesions, visual prognosis for VHL patients is good: In one major study approximately 8% of the eyes of VHL patients had poor visual acuity of 20/200 or worse with approx. 8% of these eyes requiring enucleation [82]. Genotyping of young patients presenting with paraganglioma or pheochromocytoma can lead to early detection of retinal hemangioblastomas and other VHL manifestations in family members [83].



**Figure 2: Fundoscopic photograph illustrating a retinal hemangioblastoma (arrows).**

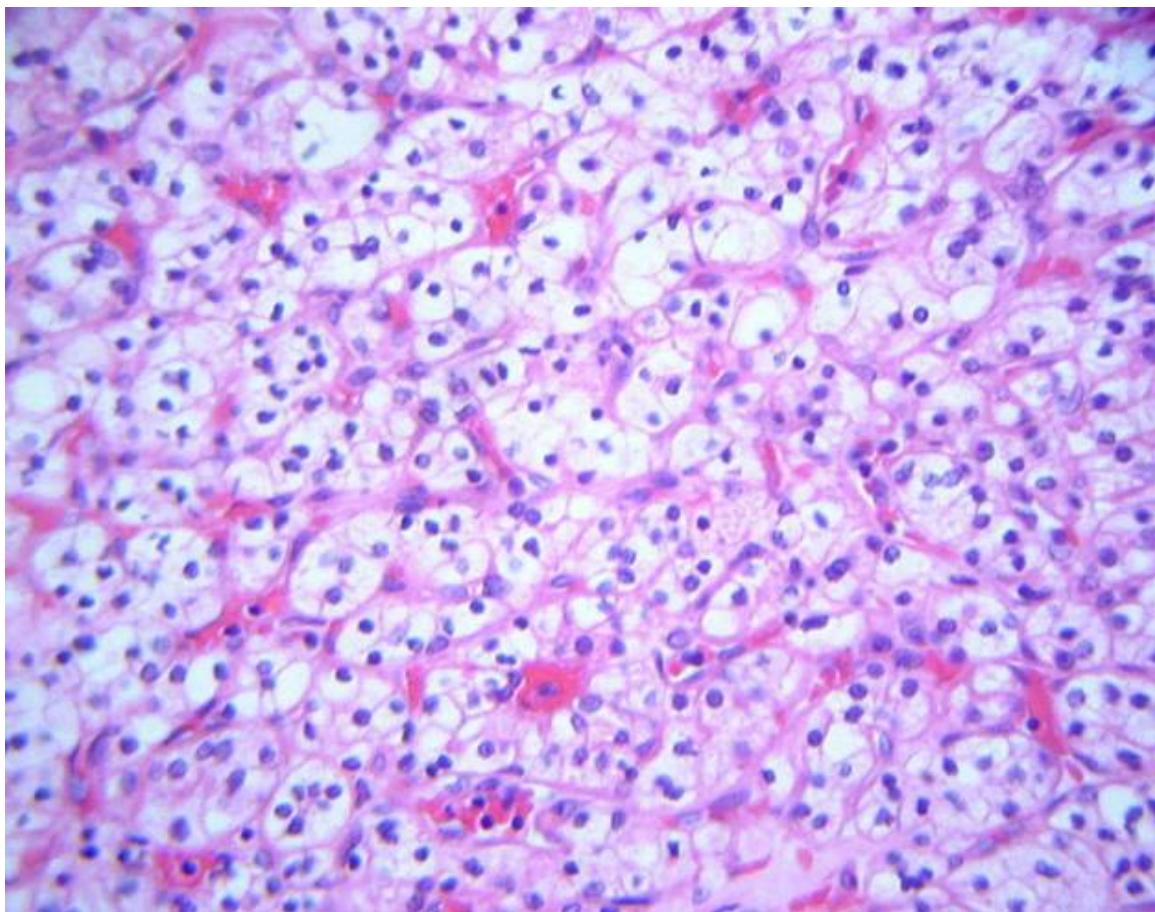
## RENAL MANIFESTATIONS IN VHL SYNDROME

Patients with VHL disease can develop renal cysts and renal cell carcinomas [84-89]. Renal cell carcinomas are malignant tumors. The pathogenetic relationship between renal cysts and renal cell carcinoma has remained unclear. One study suggests rare transition from a cyst to a solid lesion [90]. Histologically, renal cell carcinomas in VHL disease are always of clear-cell type (figure 3), and small carcinomas tend to be low

grade [91]. While small renal tumors in VHL disease tend to be low grade and minimally invasive [91], their rate of growth varies widely [92, 93].

Renal manifestations are usually multiple and bilateral in patients with VHL disease. They remain asymptomatic for a long time. Advanced cases of renal cell carcinoma may present with hematuria, flank pain, or a flank mass.

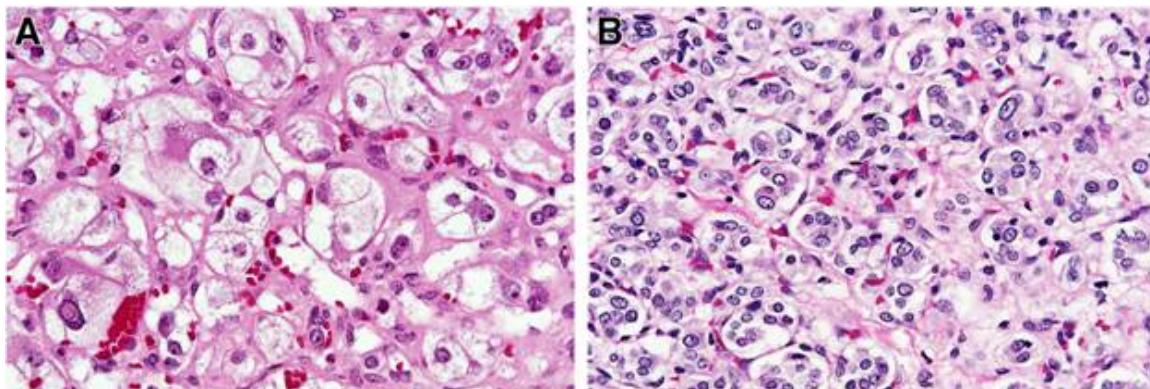
Renal cysts in VHL disease usually do not require treatment. Therapy of renal cell carcinoma is targeted towards prevention of metastasis and preservation of kidney function. In contrast to sporadic renal cell carcinoma, nephrectomy is not recommended as primary therapy for VHL-disease-associated renal cell carcinomas. Most specialized centers recommend nephron-sparing surgery for carcinomas that exceed 3 cm in size [85, 94]. Recent work supports the hypothesis, that a diameter of 4 cm is also acceptable [95]. Although the gold standard for treating renal tumors are open and laparoscopic partial nephrectomy, alternative therapies including cryotherapy and radiofrequency ablation are presently utilized [96]. Possible substances for treatment of metastasized disease include pazopanib [97].



**Figure 3. VHL disease associated renal cell carcinomas are of clear cell type (H&E stain).**

## PHEOCHROMOCYTOMAS IN VHL SYNDROME

Pheochromocytomas usually are sporadic but in more than 33% of patients can occur in a familial syndrome such as the VHL syndrome [98, 99]. Presence of pheochromocytomas defines VHL disease type 2 (A,B,C) (Figure 4, modified from [100]).



**Figure 4. VHL-associated pheochromocytoma**

In a series of 246 patients with VHL syndrome, 64 patients (26%) were found to have a pheochromocytoma [101]. In one third, these tumors were “nonfunctional” and did not cause symptoms of catecholamine excess such as hypertension. Utilizing 18F-DOPA functional imaging in 52 patients with von Hippel-Lindau disease, 4 of 10 patients with positive adrenal uptake had elevated catecholamine levels, i.e. 6 of 10 pheochromocytomas were “silent” [102]. Asymptomatic patients with pheochromocytoma have also been reported in 15 of 150 patients seen at the Mayo Clinic [103] and in 19 of 33 patients with adrenal pheochromocytomas that were incidentally discovered [104]. The mean age at diagnosis of VHL patients reported by Walther et al. 1999 [101] was 29 years with a range from age 6 to age 54 years. Bilateral pheochromocytomas in this group were found in 39% of patients. Extra-adrenal pheochromocytomas or paragangliomas in patients with VHL disease can be sympathetic or parasympathetic and, if located in the head and neck, are mostly unable to produce and secrete catecholamines [99, 105-108]. Genotyping of young patients presenting with a functional paraganglioma can lead to detection of other family members with VHL disease [83]. Recent studies in Chinese and Asian Indian patients revealed novel genotype-phenotype correlations in VHL disease [109-111]. Loss of the ability of VHL protein to degrade HIF under hypoxic conditions can promote the development of bilateral pheochromocytomas [112].

In general, extra-adrenal pheochromocytomas have a higher frequency and potential of malignancy than pheochromocytomas located in the adrenal gland [113]. Malignant pheochromocytomas (defined as the presence of chromaffin tissue at locations where such tissue should not be present, i.e. lungs, liver, bone, lymph nodes) are uncommon in patients with VHL syndrome [92, 99, 100, 114, 115]. However, even in children as young

as 12 years of age metastatic pheochromocytomas in VHL disease have been observed [116]. Unfortunately, there are no markers yet that can reliably distinguish a benign from a malignant pheochromocytoma [117].

The detection of a pheochromocytoma in patients with VHL syndrome is particularly important, given the possible need for surgical interventions for other tumors such as CNS hemangioblastomas. Just like in any patient with an undetected pheochromocytoma, surgery and other factors can lead to life-threatening hypertensive attacks intraoperatively. During pregnancy, surgery usually should take place in the second trimester and may include interval adrenalectomy [118]. Screening for pheochromocytoma in at high risk patients (VHL type 2A,B,C) should include measurements of urinary catecholamines and fractionated metanephrenes, as well as of plasma free metanephrenes, acknowledging that normetanephrine is the predominant biochemical marker for adrenal and extra-adrenal VHL-associated pheochromocytomas, and that methoxytyramine a helpful marker for head and neck paragangliomas [119, 120]. Determining the screening age for pheochromocytoma should likely be made dependent on potential beneficial and negative sequelae that could occur if screening were not performed, i.e. malignancy (lower than 5% in VHL disease!), hypertension, radiation exposure from frequent imaging. There is marked intrafamilial variation, and the penetrance of pheochromocytoma may be as late as age 54 years [101, 121]. In a recent study of 37 Danish patients from the national VHL research database and additional international children and adolescents with VHL disease the age range of clinical manifestations (mostly retinal and CNS hemangioblastomas) was from 6 to 17 years of age [122].

At present, experts recommend pheochromocytoma screening be started already in childhood at the age of 8 years [99, 123] [9]. The adrenal glands (suprarenal) can be visualized while undergoing an MRI to look for renal abnormalities in patients with VHL disease. More than 70% of pheochromocytomas presenting in children are due to VHL disease. The diagnosis can further be established by abdominal computed tomography and/or magnetic resonance imaging [9].

Given the low frequency rate of extra-adrenal pheochromocytomas (approx. 15%) and malignant pheochromocytomas (< 5%) in patients with VHL disease, only selected patients should undergo <sup>123</sup>I-MIBG scanning to search for extra-adrenal or metastatic lesions, before operation [113, 124]. Importantly, (isolated) pheochromocytoma can be the presenting manifestation of VHL syndrome [3, 125]. Approximately 50% of patients with apparently isolated familial pheochromocytoma have VHL disease and approx. 10% of patients with apparently sporadic pheochromocytoma [99, 126, 127], acknowledging that a family history is not always present or reliably obtainable [128] and that even if VHL germline mutations were detected by certified laboratory testing in patients with non-syndromic pheochromocytoma (i.e. in 3 of 182 patients, [129], misdiagnoses can occur.

Treatment of nonmetastatic adrenal pheochromocytoma consists of adrenal sparing (partial) tumor removal, either uni- or bilateral, depending on whether one or both adrenal glands are affected [130-132]. Endoscopic adrenal sparing removal of the pheochromocytoma should be performed in most if not all patients. [133]. Cortical sparing adrenal surgery exhibits < 5% recurrence rates after 10 years of follow-up while preserving normal glucocorticoid function in more than 50% of patients [134]. The perioperative outcome of patients with VHL disease is very good, although they may have a very high noradrenergic secretion compared to patients with neurofibromatosis type 1 and multiple endocrine neoplasia type 2A [135]. Occasional VHL patients with pheochromocytomas exhibit a decline in tumor size after 19 months of sunitinib therapy [136].

Whether nonfunctional pheochromocytomas (clinically only remarkable as adrenal masses on imaging with normal fractionated urinary metanephrenes and/or normal plasma free metanephrenes) require treatment/surgery remains unclear. Important aspects in this context are the questions how fast such tumors grow, if and when they cause symptoms, and if and when (at what size) they metastasize [137, 138].

## **EPIDIDYMAL CYSTADENOMAS IN VHL SYNDROME**

These non-malignant tumors occur in about half of all male VHL patients [139]. Epididymal cystadenomas consist of cystic and adenomatous areas and arise typically in the caput epididymidis (figure 5). The tumors have been suggested to arise from microscopic precursor structures, which are abundantly observed in the efferent ductiles of the caput epididymidis of VHL patients [140, 141]. Epididymal cystadenomas rarely cause symptoms and have no potential for malignant transformation. In rare cases bilateral obstruction of efferent ductiles and spermatic cords may occur resulting in sterility. Epididymal cystadenomas are well visible on ultrasound.

Women who are affected by VHL disease develop cystadenomas of the broad ligaments, which play no major clinical role.



**Figure 5. VHL disease associated epididymal cystadenomas consistently reveal papillary (pap), tubular (tu) and cystic (cy) architecture [140].**

#### PANCREATIC MANIFESTATIONS IN VHL SYNDROME

Pancreatic manifestations include pancreatic neuroendocrine tumors, cysts and cystadenomas. Overall, 35–70% of VHL patients develop a pancreatic manifestation [142–144]. Distinguishing between a benign microcystic adenoma and a pancreatic neuroendocrine tumor can be difficult. Similar to hemangioblastomas, pancreatic neuroendocrine tumors reveal a staggering growth pattern [145].

Most pancreatic tumors in VHL disease are asymptomatic. Extremely rare – and depending on size and location – are bile duct obstruction and/or pancreatic insufficiency. Most patients do not need any treatment. This is important for patients with pancreatic cysts and/or microcystic adenomas. In a recent study of 48 patients with VHL disease, branch duct intraductal papillary mucinous neoplasms were present in 10% [146].

(68)Ga-DOTATOC PET/CT appears to be a very sensitive screening tool in detecting pancreatic neuroendocrine tumors in VHL disease [147]. All patients should be investigated by MRI with intravenous contrast administration. It is very important that imaging is performed in the early arterial phase. In contrast to the multicystic

appearance of microcystic adenoma, neuroendocrine pancreatic tumors present as solid lesions and need to be considered for surgical removal. Mutations in exon 3, especially of codons 161/167 are at enhanced risk for metastatic PanNETs [148]. However, most VHL associated pancreatic neuroendocrine tumors are small and slowly growing. In general, VHL associated pancreatic neuroendocrine tumors have a favorable prognosis compared to sporadic tumors [149]. Tumors with a diameter over 2.8 cm should be treated surgically to avoid metastasis according to a recent multicenter study [148]. Resection of pancreatic tumors may also be performed if the patient is undergoing laparotomy for other lesions [150, 151]. In cases of biliary obstruction and/or pancreatic insufficiency treatment consists of placing biliary stents and/or replacing pancreatic enzymes. A new option is organ-sparing endoscopic resection which can be applied to tumors in the tail and the body of the pancreas [152]. Chemotherapy is currently under investigation. Occasional patients respond well to somatostatin analog therapy [153].

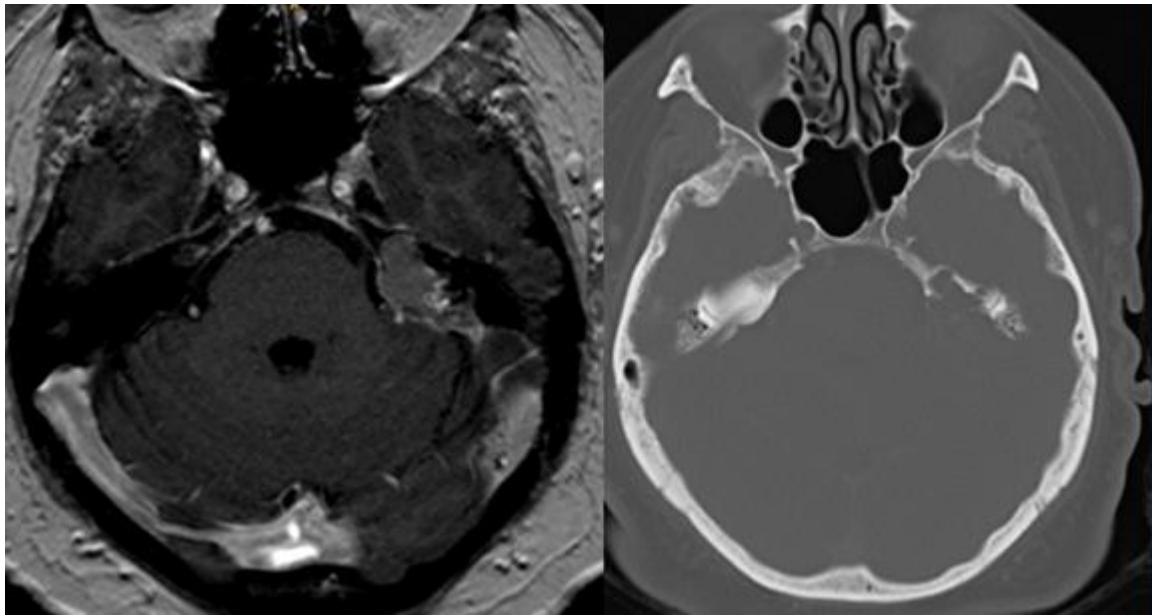
## **ENDOLYMPHATIC SAC TUMORS IN VHL SYNDROME**

Endolymphatic sac tumors are benign cystadenomas that arise in the vestibular aqueduct and may extend into the extraosseous portion of the endolymphatic sac, into the petrous bone, and into the inner ear [154, 155]. Large tumors have been reported to extend into the brain [50]. The vestibular aqueduct connects the inner ear with the extraosseous portion of the endolymphatic sac, a dural fold at the posterior surface of the petrous bone. Despite their benign nature these lesions grow locally invasive into the temporal bone due to their origin in the osseous portion of the vestibular aqueduct.

Patients may present with hypakusis or hearing loss (100%), tinnitus (77%), disequilibrium (62%), and facial paresis (8%) [156]. Hearing loss and vestibulopathy may occur suddenly due to tumor-associated intralabyrinthine hemorrhage, or insidiously, consistent with endolymphatic hydrops [157].

Diagnosis is established by MRI, which shows homogeneous or variable patterns of patchy enhancement and CT (bone window) revealing widening of the osseous portion of the vestibular aqueduct (figure 6).

Radical resection allows for complete excision of the tumors. The preoperative level of hearing can usually be preserved. The timing of surgery depends on the severity of symptoms, the slow but variable growth of the tumors, the possibility of injury to the 7<sup>th</sup> and 8<sup>th</sup> cranial nerves and possible bilateral occurrence. Hearing preserving early surgical resection is recommended to enhance the opportunity for complete resection and decrease irreversible audiovestibular symptoms [47]. Endolymphatic hydrops can be detected early on contrast-enhanced delayed FLAIR MRI even before the tumoral mass itself is visible [45]. The role of radiation in the treatment of these tumors remains controversial (see [50] for a review).



**Figure 6. VHL-associated ELST (endolymphatic sac tumor). MRI reveals an inhomogenously enhancing mass in the left temporal bone, CT shows osteolysis.**

## PROGNOSIS IN VHL SYNDROME

Before the introduction of clinical screening programs and timely prophylactic treatment of VHL associated lesions, the life expectancy of affected patients used to be impaired with a median survival around 50 years [2, 11]. Nowadays, life expectancy may be significantly improved by clinical screening programs with early detection and treatment of VHL manifestations [158]. Modern management of VHL has meanwhile achieved a life expectancy of additional 10 years [159].

## VHL SCREENING

All patients with hemangioblastoma, the index tumor of VHL disease, should receive a careful family history, screening for detection of other lesions associated with VHL (table 2), and should undergo genetic testing for VHL disease [160, 161].

Once the diagnosis of VHL disease is established, patients should undergo an annual screening program in order to identify manifestations early before deficits or metastasis occur. Screening includes periodic contrast enhanced MRI scans. CNS gadolinium accumulation will eventually occur in virtually all VHL patients, however, no pathological effect of this accumulation is known so far [47]. The screening programs vary slightly between different centers. An example of a routine surveillance protocol for von Hippel–Lindau disease (modified from Maher [162]) is presented in table 2:

<b>Table 2: Screening Procedures</b>
--------------------------------------

<b>Screen for retinal hemangioblastoma:</b> Annual ophthalmic examinations (direct and indirect ophthalmoscopy), beginning at age 1 (from birth on).
<b>Screen for CNS hemangioblastoma:</b> Contrast enhanced MRI brain and full spine, beginning at age 12. Annual or biennial depending on manifestations.
<b>Screen for renal cell carcinoma and pancreatic tumors:</b> MRI (or ultrasound) examinations of the abdomen every 12 months, beginning from the age of 12 years.
<b>Screen for pheochromocytoma:</b> Annual blood pressure monitoring and 24-h urine studies for catecholamine metabolites starting at age 6. Alternatively measuring plasma metanephrenes. MRI starting at age 12.
<b>Screen for epididymal cystadenoma:</b> Ultrasonography of the testes starting at 18 years.
<b>Screen for ELST with audiogram:</b> Biennial audiogram starting at age 16
Additional investigations in response to symptoms or signs of specific complications.

## MOLECULAR ASPECTS IN VHL SYNDROME

Patients with VHL disease carry a germline mutation of the VHL tumor suppressor gene (VHL). The VHL gene had originally been linked to chromosome 3p25 in 1988 [163] and was subsequently cloned in 1993 [164]. The evolutionarily conserved VHL gene encodes two protein products, a 30-kDa full-length form (p30) and a 19-kDa form (p19). The protein product of the VHL gene (pVHL) interacts with elongins B, C and Cullin-2 to form the VBC complex, an E3 ubiquitin ligase [165]. This multiprotein complex targets proteins for ubiquitin-mediated degradation, analogous to the *S. cerevisiae* SCF (Skp1/Cdc53/F-box) ubiquitination machinery [166-168]. Major targets of the VBC complex include the regulatory subunits of the heterodimeric transcription factor, hypoxia inducible factor (HIF). Therefore, VHL inactivation causes an up-regulation of HIF and downstream proteins under normoxic conditions. Many of the HIF targets are involved in the adaptation to acute or chronic hypoxia and include genes involved in the uptake and metabolism of glucose (GLUT-1), angiogenesis (VEGF, PDGF), control of extracellular pH (CA9), mitogenesis (TGF $\alpha$ ), and erythropoiesis (erythropoietin) [169-172].

In addition to HIF degradation, the VHL protein is involved in HIF-independent cellular processes. It directs the proper deposition of fibronectin and collagen IV within the extracellular matrix [173]. Furthermore, it works to stabilize microtubules and foster the maintenance of primary cilium. The VHL protein also promotes the stabilization and activation of p53 and, in neuronal cells, promotes apoptosis by downregulation of Jun-B [173]. It was furthermore shown that acute VHL inactivation causes a senescent-like phenotype independently of HIF but dependent on the retinoblastoma protein (Rb) and the SWI2/SNF2 chromatin remodeler p400 [174].

Biallelic inactivation of the VHL tumor suppressor gene is suspected as initiating step of tumorigenesis. In general, two genetic events or “hits” are thought to occur to the two alleles of a tumor suppressor gene: The first hit is represented by a germline mutation and the second hit is characterized by inactivation of the wild-type allele [175]. More

recent work suggests, that the biallelic VHL inactivation is only a first step in VHL tumorigenesis.

Tumorigenesis in VHL disease shares fundamental principles in the different affected organ systems: Germline VHL inactivation leads to persistence of microscopic developmentally arrested structures. These microscopic cell clusters already reveal biallelic VHL inactivation and consequent up-regulation of hypoxia inducible factor and down-stream targets like VEGF, EPO etc. This principle has been shown for CNS hemangioblastomas [61, 62], renal cell carcinomas [176], endolymphatic sac tumors [154] and epididymal cystadenomas [140] and is most likely valid for all VHL associated tumors.

Current research for pharmacotherapy of VHL disease targets proteins of the HIF cascade [33]. Additionally investigated are compounds that lead to upregulation and re-functionalization of mutated VHL protein [177].

## REFERENCES

1. Maher, E.R., L. Iselius, J.R. Yates, M. Littler, C. Benjamin, R. Harris, J. Sampson, A. Williams, M.A. Ferguson-Smith, and N. Morton, Von Hippel-Lindau disease: a genetic study. *J Med Genet*, 1991. 28(7): p. 443-7.
2. Neumann, H.P. and O.D. Wiestler, Clustering of features and genetics of von Hippel-Lindau syndrome [letter; comment]. *Lancet*, 1991. 338(8761): p. 258.
3. Maher, E.R., J.R. Yates, R. Harries, C. Benjamin, R. Harris, A.T. Moore, and M.A. Ferguson-Smith, Clinical features and natural history of von Hippel-Lindau disease [see comments]. *Q J Med*, 1990. 77(283): p. 1151-63.
4. von Hippel, E., Ueber eine seltene Erkrankung der Netzhaut. *Graefes Arch Ophthalmol.*, 1904. 59: p. 83-106.
5. Lindau, A., Studien über Kleinhirnzysten. Bau, Pathogenese und Beziehung zur Angiomatosis retinae. *Acta Path et Microbiol Scand*, 1926. Suppl 1: p. 1-126.
6. Lindau, A., Zur Frage der Angiomatosis retinae und ihrer Hirnkomplikation. *Acta Ophthalmol (Copenh)*, 1927. 4: p. 193-226.
7. Zbar, B., T. Kishida, F. Chen, L. Schmidt, E.R. Maher, F.M. Richards, P.A. Crossey, A.R. Webster, N.A. Affara, M.A. Ferguson-Smith, H. Brauch, D. Glavac, H.P. Neumann, S. Tisherman, J.J. Mulvihill, D.J. Gross, T. Shuin, J. Whaley, B. Seizinger, N. Kley, S. Olschwang, C. Boisson, S. Richard, C.H. Lips, M. Lerman, and et al., Germline mutations in the Von Hippel-Lindau disease (VHL) gene in families from North America, Europe, and Japan. *Hum Mutat*, 1996. 8(4): p. 348-57.
8. DeLellis, R.A., P.U. Heitz, R.V. Lloyd, and C. Eng, eds. *Pathology and molecular genetics of endocrine tumours. WHO classification of tumours of endocrine organs*. 2003, IARC press: Lyon.

9. Lonser, R.R., G.M. Glenn, M. Walther, E.Y. Chew, S.K. Libutti, W.M. Linehan, and E.H. Oldfield, von Hippel-Lindau disease. *Lancet*, 2003. 361(9374): p. 2059-67.
10. Maher, E.R. and W.G. Kaelin, Jr., von Hippel-Lindau disease. *Medicine (Baltimore)*, 1997. 76(6): p. 381-91.
11. Lamiell, J.M., F.G. Salazar, and Y.E. Hsia, von Hippel-Lindau disease affecting 43 members of a single kindred. *Medicine (Baltimore)*, 1989. 68(1): p. 1-29.
12. Friedrich, C.A., Von Hippel-Lindau syndrome. A pleomorphic condition. *Cancer*, 1999. 86(11 Suppl): p. 2478-82.
13. Wanebo, J., R. Lonser, G. Glenn, and E. Oldfield, The natural history of hemangioblastomas of the central nervous system in patients with von Hippel-Lindau disease. *J Neurosurg*, 2003. 98(1): p. 82-94.
14. Lonser, R.R., J.A. Butman, K. Huntoon, A.R. Asthagiri, T. Wu, K.D. Bakhtian, E.Y. Chew, Z. Zhuang, W.M. Linehan, and E.H. Oldfield, Prospective natural history study of central nervous system hemangioblastomas in von Hippel-Lindau disease. *J Neurosurg*, 2014. 120(5): p. 1055-62.
15. Neumann, H.P., Prognosis of von Hippel-Lindau syndrome. *Vasa*, 1987. 16(4): p. 309-11.
16. Lonser, R.R., A.O. Vortmeyer, J.A. Butman, S. Glasker, M.A. Finn, J.M. Ammerman, M.J. Merrill, N.A. Edwards, Z. Zhuang, and E.H. Oldfield, Edema is a precursor to central nervous system peritumoral cyst formation. *Ann Neurol*, 2005. 58(3): p. 392-9.
17. Carpenter, G., H. Schwartz, and A. Walker, Neurogenic polycythemia. *Ann Intern Med*, 1943. 19(3): p. 470-81.
18. Glasker, S., M.T. Kruger, J.H. Klingler, M. Wlodarski, J. Klompen, B. Schatlo, B. Hippchen, H.P. Neumann, and V. Van Velthoven, Hemangioblastomas and neurogenic polyglobulia. *Neurosurgery*, 2013. 72(6): p. 930-5; discussion 935.
19. Resche, F., J.P. Moisan, J. Mantoura, A. de Kersaint-Gilly, M.J. Andre, I. Perrin-Resche, D. Menegalli-Boggelli, Y. Lajat, and S. Richard, Haemangioblastoma, haemangioblastomatosis, and von Hippel-Lindau disease. *Adv Tech Stand Neurosurg*, 1993. 20: p. 197-304.
20. Filling-Katz, M.R., P.L. Choyke, E. Oldfield, L. Charnas, N.J. Patronas, G.M. Glenn, M.B. Gorin, J.K. Morgan, W.M. Linehan, B.R. Seizinger, and et al., Central nervous system involvement in Von Hippel-Lindau disease. *Neurology*, 1991. 41(1): p. 41-6.
21. Weil, R., R. Lonser, H. DeVroom, J. Wanebo, and E. Oldfield, Surgical management of brainstem hemangioblastomas in patients with von Hippel-Lindau disease. *J Neurosurg*, 2003. 98(1): p. 95-105.
22. van Velthoven, V., P.C. Reinacher, J. Klisch, H.P. Neumann, and S. Glasker, Treatment of intramedullary hemangioblastomas, with special attention to von Hippel-Lindau disease. *Neurosurgery*, 2003. 53(6): p. 1306-13.
23. Chang, S.D., J.A. Meisel, S.L. Hancock, D.P. Martin, M. McManus, and J.R. Adler, Jr., Treatment of hemangioblastomas in von Hippel-Lindau disease with

- linear accelerator-based radiosurgery. *Neurosurgery*, 1998. 43(1): p. 28-34; discussion 34-5.
24. Patrice, S.J., P.K. Sneed, J.C. Flickinger, D.C. Shrieve, B.E. Pollock, E. Alexander, 3rd, D.A. Larson, D.S. Kondziolka, P.H. Gutin, W.M. Wara, M.W. McDermott, L.D. Lunsford, and J.S. Loeffler, Radiosurgery for hemangioblastoma: results of a multiinstitutional experience. *Int J Radiat Oncol Biol Phys*, 1996. 35(3): p. 493-9.
25. Niemela, M., Y.J. Lim, M. Soderman, J. Jaaskelainen, and C. Lindquist, Gamma knife radiosurgery in 11 hemangioblastomas. *J Neurosurg*, 1996. 85(4): p. 591-6.
26. Pan, L., E.M. Wang, B.J. Wang, L.F. Zhou, N. Zhang, P.W. Cai, and J.Z. Da, Gamma knife radiosurgery for hemangioblastomas. *Stereotact Funct Neurosurg*, 1998. 70 Suppl 1: p. 179-86.
27. Jawahar, A., D. Kondziolka, Y.I. Garces, J.C. Flickinger, B.E. Pollock, and L.D. Lunsford, Stereotactic radiosurgery for hemangioblastomas of the brain. *Acta Neurochirurgica*, 2000. 142(6): p. 641-4.
28. Madhusudan, S., G. Deplanque, J.P. Braybrooke, E. Cattell, M. Taylor, P. Price, M.D. Tsaloumas, N. Moore, S.M. Huson, C. Adams, P. Frith, P. Scigalla, and A.L. Harris, Antiangiogenic therapy for von Hippel-Lindau disease. *JAMA*, 2004. 291(8): p. 943-4.
29. Richard, S., L. Croisille, J. Yvert, N. Casadeval, P. Eschwege, N. Aghakhani, P. David, A. Gaudric, P. Scigalla, and O. Hermine, Paradoxical secondary polycythemia in von Hippel-Lindau patients treated with anti-vascular endothelial growth factor receptor therapy. *Blood*, 2002. 99(10): p. 3851-3.
30. Aiello, L.P., D.J. George, M.T. Cahill, J.S. Wong, J. Cavallerano, A.L. Hannah, and W.G. Kaelin, Jr., Rapid and durable recovery of visual function in a patient with von hippel-lindau syndrome after systemic therapy with vascular endothelial growth factor receptor inhibitor su5416. *Ophthalmology*, 2002. 109(9): p. 1745-51.
31. Girmens, J.F., A. Erginay, P. Massin, P. Scigalla, A. Gaudric, and S. Richard, Treatment of von Hippel-Lindau retinal hemangioblastoma by the vascular endothelial growth factor receptor inhibitor SU5416 is more effective for associated macular edema than for hemangioblastomas. *Am J Ophthalmol*, 2003. 136(1): p. 194-6.
32. Schuch, G., M. de Wit, J. Holtje, E. Laack, G. Schilling, D.K. Hossfeld, W. Fiedler, P. Scigalla, and M.S. Jacobs, Case 2. Hemangioblastomas: diagnosis of von Hippel-Lindau disease and antiangiogenic treatment with SU5416. *J Clin Oncol*, 2005. 23(15): p. 3624-6.
33. Jonasch, E., I.E. McCutcheon, S.G. Waguespack, S. Wen, D.W. Davis, L.A. Smith, N.M. Tannir, D.S. Gombos, G.N. Fuller, and S.F. Matin, Pilot trial of sunitinib therapy in patients with von Hippel-Lindau disease. *Ann Oncol*, 2011. 22(12): p. 2661-6.
34. Knickelbein, J.E., N. Jacobs-El, W.T. Wong, H.E. Wiley, C.A. Cukras, C.B. Meyerle, and E.Y. Chew, Systemic Sunitinib Malate Treatment for Advanced

- Juxtapapillary Retinal Hemangioblastomas Associated with von Hippel-Lindau Disease. *Ophthalmol Retina*, 2017. 1(3): p. 181-187.
35. Reyes-Botero, G., J. Gallego Perez-Larraya, and M. Sanson, Sporadic CNS hemangioblastomatosis, response to sunitinib and secondary polycythemia. *J Neurooncol*, 2012. 107(2): p. 439-40.
36. Roma, A., M. Maruzzo, U. Basso, A. Brunello, R. Zamarchi, E. Bezzon, F. Pomerri, S. Zovato, G. Opocher, and V. Zagonel, First-Line sunitinib in patients with renal cell carcinoma (RCC) in von Hippel-Lindau (VHL) disease: clinical outcome and patterns of radiological response. *Fam Cancer*, 2015. 14(2): p. 309-16.
37. Pilie, P., E. Hasanov, S.F. Matin, A.H.H. Woodson, V.D. Marcott, S. Bird, R.S. Slack, G.N. Fuller, I.E. McCutcheon, and E. Jonasch, Pilot study of dovitinib in patients with von Hippel-Lindau disease. *Oncotarget*, 2018. 9(34): p. 23390-23395.
38. Kim, B.Y., E. Jonasch, and I.E. McCutcheon, Pazopanib therapy for cerebellar hemangioblastomas in von Hippel-Lindau disease: case report. *Target Oncol*, 2012. 7(2): p. 145-9.
39. Migliorini, D., S. Haller, D. Merkler, A. Pugliesi-Rinaldi, A. Koka, K. Schaller, B. Leemann, and P.Y. Dietrich, Recurrent multiple CNS hemangioblastomas with VHL disease treated with pazopanib: a case report and literature review. *CNS Oncol*, 2015. 4(6): p. 387-92.
40. Taylor, D.G., A. Ilyas, G.U. Mehta, C.J. Chen, D. Schiff, E.H. Oldfield, and A.R. Asthagiri, Variable response of CNS hemangioblastomas to Pazopanib in a single patient with von Hippel-Lindau disease: Case report. *J Clin Neurosci*, 2018. 50: p. 154-156.
41. Glasker, S. and V. Van Velthoven, Risk of hemorrhage in hemangioblastomas of the central nervous system. *Neurosurgery*, 2005. 57(1): p. 71-6; discussion 71-6.
42. Seong Eom, K., D. Won Kim, S. Sung Choi, K. Ha Choi, and T. Young Kim, Preoperative embolization of a cerebellar haemangioblastoma using Onyx: case report and literature review. *Neurol Neurochir Pol*, 2011. 45(3): p. 292-6.
43. Glasker, S., J.H. Klingler, C. Wurtenberger, K. Muller, C. Hader, J. Zentner, H.P. Neumann, and V. Van Velthoven, Essentials and pitfalls in treatment of hemangioblastoma and von Hippel-Lindau disease. *Central European Neurosurgery*, 2010. 71(2): p. 80-7.
44. Glasker, S., M.J. Shah, B. Hippchen, H.P. Neumann, and V. Van Velthoven, Doppler-Sonographically Guided Resection of CNS Hemangioblastomas. *Neurosurgery*, 2011. 68: p. ons267-ons275.
45. Takeshima, Y., Y. Tanaka, Y. Hironaka, Y. Shida, and H. Nakase, Visualization of vascular structure of spinal hemangioblastoma using intraoperative indocyanine green videoangiography and temporary feeder occlusion. *Eur Spine J*, 2015.

46. Benedetto, N., F. Aquila, and R. Vannozzi, Use of near-infrared indocyanine videoangiography and Flow 800 in the resectioning of a spinal cord haemangioblastoma. *Br J Neurosurg*, 2013. 27(6): p. 847-9.
47. Hojo, M., Y. Arakawa, T. Funaki, K. Yoshida, T. Kikuchi, Y. Takagi, Y. Araki, A. Ishii, T. Kunieda, J.C. Takahashi, and S. Miyamoto, Usefulness of tumor blood flow imaging by intraoperative indocyanine green videoangiography in hemangioblastoma surgery. *World Neurosurg*, 2014. 82(3-4): p. e495-501.
48. Tamura, Y., Y. Hirota, S. Miyata, Y. Yamada, A. Tucker, and T. Kuroiwa, The use of intraoperative near-infrared indocyanine green videoangiography in the microscopic resection of hemangioblastomas. *Acta Neurochir (Wien)*, 2012. 154(8): p. 1407-12; discussion 1412.
49. Hwang, S.W., A.M. Malek, R. Schapiro, and J.K. Wu, Intraoperative use of indocyanine green fluorescence videography for resection of a spinal cord hemangioblastoma. *Neurosurgery*, 2010. 67(3 Suppl Operative): p. ons300-3; discussion ons303.
50. Hao, S., D. Li, G. Ma, J. Yang, and G. Wang, Application of intraoperative indocyanine green videoangiography for resection of spinal cord hemangioblastoma: advantages and limitations. *J Clin Neurosci*, 2013. 20(9): p. 1269-75.
51. Vortmeyer, A.O., J.R. Gnarra, M.R. Emmert-Buck, D. Katz, W.M. Linehan, E.H. Oldfield, and Z. Zhuang, von Hippel-Lindau gene deletion detected in the stromal cell component of a cerebellar hemangioblastoma associated with von Hippel-Lindau disease. *Hum Pathol*, 1997. 28(5): p. 540-3.
52. Becker, I., W. Paulus, and W. Roggendorf, Histogenesis of stromal cells in cerebellar hemangioblastomas. *Am J Pathol*, 1989. 134: p. 271-5.
53. Lach, B., A. Gregor, P. Rippstein, and A. Omulecka, Angiogenic histogenesis of stromal cells in hemangioblastoma: ultrastructural and immunohistochemical study. *Ultrastruct Pathol*, 1999. 23(5): p. 299-310.
54. Omulecka, A., B. Lach, J. Alwasia, and A. Gregor, Immunohistochemical and ultrastructural studies of stromal cells in hemangioblastoma. *Folia Neuropathol*, 1995. 33(1): p. 41-50.
55. Ishizawa, K., T. Komori, and T. Hirose, Stromal cells in hemangioblastoma: Neuroectodermal differentiation and morphological similarities to ependymoma. *Pathol Int*, 2005. 55(7): p. 377-85.
56. Bohling, T., K.H. Plate, M. Haltia, K. Alitalo, and H.P.H. Neumann, Von Hippel-Lindau disease and capillary hemangioblastoma, in *Pathology and genetics of tumours of the central nervous system*, P. Kleihues and W.K. Cawenee, Editors. 2000, World Health Organization: Lyon. p. 223-226.
57. Cruz-Sanchez, F.F., M.L. Rossi, S. Rodriguez-Prados, N. Nakamura, J.T. Hughes, and H.B. Coakham, Haemangioblastoma: histological and immunohistological study of an enigmatic cerebellar tumour. *Histol Histopathol*, 1990. 5(4): p. 407-13.

58. Ding, X.H., L.F. Zhou, Y.Z. Tan, Y. Zhao, and J.J. Zhu, Histologic and histogenetic investigations of intracranial hemangioblastomas. *Surg Neurol*, 2007. 67(3): p. 239-45; discussion 245.
59. Shively, S.B., E.A. Falke, J. Li, M.G. Tran, E.R. Thompson, P.H. Maxwell, E. Roessler, E.H. Oldfield, R.R. Lonser, and A.O. Vortmeyer, Developmentally arrested structures preceding cerebellar tumors in von Hippel-Lindau disease. *Mod Pathol*, 2011. 24(8): p. 1023-30.
60. Vortmeyer, A.O., M. Tran, W. Zeng, S. Glasker, C. Riley, M. Tsokos, B. Ikejiri, M.J. Merrill, M. Raffeld, Z. Zhuang, R.R. Lonser, P.H. Maxwell, and E.H. Oldfield, Evolution of VHL tumorigenesis in nerve root tissue. *J Pathol*, 2006. 210(3): p. 374-82.
61. Vortmeyer, A.O., Q. Yuan, Y.S. Lee, Z. Zhuang, and E.H. Oldfield, Developmental effects of von Hippel-Lindau gene deficiency. *Annals of Neurology*, 2004. 55(5): p. 721-8.
62. Vortmeyer, A.O., S. Frank, S.Y. Jeong, K. Yuan, B. Ikejiri, Y.S. Lee, D. Bhowmick, R.R. Lonser, R. Smith, G. Rodgers, E.H. Oldfield, and Z. Zhuang, Developmental arrest of angioblastic lineage initiates tumorigenesis in von Hippel-Lindau disease. *Cancer Research*, 2003. 63(21): p. 7051-5.
63. Glasker, S., J. Li, J.B. Xia, H. Okamoto, W. Zeng, R.R. Lonser, Z. Zhuang, E.H. Oldfield, and A.O. Vortmeyer, Hemangioblastomas share protein expression with embryonal hemangioblast progenitor cell. *Cancer Research*, 2006. 66(8): p. 4167-4172.
64. Barresi, V., E. Vitarelli, G. Branca, M. Antonelli, F. Giangaspero, and G. Barresi, Expression of Brachyury in Hemangioblastoma: Potential Use in Differential Diagnosis. *Am J Surg Pathol*, 2012.
65. Park, D.M., Z. Zhuang, L. Chen, N. Szerlip, I. Maric, J. Li, T. Sohn, S.H. Kim, I.A. Lubensky, A.O. Vortmeyer, G.P. Rodgers, E.H. Oldfield, and R.R. Lonser, von Hippel-Lindau Disease-Associated Hemangioblastomas Are Derived from Embryologic Multipotent Cells. *PLoS Med*, 2007. 4(2): p. e60.
66. Barresi, V., A. Ieni, G. Branca, and G. Tuccari, Brachyury: a diagnostic marker for the differential diagnosis of chordoma and hemangioblastoma versus neoplastic histological mimickers. *Dis Markers*, 2014. 2014: p. 514753.
67. Zec, N., P. Cera, and J. Towfighi, Extramedullary hematopoiesis in cerebellar hemangioblastoma. *Neurosurgery*, 1991. 29(1): p. 34-7.
68. Stein, A.A., A.O. Schilp, and R.D. Whitfield, The histogenesis of hemangioblastoma of the brain. A review of twenty-one cases. *J Neurosurg*, 1960. 17: p. 751-61.
69. Vortmeyer, A.O., S. Frank, S.Y. Jeong, K. Yuan, B. Ikejiri, Y.S. Lee, D. Bhowmick, R.R. Lonser, R. Smith, G. Rodgers, E.H. Oldfield, and Z. Zhuang, Developmental arrest of angioblastic lineage initiates tumorigenesis in von Hippel-Lindau disease. *Cancer Res*, 2003. 63(21): p. 7051-5.
70. Vortmeyer, A.O., Q. Yuan, Y.S. Lee, Z. Zhuang, and E.H. Oldfield, Developmental effects of von Hippel-Lindau gene deficiency. *Ann Neurol*, 2004. 55(5): p. 721-8.

71. Vortmeyer, A.O., M.G. Tran, W. Zeng, S. Glasker, C. Riley, M. Tsokos, B. Ikejiri, M.J. Merrill, M. Raffeld, Z. Zhuang, R.R. Lonser, P.H. Maxwell, and E.H. Oldfield, Evolution of VHL tumourigenesis in nerve root tissue. *J Pathol*, 2006. 210(3): p. 374-82.
72. Glasker, S., J. Smith, M. Raffeld, J. Li, E.H. Oldfield, and A.O. Vortmeyer, VHL-deficient vasculogenesis in hemangioblastoma. *Exp Mol Pathol*, 2014. 96(2): p. 162-7.
73. Zhuang, Z., J.M. Frerich, K. Huntoon, C. Yang, M.J. Merrill, Z. Abdullaev, S.D. Pack, S.B. Shively, G. Stamp, and R.R. Lonser, Tumor derived vasculogenesis in von Hippel-Lindau disease-associated tumors. *Sci Rep*, 2014. 4: p. 4102.
74. Chan, C.C., A.O. Vortmeyer, E.Y. Chew, W.R. Green, D.M. Matteson, D.F. Shen, W.M. Linehan, I.A. Lubensky, and Z. Zhuang, VHL gene deletion and enhanced VEGF gene expression detected in the stromal cells of retinal angioma. *Arch Ophthalmol*, 1999. 117(5): p. 625-30.
75. Singh, A.D., M. Nouri, C.L. Shields, J.A. Shields, and N. Perez, Treatment of retinal capillary hemangioma. *Ophthalmology*, 2002. 109(10): p. 1799-806.
76. Schmidt, D., E. Natt, and H.P. Neumann, Long-term results of laser treatment for retinal angiomas in von Hippel-Lindau disease. *Eur J Med Res*, 2000. 5(2): p. 47-58.
77. Gaudric, A., V. Krivacic, G. Duguid, P. Massin, S. Giraud, and S. Richard, Vitreoretinal surgery for severe retinal capillary hemangiomas in von hippel-lindau disease. *Ophthalmology*, 2011. 118(1): p. 142-9.
78. Ach, T., D. Thiemeier, A.E. Hoeh, K.B. Schaal, and S. Dithmar, Intravitreal bevacizumab for retinal capillary haemangioma: longterm results. *Acta Ophthalmol*, 2010. 88(4): p. e137-8.
79. Slim, E., J. Antoun, H.R. Kourie, A. Schakkal, and G. Cherfan, A case series and literature review. *Can J Ophthalmol*, 2014. 49(5): p. 450-7.
80. Matsuo, T., K. Himei, K. Ichimura, H. Yanai, S. Nose, T. Mimura, Y. Miyoshi, and T. Tsushima, Long-term effect of external beam radiotherapy of optic disc hemangioma in a patient with von Hippel-Lindau disease. *Acta Med Okayama*, 2011. 65(2): p. 135-41.
81. Ueba, T., H. Abe, J. Matsumoto, T. Higashi, and T. Inoue, Efficacy of indocyanine green videography and real-time evaluation by FLOW 800 in the resection of a spinal cord hemangioblastoma in a child: case report. *J Neurosurg Pediatr*, 2012. 9(4): p. 428-31.
82. Chew, E.Y., Ocular manifestations of von Hippel-Lindau disease: clinical and genetic investigations. *Trans Am Ophthalmol Soc*, 2005. 103: p. 495-511.
83. Bahougne, T., P. Romanet, A. Mohamed, K. Caselles, T. Cuny, A. Barlier, and P. Niccoli, Functioning Mediastinal Paraganglioma Associated with a Germline Mutation of von Hippel-Lindau Gene. *J Clin Med*, 2018. 7(6).
84. Lubensky, I.A., J.R. Gnarra, P. Bertheau, M.M. Walther, W.M. Linehan, and Z. Zhuang, Allelic deletions of the VHL gene detected in multiple microscopic clear cell renal lesions in von Hippel-Lindau disease patients. *Am J Pathol*, 1996. 149(6): p. 2089-94.

85. Neumann, H.P., P. Riegler, W. Huber, R. Corradini, A. Sessa, D. Fontana, U. Wetterauer, and G. Janetschek, The challenge of kidney lesions in von Hippel-Lindau disease. [Review] [39 refs]. Contributions to Nephrology, 2001. 136: p. 193-207.
86. Richard, S., D. Chauveau, Y. Chretien, C. Beigelman, A. Denys, J.P. Fendler, G. Fromont, F. Paraf, O. Helenon, S. Nizard, and et al., Renal lesions and pheochromocytoma in von Hippel-Lindau disease. Adv Nephrol Necker Hosp, 1994. 23: p. 1-27.
87. Grubb, R.L., 3rd, P.L. Choyke, P.A. Pinto, W.M. Linehan, and M.M. Walther, Management of von Hippel-Lindau-associated kidney cancer. Nat Clin Pract Urol, 2005. 2(5): p. 248-55.
88. Kaelin, W.G., Jr., The von Hippel-Lindau tumor suppressor gene and kidney cancer. Clin Cancer Res, 2004. 10(18 Pt 2): p. 6290S-5S.
89. Linehan, W.M., M.M. Walther, and B. Zbar, The genetic basis of cancer of the kidney. Journal of Urology., 2003. 170(6 Pt 1): p. 2163-72.
90. Choyke, P.L., G.M. Glenn, M.M. Walther, B. Zbar, G.H. Weiss, R.B. Alexander, W.S. Hayes, J.P. Long, K.N. Thakore, and W.M. Linehan, The natural history of renal lesions in von Hippel-Lindau disease: a serial CT study in 28 patients. American Journal of Roentgenology, 1992. 159: p. 1229-34.
91. Poston, C.D., G.S. Jaffe, I.A. Lubensky, D. Solomon, B. Zbar, W.M. Linehan, and M.M. Walther, Characterization of the renal pathology of a familial form of renal cell carcinoma associated with von Hippel-Lindau disease: clinical and molecular genetic implications. Journal of Urology, 1995. 153(1): p. 22-6.
92. Walther, M.M., P.L. Choyke, G. Glenn, J.C. Lyne, W. Rayford, D. Venzon, and W.M. Linehan, Renal cancer in families with hereditary renal cancer: prospective analysis of a tumor size threshold for renal parenchymal sparing surgery. J Urol, 1999. 161(5): p. 1475-9.
93. Jilg, C.A., H.P. Neumann, S. Glasker, O. Schafer, P.U. Ardel, M. Schwardt, and W. Schultze-Seemann, Growth Kinetics in Von Hippel-Lindau-Associated Renal Cell Carcinoma. Urol Int. 2012;88(1):71-8.
94. Walther, M.M., P.L. Choyke, G. Weiss, C. Manolatos, J. Long, R. Reiter, R.B. Alexander, and W.M. Linehan, Parenchymal sparing surgery in patients with hereditary renal cell carcinoma. J Urol, 1995. 153(3 Pt 2): p. 913-6.
95. Jilg, C.A., H.P. Neumann, S. Glasker, O. Schafer, C. Leiber, P.U. Ardel, M. Schwardt, and W. Schultze-Seemann, Nephron sparing surgery in von Hippel-Lindau associated renal cell carcinoma; clinicopathological long-term follow-up. Fam Cancer, 2012.
96. Allasia, M., F. Soria, A. Battaglia, C. Gazzera, M. Calandri, M.P. Caprino, B. Lucatello, A. Veltri, M. Maccario, B. Pasini, A. Bosio, P. Gontero, and P. Destefanis, Radiofrequency Ablation for Renal Cancer in Von Hippel-Lindau Syndrome Patients: A Prospective Cohort Analysis. Clin Genitourin Cancer, 2017.
97. Welsh, S.J. and K. Fife, Pazopanib for the treatment of renal cell carcinoma. Future Oncol, 2015. 11(8): p. 1169-79.

98. Jafri, M. and E.R. Maher, The genetics of phaeochromocytoma: using clinical features to guide genetic testing. *Eur J Endocrinol*, 2012. 166(2): p. 151-8.
99. Neumann, H.P., B. Bausch, S.R. McWhinney, B.U. Bender, O. Gimm, G. Franke, J. Schipper, J. Klisch, C. Altehoefer, K. Zerres, A. Januszewicz, C. Eng, W.M. Smith, R. Munk, T. Manz, S. Glaesker, T.W. Apel, M. Treier, M. Reineke, M.K. Walz, C. Hoang-Vu, M. Brauckhoff, A. Klein-Franke, P. Klose, H. Schmidt, M. Maier-Woelfle, M. Peczkowska, and C. Szmigelski, Germ-line mutations in nonsyndromic pheochromocytoma. *N Engl J Med*, 2002. 346(19): p. 1459-66.
100. Koch, C.A., D. Mauro, M.M. Walther, W.M. Linehan, A.O. Vortmeyer, R. Jaffe, K. Pacak, G.P. Chrourous, Z. Zhuang, and I.A. Lubensky, Pheochromocytoma in von hippel-lindau disease: distinct histopathologic phenotype compared to pheochromocytoma in multiple endocrine neoplasia type 2. *Endocr Pathol*, 2002. 13(1): p. 17-27.
101. Walther, M.M., R. Reiter, H.R. Keiser, P.L. Choyke, D. Venzon, K. Hurley, J.R. Gnarra, J.C. Reynolds, G.M. Glenn, B. Zbar, and W.M. Linehan, Clinical and genetic characterization of pheochromocytoma in von Hippel-Lindau families: comparison with sporadic pheochromocytoma gives insight into natural history of pheochromocytoma. *J Urol*, 1999. 162(3 Pt 1): p. 659-64.
102. Weisbrod, A.B., M. Kitano, K. Gesuwan, C. Millo, P. Herscovitch, N. Nilubol, W.M. Linehan, and E. Kebebew, Clinical utility of functional imaging with 18F-FDOPA in Von Hippel-Lindau syndrome. *J Clin Endocrinol Metab*, 2012. 97(4): p. E613-7.
103. Kudva, Y.C., W.F. Young, G.B. Thompson, and e. al., Adrenal incidentaloma: an important component of the clinical presentation spectrum of benign sporadic adrenal pheochromocytoma. *Endocrinologist*, 1999. 9: p. 77-81.
104. Motta-Ramirez, G.A., E.M. Remer, B.R. Herts, I.S. Gill, and A.H. Hamrahan, Comparison of CT findings in symptomatic and incidentally discovered pheochromocytomas. *AJR Am J Roentgenol*, 2005. 185(3): p. 684-8.
105. Welander, J., P. Soderkvist, and O. Gimm, Genetics and clinical characteristics of hereditary pheochromocytomas and paragangliomas. *Endocr Relat Cancer*, 2011. 18(6): p. R253-76.
106. Papaspyprou, K., T. Mewes, H. Rossmann, C. Fottner, B. Schneider-Raetzke, O. Bartsch, M. Schreckenberger, K.J. Lackner, R.G. Amedee, and W.J. Mann, Head and neck paragangliomas: Report of 175 patients (1989-2010). *Head Neck*, 2012. 34(5): p. 632-7.
107. Gaal, J., F.H. van Nederveen, Z. Erlic, E. Korpershoek, R. Oldenburg, C.C. Boedeker, U. Kontny, H.P. Neumann, W.N. Dinjens, and R.R. de Krijger, Parasympathetic paragangliomas are part of the Von Hippel-Lindau syndrome. *J Clin Endocrinol Metab*, 2009. 94(11): p. 4367-71.
108. Boedeker, C.C., Z. Erlic, S. Richard, U. Kontny, A.P. Gimenez-Roqueplo, A. Cascon, M. Robledo, J.M. de Campos, F.H. van Nederveen, R.R. de Krijger, N. Burnichon, J. Gaal, M.A. Walter, K. Reschke, T. Wiech, J. Weber, K. Ruckauer, P.F. Plouin, V. Darrouzet, S. Giraud, C. Eng, and H.P. Neumann, Head and neck

- paragangliomas in von Hippel-Lindau disease and multiple endocrine neoplasia type 2. *J Clin Endocrinol Metab*, 2009. 94(6): p. 1938-44.
109. Liu, Q., G. Yuan, D. Tong, G. Liu, Y. Yi, J. Zhang, Y. Zhang, L.A. Wang, L. Wang, D. Zhang, R. Chen, Y. Guan, X. Yi, W. Lan, and J. Jiang, Novel genotype-phenotype correlations in five Chinese families with Von Hippel-Lindau disease. *Endocr Connect*, 2018. 7(7): p. 870-878.
110. Lomte, N., S. Kumar, V. Sarathi, R. Pandit, M. Goroshi, S. Jadhav, A.R. Lila, T. Bandgar, and N.S. Shah, Genotype phenotype correlation in Asian Indian von Hippel-Lindau (VHL) syndrome patients with pheochromocytoma/paraganglioma. *Fam Cancer*, 2018. 17(3): p. 441-449.
111. Pandit, R., K. Khadilkar, V. Sarathi, R. Kasaliwal, M. Goroshi, S. Khare, S. Nair, V. Raghavan, A. Dalvi, P. Hira, G. Fernandes, P. Sathe, A. Rojekar, G. Malhotra, G. Bakshi, G. Prakash, A. Bhansali, R. Walia, S. Kamalanathan, J. Sahoo, A. Desai, N. Bhagwat, P. Mappa, R. Rajput, S.R. Chandrashekhar, V. Shivane, P. Menon, A. Lila, T. Bandgar, and N. Shah, Germline mutations and genotype-phenotype correlation in Asian Indian patients with pheochromocytoma and paraganglioma. *Eur J Endocrinol*, 2016. 175(4): p. 311-23.
112. Astapova, O., A. Biswas, A. DiMauro, J. Moalem, and S.R. Hammes, Bilateral Pheochromocytomas in a Patient with Y175C Von Hippel-Lindau Mutation. *Case Rep Endocrinol*, 2018. 2018: p. 8967159.
113. Bhatia, K.S., M.M. Ismail, A. Sahdev, A.G. Rockall, K. Hogarth, A. Canizales, N. Avril, J.P. Monson, A.B. Grossman, and R.H. Reznek, 123I-metaiodobenzylguanidine (MIBG) scintigraphy for the detection of adrenal and extra-adrenal phaeochromocytomas: CT and MRI correlation. *Clin Endocrinol (Oxf)*, 2008. 69(2): p. 181-8.
114. Koch, C.A., A.O. Vortmeyer, R. Diallo, C. Poremba, T.J. Giordano, D. Sanders, S.R. Bornstein, G.P. Chrousos, and K. Pacak, Survivin: a novel neuroendocrine marker for pheochromocytoma. *Eur J Endocrinol*, 2002. 146(3): p. 381-8.
115. Pacak, K., G.P. Chrousos, C.A. Koch, and G. Eisenhofer, Pheochromocytoma: progress in diagnosis, therapy, and genetics., in *Adrenal Disorders*, A. Margioris and G.P. Chrousos, Editors. 2001, Humana Press: New York. p. 379-413.
116. Colvin, A., A.F. Saltzman, J. Walker, J. Bruny, and N.G. Cost, Metastatic Pheochromocytoma in an Asymptomatic 12-Year-Old With von Hippel-Lindau Disease. *Urology*, 2018.
117. Kimura, N., R. Chetty, C. Capella, W.F. Young, C.A. Koch, K.Y. Lam, R.A. DeLellis, A. Kawashima, P. Komminoth, and A. Tischler, Tumours of the Adrenal Medulla and Extraadrenal Paraganglia, in *WHO Classification of Tumours of Endocrine Organs*, R.V. Lloyd, et al., Editors. 2017, WHO Blue Book Series 2017.
118. Kaluarachchi, V.T.S., U. Bulugahapitiya, M. Arambewela, and S. Gunathilake, Successful Management of Pheochromocytoma Detected in Pregnancy by Interval Adrenalectomy in a VHL Patient. *Case Rep Endocrinol*, 2018. 2018: p. 9014585.

119. Eisenhofer, G., J.W. Lenders, H. Timmers, M. Mannelli, S.K. Grebe, L.C. Hofbauer, S.R. Bornstein, O. Tiebel, K. Adams, G. Bratslavsky, W.M. Linehan, and K. Pacak, Measurements of plasma methoxytyramine, normetanephrine, and metanephrine as discriminators of different hereditary forms of pheochromocytoma. *Clin Chem*, 2011. 57(3): p. 411-20.
120. Eisenhofer, G., J.W. Lenders, W.M. Linehan, M.M. Walther, D.S. Goldstein, and H.R. Keiser, Plasma normetanephrine and metanephrine for detecting pheochromocytoma in von Hippel-Lindau disease and multiple endocrine neoplasia type 2. *N Engl J Med*, 1999. 340(24): p. 1872-9.
121. Nielsen, S.M., W.S. Rubinstein, D.L. Thull, M.J. Armstrong, E. Feingold, M.T. Stang, J.R. Gnarr, and S.E. Carty, Genotype-phenotype correlations of pheochromocytoma in two large von Hippel-Lindau (VHL) type 2A kindreds with different missense mutations. *Am J Med Genet A*, 2011. 155A(1): p. 168-73.
122. Launbjerg, K., I. Bache, M. Galanakis, M.L. Bisgaard, and M.L.M. Binderup, von Hippel-Lindau development in children and adolescents. *Am J Med Genet A*, 2017. 173(9): p. 2381-2394.
123. Weise, M., D.P. Merke, K. Pacak, M.M. Walther, and G. Eisenhofer, Utility of plasma free metanephines for detecting childhood pheochromocytoma. *J Clin Endocrinol Metab*, 2002. 87(5): p. 1955-60.
124. Koch, C.A., Should 123I-MIBG scintigraphy be part of the workup for pheochromocytomas? *Nat Clin Pract Endocrinol Metab*, 2009. 5(2): p. 76-7.
125. Ritter, M.M., A. Frilling, P.A. Crossey, W. Hoppner, E.R. Maher, L. Mulligan, B.A. Ponder, and D. Engelhardt, Isolated familial pheochromocytoma as a variant of von Hippel-Lindau disease. *J Clin Endocrinol Metab*, 1996. 81(3): p. 1035-7.
126. Gergics, P., A. Patocs, M. Toth, P. Igaz, N. Szucs, I. Liko, F. Fazakas, I. Szabo, B. Kovacs, E. Glaz, and K. Racz, Germline VHL gene mutations in Hungarian families with von Hippel-Lindau disease and patients with apparently sporadic unilateral pheochromocytomas. *Eur J Endocrinol*, 2009. 161(3): p. 495-502.
127. Iacobone, M., F. Schiavi, M. Bottussi, E. Taschin, S. Bobisse, A. Fassina, G. Opocher, and G. Favia, Is genetic screening indicated in apparently sporadic pheochromocytomas and paragangliomas? *Surgery*, 2011. 150(6): p. 1194-201.
128. Wu, P., N. Zhang, X. Wang, X. Ning, T. Li, D. Bu, and K. Gong, Family history of von Hippel-Lindau disease was uncommon in Chinese patients: suggesting the higher frequency of de novo mutations in VHL gene in these patients. *J Hum Genet*, 2012. 57(4): p. 238-43.
129. Eisenhofer, G., C.D. Vocke, A. Elkahloun, T.T. Huynh, T. Prodanov, J.W. Lenders, H.J. Timmers, J.N. Benhammou, W.M. Linehan, and K. Pacak, Genetic Screening for von Hippel-Lindau Gene Mutations in Non-syndromic Pheochromocytoma: Low Prevalence and False-positives or Misdiagnosis Indicate a Need for Caution. *Horm Metab Res*, 2012. 44(5): p. 343-8.
130. Brunt, L.M., T.C. Lairmore, G.M. Doherty, M.A. Quasebarth, M. DeBenedetti, and J.F. Moley, Adrenalectomy for familial pheochromocytoma in the laparoscopic era. *Ann Surg*, 2002. 235(5): p. 713-20; discussion 720-1.

131. Neumann, H.P., M. Reincke, B.U. Bender, R. Elsner, and G. Janetschek, Preserved adrenocortical function after laparoscopic bilateral adrenal sparing surgery for hereditary pheochromocytoma. *J Clin Endocrinol Metab*, 1999. 84(8): p. 2608-10.
132. Walz, M.K., K. Peitgen, H.P. Neumann, O.E. Janssen, T. Philipp, and K. Mann, Endoscopic treatment of solitary, bilateral, multiple, and recurrent pheochromocytomas and paragangliomas. *World J Surg*, 2002. 26(8): p. 1005-12.
133. Dickson, P.V., C. Jimenez, G.B. Chisholm, D.L. Kennamer, C. Ng, E.G. Grubbs, D.B. Evans, J.E. Lee, and N.D. Perrier, Posterior retroperitoneoscopic adrenalectomy: a contemporary American experience. *J Am Coll Surg*, 2011. 212(4): p. 659-65; discussion 665-7.
134. Castinetti, F., D. Taieb, J.F. Henry, M. Walz, C. Guerin, T. Brue, B. Conte-Devolx, H.P. Neumann, and F. Sebag, MANAGEMENT OF ENDOCRINE DISEASE: Outcome of adrenal sparing surgery in heritable pheochromocytoma. *Eur J Endocrinol*, 2016. 174(1): p. R9-18.
135. Butz, J.J., Q. Yan, T.J. McKenzie, T.N. Weingarten, A.N. Cavalcante, I. Bancos, W.F. Young, Jr., D.R. Schroeder, D.P. Martin, and J. Sprung, Perioperative outcomes of syndromic paraganglioma and pheochromocytoma resection in patients with von Hippel-Lindau disease, multiple endocrine neoplasia type 2, or neurofibromatosis type 1. *Surgery*, 2017. 162(6): p. 1259-1269.
136. Yuan, G., Q. Liu, D. Tong, G. Liu, Y. Yi, J. Zhang, Y. Zhang, L.A. Wang, L. Wang, R. Chen, Y. Guan, X. Yi, W. Lan, and J. Jiang, A retrospective case study of sunitinib treatment in three patients with Von Hippel-Lindau disease. *Cancer Biol Ther*, 2018: p. 1-20.
137. Singer, J., C.A. Koch, W. Kassahun, P. Lamesch, G. Eisenhofer, R. Kluge, T. Lincke, M. Seiwerts, G. Borte, K. Schierle, and R. Paschke, A patient with a large recurrent pheochromocytoma demonstrating the pitfalls of diagnosis. *Nat Rev Endocrinol*, 2011. 7(12): p. 749-55.
138. Ayala-Ramirez, M., L. Feng, M.M. Johnson, S. Ejaz, M.A. Habra, T. Rich, N. Busaidy, G.J. Cote, N. Perrier, A. Phan, S. Patel, S. Waguestock, and C. Jimenez, Clinical risk factors for malignancy and overall survival in patients with pheochromocytomas and sympathetic paragangliomas: primary tumor size and primary tumor location as prognostic indicators. *J Clin Endocrinol Metab*, 2011. 96(3): p. 717-25.
139. Choyke, P.L., G.M. Glenn, J.P. Wagner, I.A. Lubensky, K. Thakore, B. Zbar, W.M. Linehan, and M.M. Walther, Epididymal cystadenomas in von Hippel-Lindau disease. *Urology*, 1997. 49(6): p. 926-31.
140. Glasker, S., M.G. Tran, S.B. Shively, B. Ikejiri, R.R. Lonser, P.H. Maxwell, Z. Zhuang, E.H. Oldfield, and A.O. Vortmeyer, Epididymal cystadenomas and epithelial tumorlets: Effects of VHL deficiency on human epididymis. *J Pathol*, 2006. 210(1): p. 32-41.
141. Mehta, G.U., S.B. Shively, H. Duong, M.G. Tran, T.J. Moncrief, J.H. Smith, J. Li, N.A. Edwards, R.R. Lonser, Z. Zhuang, M.J. Merrill, M. Raffeld, P.H. Maxwell,

- E.H. Oldfield, and A.O. Vortmeyer, Progression of epididymal maldevelopment into hamartoma-like neoplasia in VHL disease. *Neoplasia*, 2008. 10(10): p. 1146-53.
142. Lubensky, I.A., S. Pack, D. Ault, A.O. Vortmeyer, S.K. Libutti, P.L. Choyke, M.M. Walther, W.M. Linehan, and Z. Zhuang, Multiple neuroendocrine tumors of the pancreas in von Hippel-Lindau disease patients: histopathological and molecular genetic analysis. *Am J Pathol*, 1998. 153(1): p. 223-31.
143. Neumann, H.P., E. Dinkel, H. Brambs, B. Wimmer, H. Friedburg, B. Volk, G. Sigmund, P. Riegler, K. Haag, P. Schollmeyer, and et al., Pancreatic lesions in the von Hippel-Lindau syndrome. *Gastroenterology*, 1991. 101(2): p. 465-71.
144. Hough, D.M., D.H. Stephens, C.D. Johnson, and L.A. Binkovitz, Pancreatic lesions in von Hippel-Lindau disease: prevalence, clinical significance, and CT findings. *AJR Am J Roentgenol*, 1994. 162(5): p. 1091-4.
145. Weisbrod, A.B., M. Kitano, F. Thomas, D. Williams, N. Gulati, K. Gesuwan, Y. Liu, D. Venzon, I. Turkbey, P. Choyke, J. Yao, S.K. Libutti, N. Nilubol, W.M. Linehan, and E. Kebebew, Assessment of tumor growth in pancreatic neuroendocrine tumors in von Hippel Lindau syndrome. *J Am Coll Surg*, 2014. 218(2): p. 163-9.
146. Sharma, A., S. Mukewar, and S.S. Vege, Clinical Profile of Pancreatic Cystic Lesions in von Hippel-Lindau Disease: A Series of 48 Patients Seen at a Tertiary Institution. *Pancreas*, 2017. 46(7): p. 948-952.
147. Prasad, V., N. Tiling, T. Denecke, W. Brenner, and U. Plockinger, Potential role of (68)Ga-DOTATOC PET/CT in screening for pancreatic neuroendocrine tumour in patients with von Hippel-Lindau disease. *Eur J Nucl Med Mol Imaging*, 2016. 43(11): p. 2014-20.
148. Krauss, T., A.M. Ferrara, T.P. Links, U. Wellner, I. Bancos, A. Kvachenyuk, K. Villar Gomez de Las Heras, M.Y. Yukina, R. Petrov, G. Bullivant, L. von Duecker, S. Jadhav, U. Ploeckinger, S. Welin, C. Schalin-Jantti, O. Gimm, M. Pfeifer, J. Ngeow, K. Hasse-Lazar, G. Sanso, X. Qi, M.U. Ugurlu, R.E. Diaz, N. Wohllk, M. Peczkowska, J. Aberle, D.M. Lourenco, Jr., M.A.A. Pereira, M. Fragozo, A.O. Hoff, M.Q. Almeida, A.H.D. Violante, A.R.P. Quidute, Z. Zhang, M. Recasens, L.R. Diaz, T. Kunavasarat, T. Wannachalee, S. Sirinvaravong, E. Jonasch, S. Grozinsky-Glasberg, M. Fraenkel, D. Beltsevich, V.I. Egorov, D. Bausch, M. Schott, N. Tiling, G. Pennelli, S. Zschiedrich, R. Darr, J. Ruf, T. Denecke, K.H. Link, S. Zovato, E. von Dobschuetz, S. Yaremchuk, H. Amthauer, O. Makay, A. Patocs, M.K. Walz, T.B. Huber, J. Seufert, P. Hellman, R.H. Kim, E. Kuchinskaya, F. Schiavi, A. Malinoc, N. Reisch, B. Jarzab, M. Barontini, A. Januszewicz, N. Shah, W.F. Young, Jr., G. Opocher, C. Eng, H.P.H. Neumann, and B. Bausch, Preventive medicine of von Hippel-Lindau disease-associated pancreatic neuroendocrine tumors. *Endocr Relat Cancer*, 2018. 25(9): p. 783-793.
149. de Mestier, L., S. Gaujoux, J. Cros, O. Hentic, M.P. Vullierme, A. Couvelard, G. Cadiot, A. Sauvanet, P. Ruszniewski, S. Richard, and P. Hammel, Long-term Prognosis of Resected Pancreatic Neuroendocrine Tumors in von Hippel-Lindau

- Disease Is Favorable and Not Influenced by Small Tumors Left in Place. *Ann Surg.* 2015 Aug;262(2):384-8
150. Libutti, S.K., P.L. Choyke, D.L. Bartlett, H. Vargas, M. Walther, I. Lubensky, G. Glenn, W.M. Linehan, and H.R. Alexander, Pancreatic neuroendocrine tumors associated with von Hippel Lindau disease: diagnostic and management recommendations. *Surgery*, 1998. 124(6): p. 1153-9.
151. Libutti, S.K., P.L. Choyke, H.R. Alexander, G. Glenn, D.L. Bartlett, B. Zbar, I. Lubensky, S.A. McKee, E.R. Maher, W.M. Linehan, and M.M. Walther, Clinical and genetic analysis of patients with pancreatic neuroendocrine tumors associated with von Hippel-Lindau disease. *Surgery*, 2000. 128(6): p. 1022-7;discussion 1027-8.
152. von Ducker, L., M.K. Walz, C. Voss, G. Arnold, C. Eng, and H.P. Neumann, Laparoscopic organ-sparing resection of von Hippel-Lindau disease-associated pancreatic neuroendocrine tumors. *World J Surg*, 2011. 35(3): p. 563-7.
153. O'Toole, S.M. and W.M. Drake, Response to Somatostatin Analog Therapy in a Patient With von Hippel-Lindau Disease and Multiple Pancreatic Neuroendocrine Tumors. *Pancreas*, 2017. 46(7): p. e57.
154. Glasker, S., R.R. Lonser, M.G. Tran, B. Ikejiri, J.A. Butman, W. Zeng, P.H. Maxwell, Z. Zhuang, E.H. Oldfield, and A.O. Vortmeyer, Effects of VHL deficiency on endolymphatic duct and sac. *Cancer Res*, 2005. 65(23): p. 10847-53.
155. Lonser, R.R., M. Baggenstos, H.J. Kim, J.A. Butman, and A.O. Vortmeyer, The vestibular aqueduct: site of origin of endolymphatic sac tumors. *J Neurosurg*, 2008. 108(4): p. 751-6.
156. Manski, T.J., D.K. Heffner, G.M. Glenn, N.J. Patronas, A.T. Pikus, D. Katz, R. Lebovics, K. Sledjeski, P.L. Choyke, B. Zbar, W.M. Linehan, and E.H. Oldfield, Endolymphatic sac tumors. A source of morbid hearing loss in von Hippel-Lindau disease. *JAMA*, 1997. 277(18): p. 1461-6.
157. Butman, J.A., H.J. Kim, M. Baggenstos, J.M. Ammerman, J. Dambrosia, A. Patsalides, N.J. Patronas, E.H. Oldfield, and R.R. Lonser, Mechanisms of morbid hearing loss associated with tumors of the endolymphatic sac in von Hippel-Lindau disease. *JAMA*, 2007. 298(1): p. 41-8.
158. Singh, A.D., C.L. Shields, and J.A. Shields, von Hippel-Lindau disease. [Review] [205 refs]. *Survey of Ophthalmology*, 2001. 46(2): p. 117-42.
159. Wilding, A., S.L. Ingham, F. Laloo, T. Clancy, S.M. Huson, A. Moran, and D.G. Evans, Life expectancy in hereditary cancer predisposing diseases: an observational study. *J Med Genet*, 2012. 49(4): p. 264-9.
160. Glasker, S., B.U. Bender, T.W. Apel, E. Natt, V. van Velthoven, R. Scheremet, J. Zentner, and H.P. Neumann, The impact of molecular genetic analysis of the VHL gene in patients with haemangioblastomas of the central nervous system. *J Neurol Neurosurg Psychiatry*, 1999. 67(6): p. 758-62.
161. Woodward, E.R., K. Wall, J. Forsyth, F. Macdonald, and E.R. Maher, VHL mutation analysis in patients with isolated central nervous system haemangioblastoma. *Brain*, 2007. 130(Pt 3): p. 836-42.
162. Maher, E.R., Von Hippel-Lindau disease. *Curr Mol Med*, 2004. 4(8): p. 833-42.

163. Seizinger, B.R., G.A. Rouleau, L.J. Ozelius, A.H. Lane, G.E. Farmer, J.M. Lamiell, J. Haines, J.W. Yuen, D. Collins, D. Majoor-Krakauer, and et al., Von Hippel-Lindau disease maps to the region of chromosome 3 associated with renal cell carcinoma. *Nature*, 1988. 332(6161): p. 268-9.
164. Latif, F., K. Tory, J. Gnarra, M. Yao, F.M. Duh, M.L. Orcutt, T. Stackhouse, I. Kuzmin, W. Modi, L. Geil, B. Zbar, and M. Lerman, Identification of the von Hippel-Lindau disease tumor suppressor gene. *Science*, 1993. 260(5112): p. 1317-20.
165. Kaelin, W.G., Jr., Molecular basis of the VHL hereditary cancer syndrome. *Nature Reviews. Cancer.*, 2002. 2(9): p. 673-82.
166. Pause, A., S. Lee, R.A. Worrell, D.Y. Chen, W.H. Burgess, W.M. Linehan, and R.D. Klausner, The von Hippel-Lindau tumor-suppressor gene product forms a stable complex with human CUL-2, a member of the Cdc53 family of proteins. *Proc Natl Acad Sci U S A*, 1997. 94(6): p. 2156-61.
167. Lonergan, K.M., O. Iliopoulos, M. Ohh, T. Kamura, R.C. Conaway, J.W. Conaway, and W.G. Kaelin, Jr., Regulation of hypoxia-inducible mRNAs by the von Hippel-Lindau tumor suppressor protein requires binding to complexes containing elongins B/C and Cul2. *Mol Cell Biol*, 1998. 18(2): p. 732-41.
168. Lisztwan, J., G. Imbert, C. Wirbelauer, M. Gstaiger, and W. Krek, The von Hippel-Lindau tumor suppressor protein is a component of an E3 ubiquitin-protein ligase activity. *Genes Dev*, 1999. 13(14): p. 1822-33.
169. Semenza, G.L., Targeting HIF-1 for cancer therapy. *Nat Rev Cancer*, 2003. 3(10): p. 721-32.
170. Iliopoulos, O., A.P. Levy, C. Jiang, W.G. Kaelin, Jr., and M.A. Goldberg, Negative regulation of hypoxia-inducible genes by the von Hippel-Lindau protein. *Proc Natl Acad Sci U S A*, 1996. 93(20): p. 10595-9.
171. Maxwell, P.H., M.S. Wiesener, G.W. Chang, S.C. Clifford, E.C. Vaux, M.E. Cockman, C.C. Wykoff, C.W. Pugh, E.R. Maher, and P.J. Ratcliffe, The tumour suppressor protein VHL targets hypoxia-inducible factors for oxygen-dependent proteolysis. *Nature*, 1999. 399(6733): p. 271-5.
172. Krieg, M., R. Haas, H. Brauch, T. Acker, I. Flamme, and K.H. Plate, Up-regulation of hypoxia-inducible factors HIF-1alpha and HIF-2alpha under normoxic conditions in renal carcinoma cells by von Hippel-Lindau tumor suppressor gene loss of function. *Oncogene*, 2000. 19(48): p. 5435-43.
173. Frew, I.J. and W. Krek, Multitasking by pVHL in tumour suppression. *Curr Opin Cell Biol*, 2007. 19(6): p. 685-90.
174. Young, A.P., S. Schlisio, Y.A. Minamishima, Q. Zhang, L. Li, C. Grisanzio, S. Signoretti, and W.G. Kaelin, Jr., VHL loss actuates a HIF-independent senescence programme mediated by Rb and p400. *Nat Cell Biol*, 2008. 10(3): p. 361-9.
175. Knudson, A.G., Hereditary cancer: two hits revisited. *J Cancer Res Clin Oncol*, 1996. 122(3): p. 135-40.
176. Mandriota, S.J., K.J. Turner, D.R. Davies, P.G. Murray, N.V. Morgan, H.M. Sowter, C.C. Wykoff, E.R. Maher, A.L. Harris, P.J. Ratcliffe, and P.H. Maxwell,

- HIF activation identifies early lesions in VHL kidneys: evidence for site-specific tumor suppressor function in the nephron. *Cancer Cell*, 2002. 1(5): p. 459-68.
177. Ding, Z., P. German, S. Bai, Z. Feng, M. Gao, W. Si, M.M. Sobieski, C.C. Stephan, G.B. Mills, and E. Jonasch, Agents That Stabilize Mutated von Hippel-Lindau (VHL) Protein: Results of a High-Throughput Screen to Identify Compounds That Modulate VHL Proteostasis. *J Biomol Screen*. 2012 Jun;17(5):572-80